

# Prolonged recurrence-free survival following OK432-stimulated dendritic cell transfer into hepatocellular carcinoma during transarterial embolization

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## ORIGINAL ARTICLE

**Prolonged Recurrence-Free Survival following OK432-Stimulated Dendritic Cell Transfer into Hepatocellular Carcinoma during Transarterial Embolization**

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Short Title: OK432-Stimulated DC Transfer into HCC during TAE

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## SUMMARY

Despite curative locoregional treatments for hepatocellular carcinoma (HCC), tumor recurrence rates remain high. The current study was designed to assess the safety and bioactivity of infusion of dendritic cells (DCs) stimulated with OK432, a Streptococcus-derived anticancer immunotherapeutic agent, into tumor tissues following transcatheter hepatic arterial embolization (TAE) treatment in patients with HCC. DCs were derived from peripheral blood monocytes of patients with hepatitis C virus-related cirrhosis and HCC in the presence of interleukin (IL)-4 and granulocyte-macrophage colony-stimulating factor and stimulated with 0.1 KE/ml OK432 for two days. Thirteen patients were administered with  $5 \times 10^6$  of DCs through arterial catheter during the procedures of TAE treatment on day 7. The immunomodulatory effects and clinical responses were evaluated in comparison with a group of 22 historical controls treated with TAE but without DC transfer. OK432 stimulation of immature DCs promoted their maturation towards cells with activated phenotypes, high expression of a homing receptor, fairly well-preserved phagocytic capacity, greatly enhanced cytokine production and effective tumoricidal activity. Administration of OK432-stimulated DCs to patients was found to be feasible and safe. Kaplan-Meier analysis revealed prolonged recurrence-free survival of patients treated in this manner compared with the historical controls ( $P = 0.046$ , log-rank test). The bioactivity of the transferred DCs was reflected in higher serum concentrations of the cytokines IL-9, IL-15 and tumor necrosis factor- $\alpha$  and the chemokines CCL4 and CCL11. Collectively, this study suggests that a DC-based, active immunotherapeutic strategy in combination with locoregional treatments exerts beneficial antitumor effects against liver cancer.

## INTRODUCTION

Many locoregional therapeutic approaches including surgical resection, radiofrequency ablation (RFA) and transcatheter hepatic arterial embolization (TAE) have been taken in the search for curative treatments of hepatocellular carcinoma (HCC). Despite these efforts, tumor recurrence rates remain high [1,2], probably because active hepatitis and cirrhosis in the surrounding non-tumor liver tissues causes de novo development of HCC [3,4]. One strategy to reduce tumor recurrence is to enhance antitumor immune responses that may induce sufficient inhibitory effects to prevent tumor cell growth and survival [5,6]. Dendritic cells (DCs) are the most potent type of antigen-presenting cells in the human body, and are involved in the regulation of both innate and adaptive immune responses [7]. DC-based immunotherapies are believed to contribute to the eradication of residual and recurrent tumor cells.

To enhance tumor antigen presentation to T lymphocytes, DCs have been transferred with major histocompatibility complex (MHC) class I and class II genes [8] and costimulatory molecules, e.g., CD40, CD80 and CD86 [9,10], and loaded with tumor-associated antigens including tumor lysates, peptides and RNA transfection [11]. To induce natural killer (NK) and natural killer T (NKT) cell activation, DCs have been stimulated and modified to produce larger amounts of cytokines, e.g., interleukin (IL)-12, IL-18 and type I interferons [10,12]. Furthermore, DC migration into secondary lymphoid organs could be induced by expression of chemokine genes, e.g., C-C chemokine receptor-7 (CCR7) [13], and by maturation using inflammatory cytokines [14], matrix metalloproteinases and Toll-like receptor (TLR) ligands [15].

DCs stimulated with OK432, a penicillin-inactivated and lyophilized preparation of *Streptococcus pyogenes*, were recently suggested to produce large amounts of T helper-1 (Th1)-type cytokines including IL-12 and interferon- $\gamma$  (IFN- $\gamma$ )

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6 and enhance cytotoxic T lymphocyte activity compared to a standard mixture of  
7 cytokines [tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), IL-1 $\beta$ , IL-6, and prostaglandin E2 (PGE2)]  
8 [16]. Furthermore, because OK432 modulates DC maturation through TLR4 and the  $\beta_2$   
9 integrin system [16,17] and TLR4-stimulated DCs can abrogate the activity of  
10 regulatory T cells [18], OK432-stimulated DCs may contribute to the induction of  
11 antitumor immune responses partly by reducing the activity of suppressor cells.  
12 Recently, in addition to the orchestration of immune responses, OK432-activated DCs  
13 have been shown themselves to mediate strong, specific cytotoxicity toward tumor cells  
14 via CD40 / CD40 ligand interactions [19].  
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26 Recently, we have reported that combination therapy using TAE together with  
27 immature DC infusion is safe for patients with cirrhosis and HCC [20]. DCs were  
28 infused precisely into tumor tissues and contributed to the recruitment and activation of  
29 immune cells in situ. However, this approach by itself yielded limited antitumor effects  
30 probably due to insufficient stimulation of immature DCs [the preparation of which  
31 seems closely related to therapeutic outcome [21,22]]. The current study was designed  
32 to assess the safety and bioactivity of OK432-stimulated DC infusion into tumor tissues  
33 following TAE treatment in patients with cirrhosis and HCC. In addition to  
34 documenting the safety of this approach, we found that patients treated with  
35 OK432-stimulated DCs displayed unique cytokine and chemokine profiles, and, most  
36 importantly, experienced prolonged recurrence-free survival.  
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## PATIENTS AND METHODS

### Patients.

Inclusion criteria were a radiological diagnosis of primary HCC by computed tomography (CT) angiography, hepatitis C virus (HCV)-related HCC, a Karnofsky score of  $\geq 70$  %, an age of  $\geq 20$  years, informed consent, and the following normal baseline hematological parameters (within 1 week before DC administration): hemoglobin  $\geq 8.5$  g/dl; white cell count  $\geq 2,000$  / $\mu$ l; platelet count  $\geq 50,000$  / $\mu$ l; creatinine  $< 1.5$  mg/dl, and liver damage A or B [23].

Exclusion criteria included severe cardiac, renal, pulmonary, hematological or other systemic disease associated with a discontinuation risk; human immunodeficiency virus (HIV) infection; prior history of other malignancies; history of surgery, chemotherapy or radiation therapy within four weeks; immunological disorders including splenectomy and radiation to the spleen; corticosteroid or antihistamine therapy; current lactation; pregnancy; history of organ transplantation; or difficulty in follow-up.

