Arsenic uptake by aquatic macrophyte Spirodela polyrhiza L.: Interactions with phosphate and iron

Arsenic Uptake by Aquatic Macrophyte *Spirodela polyrhiza* L.: Interactions with Phosphate and Iron

M. Azizur Rahman[1]; H. Hasegawa[*, 1]; K. Ueda[1]; T. Maki[1]; M. Mahfuzur Rahman[2]

[1]Graduate School of Natural Science & Technology, Kanazawa University, Kakuma, Kanazawa 920-1192, Japan; [2]Department of Botany, Faculty of Biological Sciences, Jahangirnagar University, Savar, Dhaka-1342, Bangladesh.

*Corresponding author

E-mail: hhiroshi@t.kanazawa-u.ac.jp

Tel/Fax: 81-76-234-4792

Abstract

The uptake of arsenate (As(V)) and dimethylarsinic acid (DMAA) by aquatic macrophyte *Spirodela polyrhiza* L. was investigated to determine the influence of arsenic interaction with PO_4^{3-} and Fe ions. Plants were grown hydroponically on standard Murashige and Skoog (MS) culture solutions. Arsenic concentrations in Fe-oxide (Fe-plaque) on plant surfaces were determined by citrate-bicarbonate-ethylenediaminetetraacetic acid (CBE) technique. *Spirodela polyrhiza* L. accumulated 51-fold arsenic from arsenate solution compared to that from DMAA solution with initial concentrations of 4.0 and 0.02 μM of arsenic and phosphate, respectively. The arsenate uptake was negatively ($p < 0.001$) correlated with phosphate uptake and positively ($p < 0.05$) correlated with iron uptake. About 56% of the total arsenic was accumulated into the plant tissues while 44% was adsorbed on Fe plaque (CBE-extract), when the plants were grown on arsenate solution. The DMAA uptake into the plant was neither affected by the phosphate concentrations nor correlated ($p > 0.05$) with iron accumulation. The results suggest that adsorption of arsenate on Fe plaque of the surface of *Spirodela polyrhiza* L. contributes to the arsenic uptake significantly. Thus, arsenate uptake in *Spirodela polyrhiza* L. occurred through the phosphate uptake pathway and by physico-chemical adsorption on Fe-plaques of plant surfaces as well. The *Spirodela polyrhiza* L. uses different mechanisms for DMAA uptake.

Keywords: Arsenate, DMAA, Uptake, Interactions, Physico-chemical adsorption, Fe-plaque, *Spirodela polyrhiza* L.

1. Introduction

Arsenic is an important environmental and health concern due to its known chronic and epidemic toxicity. The main arsenic exposures to humans are through water pathway and food contamination, for instance in Bangladesh [1-3] and West Bengal, India [4] where most of the contaminations originate from natural release from rocks in the aquifer. Geogenic arsenic contamination from aquifer rocks has also been reported in Thailand [5], Vietnam, Inner Mongolia, Greece, Hungary, U.S.A., Ghana, Chile, Argentina and Mexico [6, 7]. Unfortunately, the traditional chemical and physical remediation techniques are limited due to the pattern of discharge. Hence, Phytoremediation, a plant-based green technology, is proposed as a viable alternative. Its relative inexpensiveness and eco-friendliness have made it an attractive method for water and soil remediation [8]. Some terrestrial plant species such as *Agrostis castellana*; *Agrostis delicatula* [9], *Bidens cynapiifolia* [10], Chinese brake fern (*Pteris vittata* L.) [11] and silver fern (*Pityrogramma calomelanos* L.) [12] have been reported to accumulate significant fractions of arsenic from soil. In particular, Chinese brake fern accumulates a formidable quantity of arsenic from soil [12, 13] and stores in the fronds [12, 14]. The arsenic hyperaccumulating terrestrial plants are considered for soil remediation. However, restoration of contaminated waters of ponds, lacks, ditches as well as irrigation water remains unresolved. Aquatic macrophytes could be a good tool for the environmentally sound and effective remediation of arsenic contaminated waters [15, 16]. Hence, we investigated the possible use of duckweed in aquatic phytoremediation.