Thirteen patients (4 women and 9 men) presenting at Kanazawa University Hospital between March 2004 and June 2006 were enrolled in the study, with an age range from 56 to 83 years (**Table 1**). Patients with verified radiological diagnoses of HCC stage II or more were eligible and enrolled in this study. In addition, a group of twenty-two historical controls (9 women and 13 men) treated with TAE without DC administration between July 2000 and September 2007 was included in this study. All patients received RFA therapy to increase the locoregional effects one week later [24]. They underwent ultrasound, CT scan or magnetic resonance imaging (MRI) of the abdomen about one month after treatment and at a minimum of once every three months thereafter, and tumor recurrences were followed for up to 360 days. The Institutional

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6 Review Board reviewed and approved the study protocol. This study complied with  
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8 ethical standards outlined in the Declaration of Helsinki. Adverse events were monitored  
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10 for one month after the DC infusion in terms of fever, vomiting, abdominal pain,  
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12 encephalopathy, myalgia, ascites, gastrointestinal disorder, bleeding, hepatic abscess  
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14 and autoimmune diseases.  
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### 18 **Preparation and injection of autologous DCs.**

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20 DCs were generated from blood monocyte precursors as previously reported  
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22 [25]. Briefly, peripheral blood mononuclear cells (PBMCs) were isolated by  
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24 centrifugation in Lymphoprep™ Tubes (Nycomed, Roskilde, Denmark). For generating  
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26 DCs, PBMCs were plated in 6-well tissue culture dishes (Costar, Cambridge, MA) at  
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28  $1.4 \times 10^7$  cells in 2-mL per well and allowed to adhere to plastic for 2 h. Adherent cells  
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30 were cultured in serum-free media (GMP CellGro® DC Medium; CellGro, Manassas,  
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32 VA) with 50 ng/ml recombinant human IL-4 (GMP grade; CellGro®), and 100 ng/ml  
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34 recombinant human granulocyte-macrophage colony-stimulating factor (GM-CSF)  
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36 (GMP grade; CellGro®) for 5 days to generate immature DC, and matured for a further 2  
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38 days in 0.1 KE/ml OK432 (Chugai Pharmaceuticals, Tokyo, Japan) to generate OK-DC.  
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40 On day 7, the cells were harvested for injection,  $5 \times 10^6$  cells were suspended in 5 ml  
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42 normal saline containing 1 % autologous plasma, mixed with absorbable gelatin sponge  
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44 (Gelfoam; Pharmacia & Upjohn, Peapack, NJ) and infused through an arterial catheter  
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46 following Lipiodol (iodized oil) (Lipiodol Ultrafluide, Laboratoire Guerbet,  
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48 Aulnay-Sous-Bois, France) injection during selective TAE therapy. Release criteria for  
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50 DCs were viability >80%, purity >30%, negative gram stain and endotoxin polymerase  
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52 chain reaction (PCR), and negative in process cultures from samples sent 48 h before  
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54 release. All products met all release criteria, and the DCs had a typical phenotype of  
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56  $CD14^-$  and human leukocyte antigen (HLA)-DR<sup>+</sup>.  
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### **Flow cytometry analysis.**

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6 The DC preparation was assessed by staining with the following monoclonal  
7 antibodies for 30 min on ice: anti-lin 1 (lineage cocktail 1; CD3, CD14, CD16, CD19,  
8 CD20 and CD56)-fluorescein isothiocyanate (FITC), anti-HLA-DR-peridinin  
9 chlorophyll protein (PerCP) (L243), anti-CCR7-phycoerythrin (PE) (3D12) (BD  
10 Pharmingen, San Diego, CA), anti-CD80-PE (MAB104), anti-CD83-PE (HB15a) and  
11 anti-CD86-PE (HA5.2B7) (Beckman Coulter, Fullerton, CA). Cells were analyzed on a  
12 FACSCalibur™ flow cytometer. Data analysis was performed with CELLQuest™  
13 software (Becton Dickinson, San Jose, CA).  
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#### 24 **DC phagocytosis.**

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26 Immature DCs and OK432-stimulated DCs were incubated with 1 mg/ml FITC  
27 dextran (Sigma-Aldrich, St. Louis, MO) for 30 min at 37°C and the cells were washed  
28 three times in FACS buffer before cell acquisition using a FACSCalibur™ cytometer.  
29 Control DCs (not incubated with FITC dextran) were acquired at the same time to allow  
30 background levels of fluorescence to be determined.  
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#### 38 **Enzyme-linked immunosorbent assay (ELISA).**

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40 DCs were seeded at 200,000 cells/ml, and supernatant collected after 48 h.  
41 IL-12p40 and IFN- $\gamma$  were detected using matched paired antibodies (BD Pharmingen)  
42 following standard protocols.  
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#### 48 **Cytotoxicity assays.**

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50 The ability of DCs to exert cytotoxicity was assessed in a standard <sup>51</sup>Cr release  
51 assay [19]. We used the HCC cell lines Hep3B and PLC/PRF/5 (ATCC, Manassas, VA)  
52 and a lymphoblastoid cell line T2 that expresses HLA-A\*0201 (ATCC) as target cells.  
53 Target cells were labeled with <sup>51</sup>Cr. In a 96-well plate, 2.5 x 10<sup>3</sup> target cells per well  
54 were incubated with DCs for 8 h at different effector / target (E / T) ratios in triplicates.  
55 Percent specific lysis was calculated as follows: (experimental release - spontaneous  
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6 release) / (maximum release - spontaneous release) x 100. Spontaneous release was  
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8 always < 20 % of the total.  
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### 10 11 12 **NK cell activity.**

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14 NK cell cytotoxicity against K562 erythroleukemia target cells was measured  
15 by using <sup>51</sup>Cr-release assay according to previously published methods [26] with  
16 PBMCs obtained from the patients. All experiments were performed in triplicate.  
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18 Percent cytotoxicity was calculated as follows: [(experimental cpm – spontaneous cpm)  
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20 / (total cpm – spontaneous cpm)] X 100.  
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### 26 27 **Intracellular cytokine expression.**

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29 Freshly isolated PBMCs were stimulated with 25 ng/ml phorbol 12-myristate  
30 13-acetate (PMA, Sigma-Aldrich) and 1 µg/ml ionomycin (Sigma-Aldrich) at 37 °C in  
31 humidified 7 % CO<sub>2</sub> for 4 h. To block cytokine secretion, brefeldin A (Sigma, St. Louis,  
32 MO) [27] was added to a final concentration of 10 µg/mL. After addition of stimuli, the  
33 surface staining was performed with anti-CD4-PC5 (13B8.2), anti-CD8-PerCP (SK1)  
34 and anti-CD56-PC5 (N901) (Beckman Coulter). Subsequently, the cells were  
35 permeabilized, stained for intracellular IFN-γ and IL-4 using the FastImmune™ system  
36 (BD Pharmingen), resuspended in PBS containing 1 % paraformaldehyde (PFA), and  
37 analyzed on a flow cytometer (≈10,000 gated events acquired per sample).  
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### 48 49 **IFN-γ enzyme-linked immunospot (ELISPOT) assay.**

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51 ELISPOT assays were performed as previously described with the following  
52 modifications [28-30]. HLA-A24 restricted peptide epitopes, squamous cell carcinoma  
53 antigen recognized by T cells 2 (SART2)<sub>899</sub> (SYTRLFLIL), SART3<sub>109</sub>  
54 (VYDYNCHVDL), multidrug resistance protein 3 (MRP3)<sub>765</sub> (VYSDADIFL), MRP3<sub>503</sub>  
55 (LYAWEPSFL), MRP3<sub>692</sub> (AYVPQQAWI), alpha-fetoprotein (AFP)<sub>403</sub> (KYIQESQAL),  
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57 AFP<sub>434</sub> (AYTKKAPQL), AFP<sub>357</sub> (EYSRRHPQL), human telomerase reverse  
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6 transcriptase (hTERT)<sub>167</sub> (AYQVCGPPL) (unpublished), hTERT<sub>461</sub> (VYGFVRACL),  
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8 hTERT<sub>324</sub> (VYAETKHFL), were used in this study. Negative controls consisted of an  
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10 HIV envelope-derived peptide (HIVenv<sub>584</sub>). Positive controls consisted of 10 ng/ml  
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12 PMA (Sigma) or a CMV pp65-derived peptide (CMVpp65<sub>328</sub>). The colored spots were  
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14 counted with a KS ELISpot Reader (Zeiss, Tokyo, Japan). The number of specific spots  
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16 was determined by subtracting the number of spots in the absence of antigen from the  
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18 number of spots in its presence. Responses were considered positive if more than 10  
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20 specific spots were detected and if the number of spots in the presence of antigen was at  
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22 least two fold greater than the number of spots in the absence of antigen.  
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### 26 **Cytokine and chemokine profiling.**

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28 Serum cytokine and chemokine levels were measured using the Bioplex assay  
29  
30 (Bio-Rad, Hercules, CA). Briefly, frozen serum samples were thawed at room  
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32 temperature, diluted 1 : 4 in sample diluents, and 50  $\mu$ l aliquots of diluted sample were  
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34 added in duplicate to the wells of 96-well microtiter plate containing the coated beads  
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36 for a validated panel of 27 human cytokines and chemokines (cytokine 27-plex antibody  
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38 bead kit) according to the manufacturer's instructions. These included IL-1 $\beta$ , IL-1Ra,  
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40 IL-2, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12p70, IL-13, IL-15, IL-17, basic FGF,  
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42 eotaxin, G-CSF, GM-CSF, IFN- $\gamma$ , IP-10, MCP-1, MIP-1 $\alpha$ , MIP-1 $\beta$ , PDGF-BB,  
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44 RANTES, TNF- $\alpha$ , and VEGF. Eight standards (ranging from 2 to 32,000 pg/mL) were  
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46 used to generate calibration curves for each cytokine. Data acquisition and analysis were  
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48 done using Bio-Plex Manager software version 4.1.1.  
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### 52 **Arginase activity.**

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54 Serum samples were tested for arginase activity by conversion of L-arginine to  
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56 L-ornithine [31] using a kit supplied the manufacturer (BioAssay Systems, Hayward,  
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58 CA). Briefly, sera were treated with a membrane filter (Millipore, Billerica, MA) to  
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60 remove urea, combined with the sample buffer in wells of 96-well plate, and incubated

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6 at 37°C for 2 h. Subsequently, the urea reagent was added to stop the arginase reaction.  
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8 The color produced was read at 520 nm using a microtiter plate reader.  
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### 10 11 **Statistical analysis.**

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14 Results are expressed as means  $\pm$  SD. Differences between groups were  
15 analyzed for statistical significance by the Mann-Whitney U test. Qualitative variables  
16 were compared by means of Fisher's exact test. The estimated probability of tumor  
17 recurrence-free survival was determined using the Kaplan-Meier method. The Mantel  
18 Cox log-rank test was used to compare curves between groups. Any *P* values less than  
19 0.05 were considered statistically significant. All statistical tests were two-sided.  
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## RESULTS

### Preparation of OK432-stimulated DCs.

Adherent cells isolated from PBMCs of patients with cirrhosis and HCC (**Table 1**) were differentiated into DCs in the presence of IL-4 and GM-CSF. The cells were stimulated with 0.1 KE/ml OK432 for three days.  $54.6 \pm 9.5$  % (mean  $\pm$  SD; n = 13) of OK432-stimulated cells showed high levels of MHC class II (HLA-DR) and the absence of lineage markers including CD3, CD14, CD16, CD19, CD20 and CD56, in which  $30.9 \pm 14.2$  % were CD11c-positive (myeloid DC subset) and  $14.8 \pm 11.2$  were CD123-positive (plasmacytoid DC subset), consistent with our previous observations [20]. As reported [32,33], greater proportions of the cells developed high levels of expression of the costimulatory molecules B7-1 (CD80) and B7-2 (CD86) and an activation marker (CD83) compared to DCs prepared without OK432 stimulation (**Fig. 1A**). Furthermore, the chemokine receptor CCR7 which leads to homing to lymph nodes [13,34] was also induced following OK432 stimulation.

To evaluate the endocytic and phagocytic ability of the OK432-stimulated cells, uptake of FITC-dextran was quantitated by flow cytometry (**Fig. 1B**). The cells showed lower levels of uptake due to maturation compared to DCs prepared without OK432 stimulation, while the OK432-stimulated cells derived from HCC patients preserved a moderate uptake capacity. As expected, the OK432-stimulated cells produced large amounts of the cytokines IL-12 and IFN- $\gamma$  (**Fig. 1C**). In addition, they displayed high cytotoxic activity against HCC cell lines (Hep3B and PLC/PRF/5) and a lymphoblastoid cell line (T2) although DCs without OK432 stimulation lysed none of the target cells to any great degree (**Fig. 1D**). Taken together, these results demonstrate that OK432 stimulation of IL-4 and GM-CSF-induced immature DCs derived from HCC patients promoted their maturation towards cells with activated phenotypes, high expression of a homing receptor, fairly well-preserved phagocytic capacity, greatly enhanced cytokine

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6 production and effective tumoricidal activity, consistent with previous observations  
7 [16,19].  
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### 10 11 **Safety of OK432-stimulated DC administration.** 12