In the present study, duckweed (*Spirodela polyrhiza* L.) was selected because of its fast growth, wide distribution, short life span and stability to the large scale environmental changes [17, 18]. The plant commonly grows in inland small water bodies such as ponds,

lacks, ditches in Bangladesh and West Bengal, India into which arsenic contaminated water from hand tube wells (used for household necessity) and shallow tube wells (used for irrigation) is drained. Moreover, duckweed (*Spirodela polyrhiza* L.) grows in the rice fields of south Asian countries where arsenic contaminated groundwater is the main source of irrigation during dry season. The plant is also beneficial to rice cultivation as it suppressed or reduce weed growth in the rice field.

Arsenate and arsenite are bioavailable inorganic forms of arsenic in aquatic systems [19]. The dynamics of arsenate exchange between water and adsorbing colloids are analogous to those of phosphate, though the competition for exchange sites favors phosphate over arsenate [20]. Arsenate and DMAA are the major species of arsenic in oxic aquatic systems [21]. Uptake behavior of these two arsenic species could reflect the influence of inorganic and organic arsenic species and their interactions with PO_4^{3-} and Fe ions. The comparison between inorganic (arsenate) and organic (DMAA) arsenic species uptake is important because of their limit of toxicity too.

In nature, wetland plants form dense root networks in upper wetland sediments and, under flooded conditions, pump oxygen to their roots for respiration [22]. Thus, oxygenation of the rhizosphere by wetland plants leads to precipitation of iron (oxyhydro)-oxides in the rhizosphere and on the roots of plants [23]. Precipitation of iron (oxyhydro)-oxides on roots of aquatic plants has also been reported in literatures [24]. Due to the high adsorptive affinity of arsenic for iron hydroxides, Fe plaque formation on root surface of aquatic plants might be significant in the uptake of arsenic by the plants. In the present study we reported the uptake of arsenate and DMAA in duckweed (*Spirodela polyrhiza* L.) and their interactions with PO_4^{3-} and Fe ions. The contribution of Fe-plaque formation on plant's surfaces in the arsenic uptake has also been discussed.

2. Materials and Methods

2.1. Conditions for plant cultivation

The *Spirodela polyrhiza* L., collected from a rice field in Manikgonj of Dhaka, Bangladesh, was stock-cultured in green house for 2 weeks. Then, the plants were rinsed three times with deionized (DI) water and transferred to growth chamber. In the growth chamber, the experiment was conducted with the conditions being set as 14:10 h light/dark schedule, 100-125 μE m^{-2} s^{-1} light intensity, 75% humidity, 22 ºC and 20(±2) ºC temperatures for day and night, respectively.

Modified standard Murashige and Skoog (MS) culture solution was used as growth medium in the experiment (Table 1). The control culture solution contained 0.02 μM PO_4^{3-} and other culture solutions were prepared by modifying the PO_4^{3-} concentration to 100 or 500 μM. Three test concentrations (1.0, 2.0 and 4.0 μM) of either arsenate or DMAA were added to the modified MS culture solutions. The pH of the solution was adjusted to 6.0.

Before inoculation, *Spirodela polyrhiza* L. from the stock-culture were rinsed for three times with deionized (DI) water. About 100 ml of culture solution was taken into 200-ml polystylene test vessels (118 x 86 x 60 mm). About 120 individual plants were inoculated in each of the test vessels. The experiment was arranged following the randomized design (RD) with three replicates. Stock solutions of arsenate and DMAA were made by dissolving $Na_2HAsO_4{\cdot}7H_2O$ and $(CH_3)_2AsO_2Na{\cdot}3H_2O$ in DI water, respectively. Arsenic stock solutions were added to the cultures before inoculation. The plants were grown for 12 days. Changes in the volume of cultures from evaporation and accumulation were compensated by adding DI water every 2 days throughout the experiment.

2.2. Iron plaque induction

A separate experiment was conducted to investigate the role of iron plaque on arsenic uptake in *Spirodela polyrhiza* L. Plants were grown in 1.5 L of DI water for 24 h before iron induction to minimize interferences from other elements with iron. They were then, transferred into 1 L of the MS solution containing 0.36 mM of iron as $FeSO_4 \cdot 7H_2O$ and grown for 2 days. The pH of solution was adjusted to 6.0 using either 0.1 M KOH or 0.1 M HCl. The specified standard concentration of phosphate for MS culture solution was not modified. After 2 days in high iron medium, plants were inoculated into MS culture solution for 12 days as described in the previous section, with 6.0 µM of either arsenate or DMAA.

2.3. CBE-extraction of Fe-plaques

Iron plaques from plant surfaces were extracted using citrate-bicarbonate-ethylenediaminetetraacetate (CBE)-technique, a modification of dithionite-citrate-bicarbonate (DCB)-extraction method of Taylor and Crowder [25] and Otte et al. [26]. The CBE solution was prepared from 0.03, 0.125 and 0.050 M of sodium citrate, sodium bicarbonate and EDTA, respectively. Plants were treated with 30 ml of CBE solution for 60 min. at room temperature. The plants were then, rinsed with DI water for 3 times, and the rinsed water was added to the CBE-extracts to make a total volume of 50 ml.

2.4. Sample preparation and chemical analysis

All plants were harvested after 12 days of inoculation. After rinsing with DI water for four times, the plant samples were kept on clean absorbent paper to remove the water from the plant surfaces. The samples were dried at 65 ºC until they reached a constant weight. Then, 0.10-0.20 g of dried samples was taken into 50-ml polyethylene tubes (*Digi*Tubes, SCP Science, Canada) for digestion. Five ml of 65% HNO_3 were added to the sample and then,

left to incubate for 12 hours. The samples were heated on a heating block (*Digi*PREP, SCP Science, Canada) at 95 ºC for 2 hours. After cooling to room temperature, 3 ml of 30% hydrogen peroxide were added and the samples were heated again at 105 ºC for 20 min. Then, the digests were diluted to 10 ml with DI water and taken into 15-ml polyethylene bottles (HDPE, NALGENE®, Nalge Nunc International, Rochester, NY) in readiness for analysis.

Arsenic and iron were analyzed using graphite-furnace atomic absorption spectrometer (GF-AAS, Z-8100, Hitachi, Japan). For the determination of arsenic, 5 µL of 0.05 M nickel nitrate was added to a 10-µL sample into the cuvette as matrix modifier. Certified standard reference material 1573a (tomato leaf from NIST, USA) was used to check the accuracy of analysis. Arsenic concentration in certified reference material was 0.112±0.004 µg g^{-1} while the measured arsenic concentration was 0.123±0.009 µg g^{-1}. The concentrations detected in all samples were above the instrumental limits of detection (≥ 0.01 µM in samples in water). Total phosphate was determined spectrophotometrically [27].

All chemical reagents used in this experiment were of analytical grade. Glassware and dishes were washed with detergent solution, 3 M HCl and finally rinsed with DI water for eight times before use. In each analytical batch, at least two reagent blanks and three replicate samples were included.

2.5. Data analysis

Bioaccumulation of arsenic by *Spirodela polyrhiza* L. was determined on dry weight basis [18]. The experimental data were statistically analyzed for mean separation of different arsenic treatments according to the least significant difference (LSD) at 5% level by IRRI-

181 STAT 4.0 for windows (Developed by the Biometrics unit, IRRI, Philippines) and the
182 Pearson correlation coefficient (r) was calculated by SPSS® statistical package.
183
184 ## 3. Results and Discussion
185 ### 3.1. Accumulation of As species in *S. polyrhiza* L.
186 The accumulation of arsenic in *Spirodela polyrhiza* L. from arsenate treatment is presented
187 in Fig. 1., where as the accumulation from DMAA treatment is presented in Fig. 2. The
188 results show that *Spirodela polyrhiza* L. accumulated about 51-fold arsenic, when the
189 plants were inoculated in arsenate solution compared to that in DMAA solution. Arsenic
190 contents in tissues had a strong positive correlation with the initial concentrations of
191 arsenate in culture solutions ($r = 0.979$; $p < 0.001$ at 95% confidence interval).
192
193 ### 3.2. Influence PO_4^{3-} on As uptake
194 The accumulation of arsenic in *Spirodela polyrhiza* L. decreased significantly with the
195 increase of the phosphate concentration in the culture solutions for all three arsenate
196 concentrations (Fig. 1). When the concentration of PO_4^{3-} in the culture solution was
197 increased from 0.02 to 500 µM with a constant arsenate concentration (4.0 µM), arsenic
198 accumulation into the *Spirodela polyrhiza* L. decreased by 68%. The result implies the
199 suppression of arsenic uptake in *Spirodela polyrhiza* L. by phosphate from arsenate
200 solution.
201
202 Mkandawire and Dudel [15] reported 0.26 and 1.45 µmol g^{-1} dry weight of arsenic
203 accumulation in fronds of *Lemna gibba* L. (lesser duckweed), when the PO_4^{3-}
204 concentrations in arsenate treated culture solution were 421 and 0.014 µM, respectively. In
205 another study, Mkandawire et al. [18] observed that arsenic accumulation decreased by
206 28-32%, when PO_4^{3-} concentration in arsenate treated culture solution was increased from