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14 Prior to the administration of OK432-stimulated DCs to patients, the cells were  
15 confirmed to be safe in athymic nude mice to which 100-fold cell numbers / weight  
16 were injected subcutaneously (data not shown). Subsequently, OK432-stimulated DC  
17 administration was performed during TAE therapy in humans, in which DCs were  
18 mixed together with absorbable gelatin sponge (Gelfoam) and infused through an  
19 arterial catheter following iodized oil (Lipiodol) injection as reported previously [20].  
20 Adverse events were monitored clinically and biochemically after DC infusion (**Table**  
21 **2**). A larger proportion (12 / 13) of the patients were complicated with high fever  
22 compared to those previously treated with immature DCs (5 / 10) [20], probably due to  
23 the proinflammatory responses induced by OK432-stimulated DCs. However, there  
24 were no grade III or IV National Cancer Institute Common Toxicity Criteria adverse  
25 events including vomiting, abdominal pain, encephalopathy, myalgia, ascites,  
26 gastrointestinal disorders, bleeding, hepatic abscess or autoimmune diseases associated  
27 with DC infusion and TAE in this study. There was also no clinical or serological  
28 evidence of hepatic failure or autoimmune response in any patients. Thus, concurrent  
29 treatment with OK432-stimulated DC infusions can be performed safely at the same  
30 time as TAE in patients with cirrhosis and HCC.  
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### 50 **Recurrence-free survival following DC infusion.** 51

52 A further objective of this study was to determine clinical response following  
53 DC infusion. A group of historical controls treated with TAE without DC administration  
54 was reviewed for this study (**Table 3**). The clinical characteristics including tumor  
55 burden and hepatic reserve were comparable between patients treated with TAE and  
56 OK432-stimulated DC transfer (n = 13) and those historical controls with TAE but  
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6 without DC administration (n = 22). We compared the recurrence-free survival between  
7 these patient groups. Kaplan-Meier analysis indicated that patients treated with TAE and  
8 OK432-stimulated DC transfer had prolonged recurrence-free survival compared with  
9 the historical controls that had been treated with TAE alone (recurrence rates 360 days  
10 after the treatments; 2 / 13 and 12 / 22, respectively;  $P = 0.046$ , log-rank test) (**Fig. 2**).  
11 The results demonstrated that OK432-stimulated DC transfer during TAE therapy  
12 reduces tumor recurrence in HCC patients.  
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### 22 **NK cell activity and intracellular cytokine responses in PBMCs.**

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24 To assess systemic immunomodulatory effects of OK432-stimulated DC  
25 transfer, PBMCs were isolated one and three months after treatment and NK cell  
26 cytotoxicity against K562 erythroleukemia target cells measured using the  $^{51}\text{Cr}$ -release  
27 assay (**Fig. 3**). The level of NK cell was unaltered following treatment. In addition,  
28 cytokine production capacity of lymphocyte subsets was quantitated by measuring  
29 intracellular IFN- $\gamma$  and IL-4 using flow cytometry. There were also no significant  
30 changes in terms of cytokine production capacity in the CD4 $^{+}$ , CD8 $^{+}$  and CD56 $^{+}$  subsets  
31 in the patients treated with OK432-stimulated DCs.  
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### 42 **Immune responses to peptide epitopes derived from tumor antigens.**

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44 To assess the effects on T cell responses to tumor antigens, PBMCs were  
45 obtained 4 weeks after DC infusion, pulsed with peptides derived from AFP, MRP3,  
46 SART2, SART3 and hTERT. IFN- $\gamma$  production was then quantitated in an ELISPOT  
47 assay. Cells producing IFN- $\gamma$  in response to stimulation with HLA-A24 [the most  
48 common HLA-A antigen (58.1%) in Japanese populations [35]]-restricted peptide  
49 epitopes derived from tumor antigens MRP3 and hTERT were induced in 3 of 6  
50 HLA-A24-positive patients (numbers 2, 6 and 11) after treatment with TAE and  
51 OK432-stimulated DCs (**Fig. 4**). To understand the immunological and clinical  
52 significance of the T lymphocyte responses, PBMCs obtained from the historical control  
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6 patients who had been treated with TAE without DC administration were also evaluated  
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8 by ELISPOT. Similarly, positive reactions were observed in 4 (numbers t8, t19, t20 and  
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10 t22) of 6 HLA-A24-positive patients. These data indicate that T lymphocyte responses  
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12 to HLA-A24 restricted peptide epitopes of tumor antigens were induced following the  
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14 TAE therapy, but no additional responses were observed as a result of  
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16 OK432-stimulated DC transfer in the current study.  
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### 20 **Serum levels of cytokines, chemokines and arginase activity.**

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22 To screen for immunobiological responses induced following  
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24 OK432-stimulated DC transfer, serum levels of cytokines and chemokines were  
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26 measured simultaneously using the Bio-Plex multiplex suspension array system. The  
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28 results were compared with the historical control patients treated with TAE without DC  
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30 administration. Interestingly, serum concentrations of IL-9, IL-15 and TNF- $\alpha$  were  
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32 greatly increased after OK432-stimulated DC infusion, in contrast to their reduction  
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34 following TAE treatment alone (**Fig. 5A**). Furthermore, the chemokines eotaxin  
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36 (CCL11) and MIP-1 $\beta$  (CCL4) were markedly induced after DC transfer although they  
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38 were also decreased after TAE alone. These data indicate that transfer of  
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40 OK432-stimulated DC during TAE therapy induced unique immune responses that may  
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42 be mediated by the cytokines IL-9, IL-15 and TNF- $\alpha$  and the chemokines eotaxin and  
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44 MIP-1 $\beta$ .  
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51 In addition, serum arginase activity was reported to reflect numbers of  
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53 myeloid-derived suppressor cells (MDSCs) that may inhibit T lymphocyte responses in  
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55 cancer patients [36]. Therefore, serum arginase activity was measured after  
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57 OK432-stimulated DC infusion, and it was found that it was increased 6- or 7-fold in  
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59 patients treated with TAE. However, this increase was independent of the presence or  
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activity was decreased again 4 weeks after treatment with both TAE and

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6 OK432-stimulated DC transfer but tended to be maintained at a high level in patients  
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8 treated with TAE without DC transfer. However, these differences did not reach  
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10 statistical significance ( $P > 0.05$ ). Because arginase activity is known to be relatively  
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12 high in liver and HCC cells [37], the influence of tissue injury was assessed  
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14 biochemically by measuring serum levels of alanine aminotransferase (ALT) and lactic  
15  
16 dehydrogenase (LDH) activities. We did not observed ALT or LDH elevation, indicating  
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18 that the increase of arginase activity was not due to tissue damage following treatment.  
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20 Collectively, these results demonstrate that infusion of OK432-stimulated DCs during  
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22 TAE treatment may reduce the immunosuppressive activities of MDSCs, and assist in  
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24 developing a favorable environment for the induction of antitumor immunity.  
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## DISCUSSION