207 0.014 to 421 μM. The impact of increasing phosphate concentration in culture solutions
208 was similar to that of present experiment. Thus, the magnitude of arsenic accumulation in
209 *Spirodela polyrhiza* L. in relation to PO_4^{3-} concentrations in culture solution with arsenate
210 is comparable with that in *Lemna gibba* L. This might be because AsO_4^{3-} is a sorption
211 analog of PO_4^{3-} and competes with it for uptake carriers in the plasmalemma [18].
212 Mkandawire and Dudel [15] proposed the arsenate uptake in *Lemna gibba* L. might occur
213 through the phosphate uptake pathway due to similar chemical behavior of AsO_4^{3-} and
214 PO_4^{3-}. The present findings suggest the same for *Spirodela polyrhiza* L.
215
216 In contrast, arsenic accumulation was not affected with the increase of phosphate
217 concentration in DMAA solution (Fig. 2). The results imply that the arsenate uptake into
218 the aquatic macrophyte is related to the phosphate concentration in the culture solution,
219 while DMAA uptake was not.
220
221 **3.3. Effect of As species on PO_4^{3-} uptake**
222 Phosphorus uptake in *Spirodela polyrhiza* L. decreased significantly ($p < 0.001$) with the
223 increase of arsenate concentrations in culture solutions, while DMAA had no significant
224 effect ($p > 0.05$) on its uptake. Pearson correlation analysis revealed a strong negative
225 relationship between the arsenate concentration in culture solutions and phosphate
226 concentration in plant tissues (r = -0.994; $p < 0.001$ at 95% confidence interval). On the
227 other hand, the correlation was not significant (r = -0.220; $p > 0.05$ at 95% confidence
228 interval) for DMAA. De La Rosa et al. [28] reported the reduction of phosphate uptake
229 into tumbleweed (*Salsola kali*), when the plant was exposed to arsenate.
230
231 Figure 3 shows the relationship between arsenic and phosphate concentrations in
232 *Spirodela polyrhiza* L. The correlation between arsenic and phosphate concentrations (r =

233 -0.982; $p < 0.001$ at 95% confidence interval) in *Spirodela polyrhiza* L. was stronger and
234 negative, when the plants were exposed to arsenate solution (Fig. 3a). On the other hand,
235 the correlation was very poor (r = -0.281; $p > 0.05$ at 95% confidence interval), when the
236 plants were exposed to DMAA solution (Fig. 3b). The results suggest that the phosphate
237 uptake into the aquatic macrophyte might be inhibited by arsenate while its uptake was not
238 influenced by DMAA. The reduction of phosphate uptake might be due to the desorption
239 of arsenate from iron plaque of plant surfaces. Barrow (29) investigated As(V) and P
240 competitive adsorption in soil and found that, though As(V) desorbed some previously
241 adsorbed P, a substantial portion of the bound P was not displaced by As(V).
242
243 **3.4. Influence of Fe on As species uptake**
244 Iron concentrations were positively correlated with those of arsenic (r = 0.662; $p = 0.019$
245 at 95% confidence interval) in *Spirodela polyrhiza* L. exposed to arsenate solution. On the
246 other hand, iron concentrations did not correlate with those of arsenic (r = 0.031; $p = 0.923$
247 at 95% confidence interval) in plants exposed to DMAA solution. Robinson et al. [30] also
248 reported positive correlation between arsenic and iron concentrations in aquatic plants
249 because arsenic could be adsorbed by iron oxides on plant surfaces. However, which
250 species of arsenic predominated in such adsorption was not clear from their study. The
251 present study suggest that inorganic arsenic species are more likely to be adsorbed on Fe
252 plaques on *Spirodela polyrhiza* L. Blute et al. [31] reported that arsenate correlated
253 positively with iron in plaque and negatively with iron adsorbed on the roots of *Typha*
254 *latifolia* (cattail) growing on arsenic contaminated wetland sediments. According to Blute
255 et al. [31], the ferric plaque was predominantly Fe(III) oxyhydroxide, and arsenate
256 accounted for 80% of the total adsorbed arsenic. Adsorption of arsenic on ferric iron
257 inhibited the mobility of arsenic into the roots. Another report [32] suggested the same
258 mechanism for arsenic retention by rice root.

3.5. Influence of PO_4^{3-} on As adsorption on Fe plaque of plant surfaces

Arsenic and iron concentrations in plants grown in solution with arsenate and lower phosphate were highly correlated (r = 0.994; $p < 0.001$ at 95% confidence interval) (Fig. 4a). But they were not significantly correlated when the plants were grown in solution with higher phosphate (r = -0.220 and -0.461 for 100 and 500 µM of PO_4^{3-} in solutions, respectively; $p > 0.05$) and the same arsenic species (Fig. 4b, 4c). This might attribute to the adsorption of arsenate on iron plaques of plant surfaces in lower phosphate solution, which was desorbed by phosphate in higher phosphate solution.

The adsorption of phosphate on iron plaque has been reported by Zhang et al. [33]. They demonstrated that the amounts of phosphorus accumulated in iron plaque were correlated positively to the amount of iron plaque on roots. Therefore, iron plaque on roots might act as a phosphorus pool. Beside this, there are contradictory reports on the effects of iron plaque on phosphorus uptake by plant [26, 34, 35]. The reasons for such opposite results that iron plaque affect phosphorus uptake may be due to the different plant species and the amount of iron plaque, especially to the latter. Zhang et al. [33] reported that the phosphorus concentration in shoots of rice increased by 72% with the increase of iron plaque from 0.22 to 24.5 g $^{-kg}$ dry root weight. But higher plaque deposition (28.3 g $^{-kg}$ dry root weight) on rice root surface decreased phosphate concentration.

Though Zhang et al. [33] demonstrated the adsorption of phosphate on Fe plaques of plant's root surface the role of phosphate is not clear from their study. The present study suggests that arsenate adsorbed on iron plaques of plant surfaces might be desorbed by phosphate at higher concentration.

285 **3.6. Comparison between internalized and surface adsorbed As**

286 Physico-chemical adsorption, a different mechanism for arsenic accumulation into aquatic
287 plants, has been proposed in the literature (Robinson et al. [30]. In this mechanism,
288 suspended oxides of iron (Fe plaques) on the root and lower surface of the fronds of
289 aquatic plants adsorb arsenic.

290

291 To understand the arsenate adsorption on iron plaques, iron plaques were induced on
292 *Spirodela polyrhiza* L. surfaces before expose them to the arsenic species. Arsenic
293 concentrations in plant tissues and iron plaques (CBE-extracts) were determined
294 separately. Results showed that when *Spirodela polyrhiza* L. was exposed to 6.0 µM
295 arsenate, 0.86±0.06 µmol g^{-1} dry weight of arsenic was adsorbed on iron plaques of plant
296 surfaces. On the other hand, arsenic concentration was 1.08±0.12 µmol g^{-1} dry weight into
297 the plant tissues (Table 2). The result shows that about 56% of the total arsenic is
298 distributed into the plant tissues compared to 44% in Fe-plaques. However, significantly
299 higher concentration of iron (547±5 µM g^{-1} dry weight) in CBE-extracts compared with
300 plant tissues (69.3±1.0 µM g^{-1} dry weight) (Table 2) confirms the formation of iron
301 plaques on plant surfaces. The current results imply that adsorption of arsenate on Fe
302 plaque of the surface of *Spirodela polyrhiza* L. contributes to arsenate uptake significantly.

303

304 There was no significant correlation between DMAA and phosphate concentrations in
305 *Spirodela polyrhiza* L. (Fig. 3b). Moreover, DMAA and iron concentrations in plants did
306 not correlate significantly ($p > 0.05$) in neither low nor high phosphate solutions (Fig. 4A,
307 4B and 4C). It suggests that the accumulation of DMAA might not correlate with
308 phosphate accumulation. Arsenic concentrations in Fe-plaques and plant tissues were low
309 and did not differ significantly, when the plants were exposed to DMAA (Table 2). The

results imply that DMAA less adsorbed to Fe-plaques on the plant surface and Fe has more effect on As uptake from inorganic arsenic sources.