Although many novel strategies including immunotherapies have been developed in an attempt to suppress tumor recurrence after curative treatments for HCC, recurrence rates and survival times have not been improved significantly [38]. In the current study, we first established that OK432-stimulated DC administration during TAE therapy did not cause critical adverse events in patients with cirrhosis and HCC. Most importantly, DC transfer resulted in prolonged recurrence-free survival after combination therapy with TAE and OK432-stimulated DC administration. In terms of the immunomodulatory effects of DC transfer, although NK cell activity, intracellular cytokine production, and T lymphocyte-mediated immune responses were not altered in PBMCs from treated patients, serum levels of IL-9, IL-15 and TNF- $\alpha$  and the chemokines eotaxin and MIP-1 $\beta$  were markedly enhanced after DC transfer. In addition, serum levels of arginase activity were decreased following DC transfer. Collectively, this study demonstrated the feasibility, safety, and beneficial antitumor effects of OK432-stimulated DC infusion into tumor tissues for patients with cirrhosis and HCC, suggesting the ability of an active immunotherapeutic strategy to reduce tumor recurrence after locoregional treatment of HCC.

DCs were stimulated with OK432 prior to infusion into tumor tissues through an arterial catheter. OK432 was reported to activate DCs through its binding to TLR-2 and -4 [16,39] that can be used for cancer therapy [33]. The current results indicate that OK432 stimulation of immature DCs from HCC patients promoted their maturation processes while preserving antigen uptake capacity and enhancing tumoricidal activity, consistent with previous observations [16,19] and supporting the current strategy in which OK432-stimulated DCs were directly infused into tumor tissues. Since the tumoricidal activity of unstimulated DCs was not observed in *in vitro* experiments, OK432 stimulation obviously altered the cytotoxic properties of DCs. One of the

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6 mechanisms of DC killing was reported to be CD40 / CD40 ligand interaction [19]. The  
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8 further studies are needed to determine the killing mechanisms of DCs derived from  
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10 HCC patients in a direct (TNF, TRAIL, Fas ligand, NO and perforin / granzyme) and  
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12 indirect (MHC-restricted) manner [40-43]. Although the main mechanism by which  
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14 OK432-stimulated DCs prolonged the recurrence-free survival was not elucidated, the  
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16 tumoricidal activity of mature DCs was implicated in in vivo enhancement of antigen  
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18 presentation, costimulation and inflammatory cytokine production.  
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22 Very recent reports document injection of OK432-stimulated DCs into patients  
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24 with cancer of the gastrointestinal tract or pancreas [44,45], but their antitumor effects  
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26 were not clearly defined. The current study shows for the first time that  
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28 OK432-stimulated DCs induce beneficial antitumor responses when transferred into  
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30 tumor tissues during TAE therapy. The antitumor responses may have been enhanced as  
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32 a result of optimal activation of the DCs with OK432 or combining infusion of  
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34 stimulated DCs with TAE therapy. Inappropriately activated DCs may be unable to  
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36 generate sufficient numbers of properly activated effector T lymphocytes [46]. As  
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38 shown in **Fig. 1**, all of these alterations could contribute to the further enhancement of  
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40 antitumor effects compared to those in our previous study with immature DCs [20].  
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42 Furthermore, the tumor cell death-promoting therapies, e.g. chemotherapy [47] and TAE  
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44 [48], can be expected to enhance the effects of therapeutic cancer vaccines by redressing  
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46 the immunosuppressive tumor environment.  
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51 NK cell activity and intracellular cytokine responses in CD4<sup>+</sup> and CD8<sup>+</sup> T  
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53 lymphocytes and CD56<sup>+</sup> NK cell subsets in PBMCs were not significantly changed in  
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55 patients treated with OK432-stimulated DCs. Furthermore, we did not observe tumor  
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57 antigen-specific T lymphocyte responses clearly associated with DC administration. The  
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59 data therefore suggest that the immune responses induced by the therapy applied here  
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were not systemically detectable. Because cytotoxic T lymphocyte responses were

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6 enhanced in patients receiving  $> 3 \times 10^7$  cells [49,50], the numbers of transferred  
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8 OK432-stimulated DCs were apparently not sufficient to induce responses detectable in  
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10 the peripheral blood, but were enough to exert beneficial antitumor effects. In addition,  
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12 many studies have concluded that cytotoxic T lymphocyte responses rarely predict  
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14 clinical outcomes of DC-based immunotherapies [51,52], and that in many cases, also  
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16 including our own studies [28,30], tumor-specific effector T lymphocytes coexist with  
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18 the tumors. Consistent with these observations, the current results suggest that cytotoxic  
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20 T lymphocyte responses in PBMCs are not reliable predictors of beneficial antitumor  
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22 effects in patients treated with the current OK432-stimulated DC strategy.  
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26 Serum levels of the cytokines IL-9, IL-15 and TNF- $\alpha$  and the chemokines  
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28 eotaxin and MIP-1 $\beta$  were increased following OK432-stimulated DC transfer, but  
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30 decreased after TAE therapy without DC administration. IL-9 and IL-15 belong to the  
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32 cytokine receptor common gamma chain ( $\gamma_c$ ; CD132) family, a member of the Type I  
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34 cytokine receptor family expressed on most lymphocyte populations [53]. IL-9 exerts  
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36 pleiotropic activities on T and B lymphocytes, mast cells, monocytes and hematopoietic  
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38 progenitors [54,55]. IL-15 and TNF- $\alpha$  are known to prime T lymphocytes and NK cells  
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40 when secreted by DCs [56] and to induce antitumor immune responses [57]. Eotaxin is  
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42 known to selectively recruit eosinophils also contributing to antitumor effects [58,59],  
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44 and MIP-1 $\beta$  is a chemoattractant for NK cells, monocytes and a variety of other  
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46 immune cells [60]. In addition, serum levels of arginase tended to decrease after DC  
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48 transfer. Because serum arginase activity reflects the numbers of MDSCs that inhibit T  
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50 lymphocyte responses in cancer patients [36], the patients treated with  
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52 OK432-stimulated DCs might have developed lower levels of suppressor cells.  
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54 Collectively, the results suggest that infusion of OK432-stimulated DCs may orchestrate  
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56 the immune environment in the whole body that could enhance beneficial antitumor  
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58 effects, although the precise molecular and cellular mechanisms associated with the  
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60 actions of these cytokines and chemokines were not clearly defined in the current

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analysis.