4. Conclusion:

The results of the present study show that not only internalized, but also surface adsorbed arsenic (mostly arsenate) contributes significantly to the total amount of arsenic uptake in aquatic macrophyte *Spirodela polyrhiza* L. Thus, it could be suggest that arsenic uptake in *Spirodela polyrhiza* L. occurred through the phosphate uptake pathway as well as by physico-chemical adsorption on Fe-plaques of plant's surfaces. The arsenate uptake in the plant is related to the Fe ion and phosphate concentrations in culture medium while DMAA was not. It is well reported in many previous studies that arsenate compete with phosphate for uptake carriers in the plasmalemma, which is also consistent to the present study. But the current study reports that higher phosphate concentration in the culture medium might desorbs arsenate from iron plaques of plant surfaces.

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436 **Table 1:** Modified[a] murashige & skoog (MS) nutrients for *Spirodela polyrhiza* L.
437 hydroponic culture medium
438

Nutrients	Concentration (mg l^{-1})
KNO_3	1900
NH_4NO_3	1650
$CaCl_2.2H_2O$	440
$MgSO_4.7H_2O$	370
K_2HPO_4	Modified [a]
$FeSO_4.7H_2O$	27.80
$MnSO_4.5H_2O$	22.30
$ZnSO_4.7H_2O$	8.60
H_3BO_3	6.20
KI	0.83
$Na_2MoO_4.2H_2O$	0.25
$CuSO_4.5H_2O$	0.025
$CoCl_2.6H_2O$	0.025
Na_2-EDTA	37.30

439

440 [a] The control solution contained 0.02 µM PO_4^{3-} and the modifications of the
441 solutions were 100 and 500 µM of PO_4^{3-}. The pH of the solution was adjusted to
442 6.0.
443
444
445
446

Table-2: Arsenic and iron concentrations into the tissues of *Spirodela polyrhiza* L. and Fe-plaques of the plant surfaces grown for 12 days in solution containing 6.0 µM arsenic [a]

As treatments in solutions	µmol As (g dry weight)$^{-1}$		µmol Fe (g dry weight)$^{-1}$	
	Plant tissues	CBE-extracts	Plant tissues	CBE-extracts
Control	0.04±0.01c	0.02±0.00c	65.2±0.2a	914±3a
Arsenate	1.08±0.12a	0.86±0.06a	69.3±1.0a	547±5b
DMAA	0.05±0.02b	0.08±0.03b	50.2±0.5b	484±5c

[a] Different letters indicate significant differences ($p < 0.05$) between treatments according to the least significant difference (LSD).

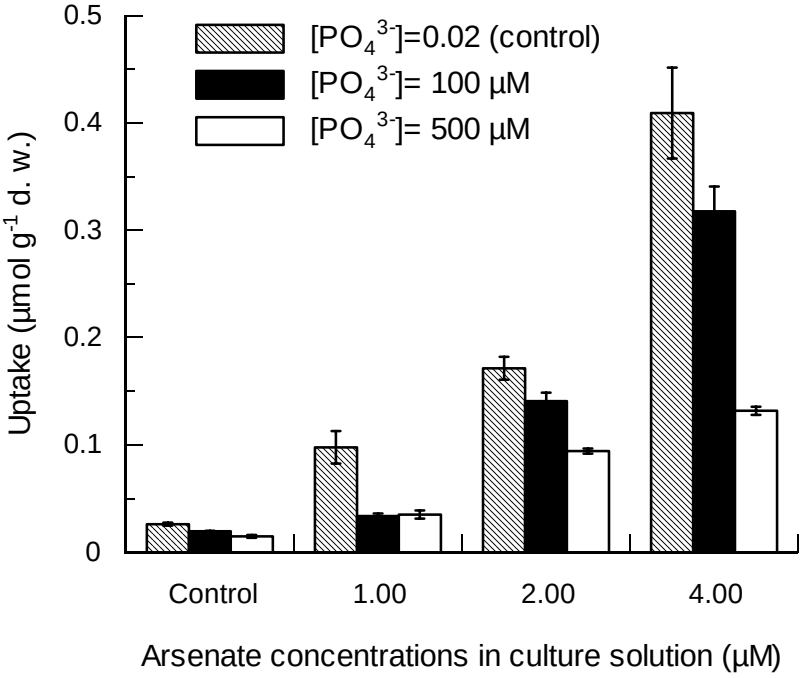


Figure 1: Arsenate uptake in *S. polyrhiza* L. affected by the PO_4^{3-} concentrations in culture solution. Each point is the average of three replicates. Error bars represent ± SD (*n*=3).