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**DISCLOSURE**

The authors have declared that no conflict of interest exists.

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## REFERENCES

1. Omata M, Tateishi R, Yoshida H, Shiina S. Treatment of hepatocellular carcinoma by percutaneous tumor ablation methods: Ethanol injection therapy and radiofrequency ablation. *Gastroenterology* 2004; 127:S159-66.
2. Belghiti J. Resection and liver transplantation for HCC. *J Gastroenterol* 2009; 44 Suppl 19:132-5.
3. Nakamoto Y, Guidotti LG, Kuhlen CV, Fowler P, Chisari FV. Immune pathogenesis of hepatocellular carcinoma. *J Exp Med* 1998; 188:341-50.
4. Ercolani G, Grazi GL, Ravaioli M, et al. Liver resection for hepatocellular carcinoma on cirrhosis: univariate and multivariate analysis of risk factors for intrahepatic recurrence. *Ann Surg* 2003; 237:536-43.
5. Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ, Schreiber RD. IFN $\gamma$  and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature* 2001; 410:1107-11.
6. Vulink A, Radford KJ, Melief C, Hart DN. Dendritic cells in cancer immunotherapy. *Adv Cancer Res* 2008; 99:363-407.
7. Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu YJ, Pulendran B, Palucka K. Immunobiology of dendritic cells. *Annu Rev Immunol* 2000; 18:767-811.
8. Lemos MP, Esquivel F, Scott P, Laufer TM. MHC class II expression restricted to CD8 $\alpha$ <sup>+</sup> and CD11b<sup>+</sup> dendritic cells is sufficient for control of *Leishmania major*. *J Exp Med* 2004; 199:725-30.
9. Ni K, O'Neill HC. The role of dendritic cells in T cell activation. *Immunol Cell Biol* 1997; 75:223-30.
10. Andrews DM, Andoniou CE, Scalzo AA, van Dommelen SL, Wallace ME, Smyth MJ, Degli-Esposti MA. Cross-talk between dendritic cells and natural killer cells in

- 1  
2  
3  
4  
5  
6  
7 viral infection. *Mol Immunol* 2005; 42:547-55.
- 8  
9 11. Heiser A, Coleman D, Dannull J, et al. Autologous dendritic cells transfected with  
10 prostate-specific antigen RNA stimulate CTL responses against metastatic prostate  
11 tumors. *J Clin Invest* 2002; 109:409-17.
- 12  
13 12. Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature*  
14 1998; 392:245-52.
- 15  
16 13. Forster R, Schubel A, Breitfeld D, Kremmer E, Renner-Muller I, Wolf E, Lipp M.  
17 CCR7 coordinates the primary immune response by establishing functional  
18 microenvironments in secondary lymphoid organs. *Cell* 1999; 99:23-33.
- 19  
20 14. MartIn-Fontecha A, Sebastiani S, Hopken UE, Ugucioni M, Lipp M,  
21 Lanzavecchia A, Sallusto F. Regulation of dendritic cell migration to the draining  
22 lymph node: impact on T lymphocyte traffic and priming. *J Exp Med* 2003;  
23 198:615-21.
- 24  
25 15. Ratzinger G, Stoitzner P, Ebner S, et al. Matrix metalloproteinases 9 and 2 are  
26 necessary for the migration of Langerhans cells and dermal dendritic cells from  
27 human and murine skin. *J Immunol* 2002; 168:4361-71.
- 28  
29 16. Nakahara S, Tsunoda T, Baba T, Asabe S, Tahara H. Dendritic cells stimulated with  
30 a bacterial product, OK-432, efficiently induce cytotoxic T lymphocytes specific to  
31 tumor rejection peptide. *Cancer Res* 2003; 63:4112-8.
- 32  
33 17. Okamoto M, Oshikawa T, Tano T, et al. Mechanism of anticancer host response  
34 induced by OK-432, a streptococcal preparation, mediated by phagocytosis and  
35 Toll-like receptor 4 signaling. *J Immunother* 2006; 29:78-86.
- 36  
37 18. Pasare C, Medzhitov R. Toll pathway-dependent blockade of CD4+CD25+ T  
38 cell-mediated suppression by dendritic cells. *Science* 2003; 299:1033-6.
- 39  
40 19. Hill KS, Errington F, Steele LP, et al. OK432-activated human dendritic cells kill  
41 tumor cells via CD40/CD40 ligand interactions. *J Immunol* 2008; 181:3108-15.
- 42  
43 20. Nakamoto Y, Mizukoshi E, Tsuji H, et al. Combined therapy of transcatheter  
44  
45  
46  
47  
48  
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2  
3  
4  
5  
6  
7 hepatic arterial embolization with intratumoral dendritic cell infusion for  
8  
9 hepatocellular carcinoma: clinical safety. *Clin Exp Immunol* 2007; 147:296-305.
- 10  
11 21. Steinman RM, Banchereau J. Taking dendritic cells into medicine. *Nature* 2007;  
12  
13 449:419-26.
- 14  
15 22. Tacken PJ, de Vries IJ, Torensma R, Figdor CG. Dendritic-cell immunotherapy:  
16  
17 from ex vivo loading to in vivo targeting. *Nat Rev Immunol* 2007; 7:790-802.
- 18  
19 23. Makuuchi M. General rules for the clinical and pathological study of primary liver  
20  
21 cancer, 2nd English ed. Tokyo, Japan: Kanehara & Co., Ltd., 2003.
- 22  
23 24. Veltri A, Moretto P, Doriguzzi A, Pagano E, Carrara G, Gandini G. Radiofrequency  
24  
25 thermal ablation (RFA) after transarterial chemoembolization (TACE) as a  
26  
27 combined therapy for unresectable non-early hepatocellular carcinoma (HCC). *Eur*  
28  
29 *Radiol* 2006; 16:661-9.
- 30  
31 25. Dhodapkar MV, Steinman RM, Sapp M, et al. Rapid generation of broad T-cell  
32  
33 immunity in humans after a single injection of mature dendritic cells. *J Clin Invest*  
34  
35 1999; 104:173-80.
- 36  
37 26. Orange JS, Brodeur SR, Jain A, et al. Deficient natural killer cell cytotoxicity in  
38  
39 patients with IKK-gamma/NEMO mutations. *J Clin Invest* 2002; 109:1501-9.
- 40  
41 27. Klausner RD, Donaldson JG, Lippincott-Schwartz J. Brefeldin A: insights into the  
42  
43 control of membrane traffic and organelle structure. *J Cell Biol* 1992; 116:1071-80.
- 44  
45 28. Mizukoshi E, Nakamoto Y, Marukawa Y, et al. Cytotoxic T cell responses to human  
46  
47 telomerase reverse transcriptase in patients with hepatocellular carcinoma.  
48  
49 *Hepatology* 2006; 43:1284-94.
- 50  
51 29. Mizukoshi E, Nakamoto Y, Tsuji H, Yamashita T, Kaneko S. Identification of  
52  
53 alpha-fetoprotein-derived peptides recognized by cytotoxic T lymphocytes in  
54  
55 HLA-A24+ patients with hepatocellular carcinoma. *Int J Cancer* 2006;  
56  
57 118:1194-204.
- 58  
59 30. Mizukoshi E, Honda M, Arai K, Yamashita T, Nakamoto Y, Kaneko S. Expression  
60

- 1  
2  
3  
4  
5  
6  
7 of multidrug resistance-associated protein 3 and cytotoxic T cell responses in  
8 patients with hepatocellular carcinoma. *J Hepatol* 2008; 49:946-54.
- 9  
10  
11 31. Rodriguez PC, Quiceno DG, Zabaleta J, et al. Arginase I production in the tumor  
12 microenvironment by mature myeloid cells inhibits T-cell receptor expression and  
13 antigen-specific T-cell responses. *Cancer Res* 2004; 64:5839-49.
- 14  
15  
16  
17 32. Itoh T, Ueda Y, Okugawa K, et al. Streptococcal preparation OK432 promotes  
18 functional maturation of human monocyte-derived dendritic cells. *Cancer Immunol*  
19 *Immunother* 2003; 52:207-14.
- 20  
21  
22  
23 33. Kuroki H, Morisaki T, Matsumoto K, Onishi H, Baba E, Tanaka M, Katano M.  
24 Streptococcal preparation OK-432: a new maturation factor of monocyte-derived  
25 dendritic cells for clinical use. *Cancer Immunol Immunother* 2003; 52:561-8.
- 26  
27  
28  
29 34. Gunn MD, Kyuwa S, Tam C, Kakiuchi T, Matsuzawa A, Williams LT, Nakano H.  
30 Mice lacking expression of secondary lymphoid organ chemokine have defects in  
31 lymphocyte homing and dendritic cell localization. *J Exp Med* 1999; 189:451-60.
- 32  
33  
34  
35 35. Imanishi T, Akaza T, Kimura A, Tokunaga K, Gojobori T. HLA 1991, Proceeding of  
36 the Eleventh International Histocompatibility Workshop and Conference. Tokyo,  
37 Japan: Oxford University Press, 1992.
- 38  
39  
40  
41 36. Zea AH, Rodriguez PC, Atkins MB, et al. Arginase-producing myeloid suppressor  
42 cells in renal cell carcinoma patients: a mechanism of tumor evasion. *Cancer Res*  
43 2005; 65:3044-8.
- 44  
45  
46  
47 37. Chrzanowska A, Krawczyk M, Baranczyk-Kuzma A. Changes in arginase  
48 isoenzymes pattern in human hepatocellular carcinoma. *Biochem Biophys Res*  
49 *Commun* 2008; 377:337-40.
- 50  
51  
52  
53 38. Caldwell S, Park SH. The epidemiology of hepatocellular cancer: from the  
54 perspectives of public health problem to tumor biology. *J Gastroenterol* 2009; 44  
55 *Suppl* 19:96-101.
- 56  
57  
58  
59 39. Okamoto M, Oshikawa T, Tano T, et al. Involvement of Toll-like receptor 4  
60

- 1  
2  
3  
4  
5  
6  
7 signaling in interferon-gamma production and antitumor effect by streptococcal  
8 agent OK-432. *J Natl Cancer Inst* 2003; 95:316-26.
- 9  
10  
11 40. Liu S, Yu Y, Zhang M, Wang W, Cao X. The involvement of TNF-alpha-related  
12 apoptosis-inducing ligand in the enhanced cytotoxicity of IFN-beta-stimulated  
13 human dendritic cells to tumor cells. *J Immunol* 2001; 166:5407-15.
- 14  
15  
16 41. Lu G, Janjic BM, Janjic J, Whiteside TL, Storkus WJ, Vujanovic NL. Innate direct  
17 anticancer effector function of human immature dendritic cells. II. Role of TNF,  
18 lymphotoxin-alpha(1)beta(2), Fas ligand, and TNF-related apoptosis-inducing  
19 ligand. *J Immunol* 2002; 168:1831-9.
- 20  
21  
22 42. Nicolas A, Cathelin D, Larmonier N, et al. Dendritic cells trigger tumor cell death  
23 by a nitric oxide-dependent mechanism. *J Immunol* 2007; 179:812-8.
- 24  
25  
26 43. Stry G, Bangert C, Tauber M, Strohal R, Kopp T, Stingl G. Tumoricidal activity of  
27 TLR7/8-activated inflammatory dendritic cells. *J Exp Med* 2007; 204:1441-51.
- 28  
29  
30 44. West E, Morgan R, Scott K, et al. Clinical grade OK432-activated dendritic cells: in  
31 vitro characterization and tracking during intralymphatic delivery. *J Immunother*  
32 2009; 32:66-78.
- 33  
34  
35 45. Hirooka Y, Itoh A, Kawashima H, et al. A combination therapy of gemcitabine with  
36 immunotherapy for patients with inoperable locally advanced pancreatic cancer.  
37 *Pancreas* 2009; 38:e69-74.
- 38  
39  
40 46. Melief CJ. Cancer immunotherapy by dendritic cells. *Immunity* 2008; 29:372-83.
- 41  
42  
43 47. Zitvogel L, Apetoh L, Ghiringhelli F, Kroemer G. Immunological aspects of cancer  
44 chemotherapy. *Nat Rev Immunol* 2008; 8:59-73.
- 45  
46  
47 48. Ayaru L, Pereira SP, Alisa A, et al. Unmasking of alpha-fetoprotein-specific CD4(+)  
48 T cell responses in hepatocellular carcinoma patients undergoing embolization. *J*  
49 *Immunol* 2007; 178:1914-22.
- 50  
51  
52 49. Thurner B, Haendle I, Roder C, et al. Vaccination with mage-3A1 peptide-pulsed  
53 mature, monocyte-derived dendritic cells expands specific cytotoxic T cells and  
54  
55  
56  
57  
58  
59  
60