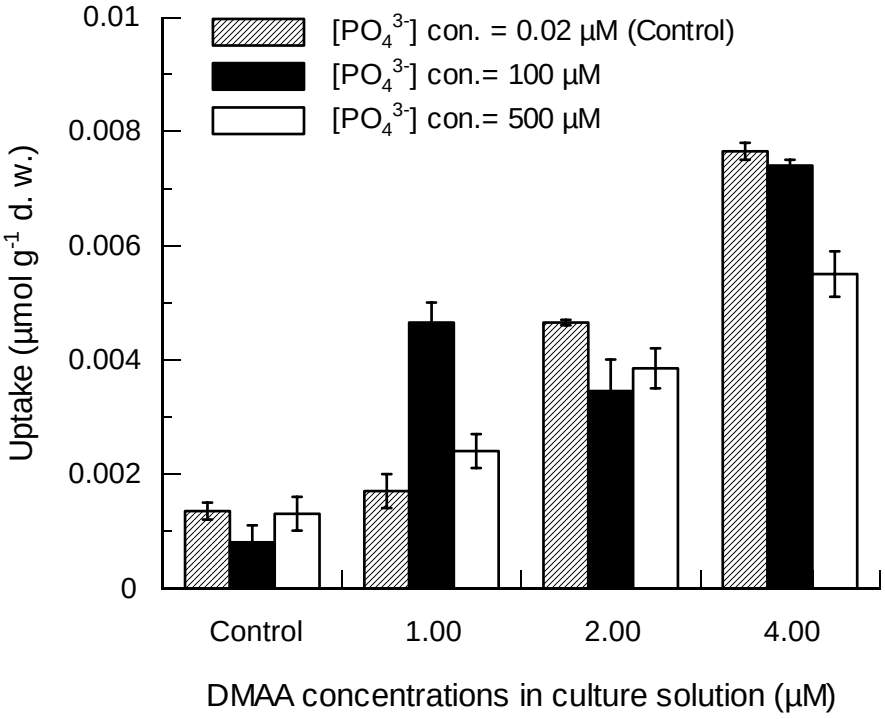


Figure 2: DMAA uptake in *S. polyrhiza* L. affected by the PO_4^{3-} concentrations in culture solution. Each point is the average of three replicates. Error bars represent ± SD (*n*=3).

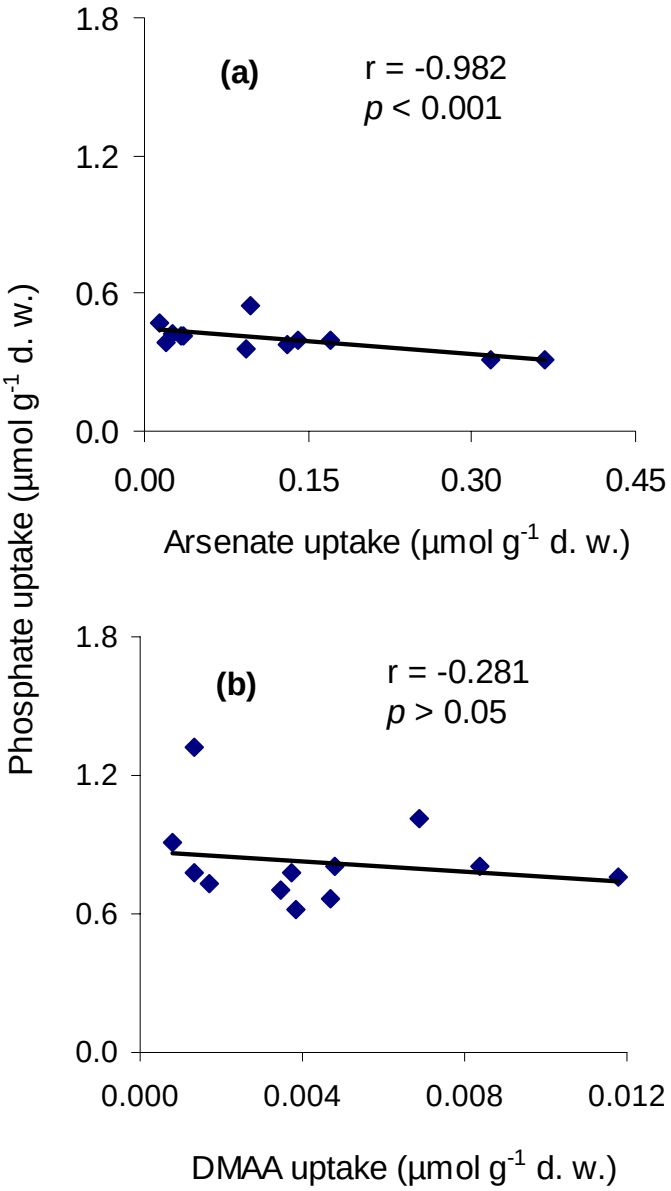
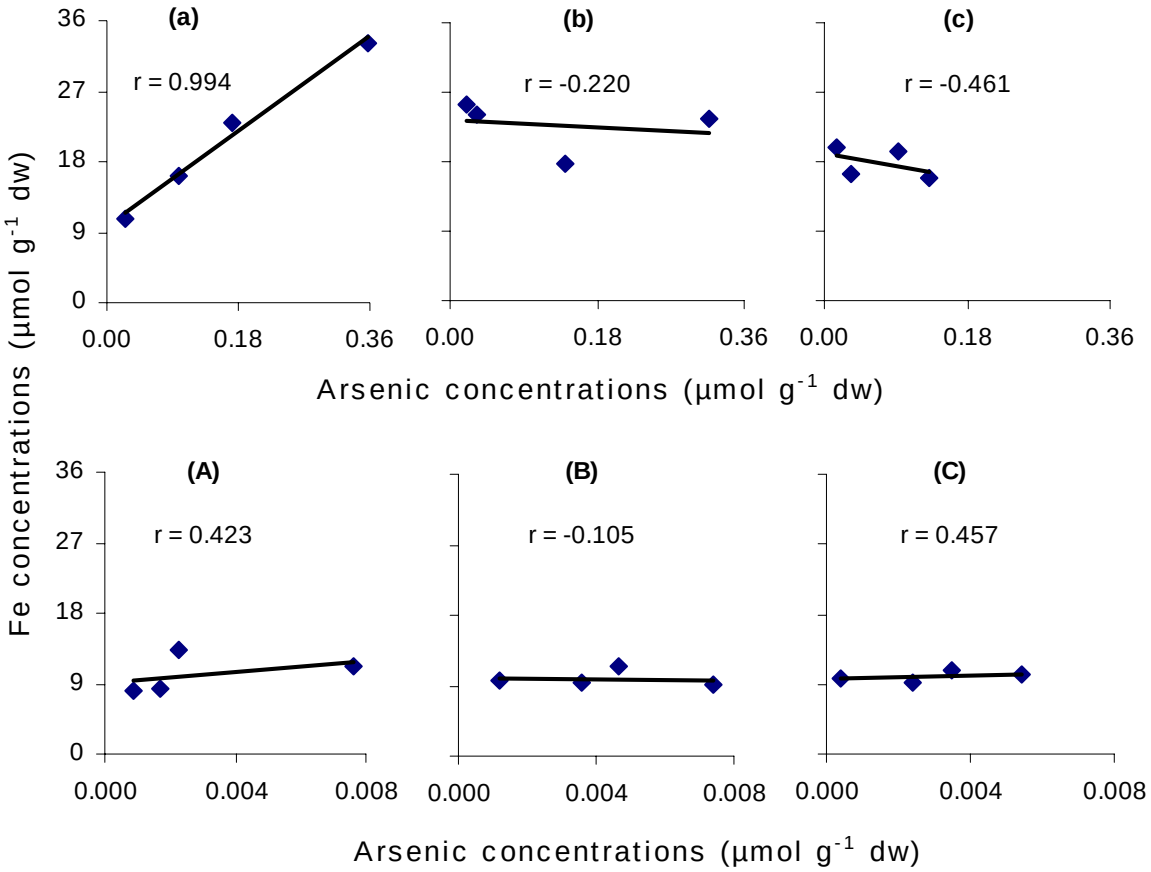


Figure 3: Relationship between arsenic and phosphate uptake in *S. polyrhiza* L. when the plant was exposed to arsenate (a) and DMAA (b).



510

511 **Figure 4:** Correlation between arsenic and iron concentrations in *S. polyrhiza* L. when the
512 plant was exposed to arsenate (above) and DMAA (bellow). PO_4^{3-} = 0.02 µM (a,
513 A); PO_4^{3-} = 100 µM (b, B); PO_4^{3-} = 500 µM (c, C).