- 1  
2  
3  
4  
5  
6  
7 induces regression of some metastases in advanced stage IV melanoma. *J Exp Med*  
8 1999; 190:1669-78.  
9
- 10  
11 50. Banchereau J, Palucka AK, Dhodapkar M, et al. Immune and clinical responses in  
12 patients with metastatic melanoma to CD34(+) progenitor-derived dendritic cell  
13 vaccine. *Cancer Res* 2001; 61:6451-8.  
14  
15  
16  
17 51. Engell-Noerregaard L, Hansen TH, Andersen MH, Thor Straten P, Svane IM.  
18 Review of clinical studies on dendritic cell-based vaccination of patients with  
19 malignant melanoma: assessment of correlation between clinical response and  
20 vaccine parameters. *Cancer Immunol Immunother* 2009; 58:1-14.  
21  
22  
23  
24  
25 52. Itoh K, Yamada A, Mine T, Noguchi M. Recent advances in cancer vaccines: an  
26 overview. *Jpn J Clin Oncol* 2009; 39:73-80.  
27  
28  
29 53. Sugamura K, Asao H, Kondo M, Tanaka N, Ishii N, Nakamura M, Takeshita T. The  
30 common gamma-chain for multiple cytokine receptors. *Adv Immunol* 1995;  
31 59:225-77.  
32  
33  
34  
35 54. Temann UA, Geba GP, Rankin JA, Flavell RA. Expression of interleukin 9 in the  
36 lungs of transgenic mice causes airway inflammation, mast cell hyperplasia, and  
37 bronchial hyperresponsiveness. *J Exp Med* 1998; 188:1307-20.  
38  
39  
40  
41 55. McMillan SJ, Bishop B, Townsend MJ, McKenzie AN, Lloyd CM. The absence of  
42 interleukin 9 does not affect the development of allergen-induced pulmonary  
43 inflammation nor airway hyperreactivity. *J Exp Med* 2002; 195:51-7.  
44  
45  
46  
47  
48 56. de Saint-Vis B, Fugier-Vivier I, Massacrier C, et al. The cytokine profile expressed  
49 by human dendritic cells is dependent on cell subtype and mode of activation. *J*  
50 *Immunol* 1998; 160:1666-76.  
51  
52  
53  
54 57. Shanmugham LN, Petrarca C, Frydas S, et al. IL-15 an immunoregulatory and  
55 anti-cancer cytokine. Recent advances. *J Exp Clin Cancer Res* 2006; 25:529-36.  
56  
57  
58 58. Kataoka S, Konishi Y, Nishio Y, Fujikawa-Adachi K, Tominaga A. Antitumor  
59 activity of eosinophils activated by IL-5 and eotaxin against hepatocellular  
60

1  
2  
3  
4  
5  
6  
7 carcinoma. *DNA Cell Biol* 2004; 23:549-60.

- 8  
9 59. Simson L, Ellyard JI, Dent LA, Matthaei KI, Rothenberg ME, Foster PS, Smyth MJ,  
10 Parish CR. Regulation of carcinogenesis by IL-5 and CCL11: a potential role for  
11 eosinophils in tumor immune surveillance. *J Immunol* 2007; 178:4222-9.  
12  
13 60. Bystry RS, Aluvihare V, Welch KA, Kallikourdis M, Betz AG. B cells and  
14 professional APCs recruit regulatory T cells via CCL4. *Nat Immunol* 2001;  
15 2:1126-32.  
16  
17  
18  
19  
20  
21  
22  
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## FIGURE LEGENDS

**Figure 1.** Effects of OK432 stimulation on the properties of dendritic cells (DCs) generated from blood monocyte precursors in patients with cirrhosis and hepatocellular carcinoma (HCC) (n = 13). **(A)** lin<sup>+</sup>HLA-DR<sup>-</sup> subsets with [OK432(+)] and without [OK432(-)] stimulation were analyzed for surface expression of CD80, CD83, CD86 and CCR7. Dot plots of a representative case are shown in the left-hand panel. Mean percentages ( $\pm$  SD) of positive cells are indicated in the right-hand panel. OK432 stimulation resulted in the expression of high levels of CD80, CD83, CD86 and CCR7 in the lin<sup>+</sup>HLA-DR<sup>-</sup> DC subset. **(B)** DC subsets with and without OK432 stimulation were incubated with fluorescein isothiocyanate (FITC) dextran for 30 min and the uptake was determined by flow cytometry. A representative analysis is shown in the upper panel. Mean fluorescence intensities (MFIs) ( $\pm$  SD) of the positive cells are indicated in the lower panel. OK432-stimulated cells showed lower levels of uptake due to maturation. **(C)** DC supernatants were harvested and the concentrations of IL-12 and IFN- $\gamma$  measured by ELISA. OK432-stimulated cells produced large amounts of the cytokines. The data indicate means  $\pm$  SD of the groups with and without the stimulation. All comparisons in A - C [OK432(+) vs. OK432(-)] were statistically significant by the Mann-Whitney U test ( $P < 0.005$ ). **(D)** Tumoricidal activity of DCs assessed by incubation with <sup>51</sup>Cr-labeled Hep3B, PLC/PRF/5 and T2 targets for 8 h at the indicated E / T (effector / target) cell ratios. OK432-stimulated cells displayed high cytotoxic activity against the target cells. The results are representative of the cases studied.

**Figure 2.** Recurrence-free survival of patients treated with transcatheter hepatic arterial embolization (TAE) with (TAE + OK-DC; n = 13) and without (TAE: historical controls; n = 22) OK432-stimulated DC administration. Time zero is the date of TAE. All patients underwent ultrasound, CT scan or MRI of the abdomen about 1 month after treatment and at a minimum of once every 3 months thereafter. Kaplan-Meier analysis

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6 indicated that TAE + OK-DC treatment prolonged recurrence-free survival compared  
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8 with the TAE-alone group (recurrence rates 360 days after the treatments; 2 / 13 and 12  
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10 / 22, respectively;  $P = 0.046$ , log-rank test).  
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14 **Figure 3.** Natural killer (NK) cell activity and intracellular cytokine production in  
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16 PBMCs of patients treated with OK432-stimulated DCs during TAE therapy (n = 13).  
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18 PBMCs were isolated before and one and three months after treatment and used for the  
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20 analyses. Upper panel, NK cell cytotoxicity against K562 erythroleukemia target cells  
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22 was evaluated at the E / T cell ratios shown. NK cell activities were not changed  
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24 following treatment. Middle and lower panels, PBMCs were stimulated with phorbol  
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26 12-myristate 13-acetate (PMA) and ionomycin, stained for CD4, CD8 and CD56  
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28 expression, permeabilized, and stained for intracellular IFN- $\gamma$  and IL-4. Percentages of  
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30 cytokine-positive cells were quantitated by flow cytometry. There were no significant  
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32 changes in terms of cytokine production capacity in the CD4<sup>+</sup>, CD8<sup>+</sup> and CD56<sup>+</sup> subsets  
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34 following the treatments. The data are given as means  $\pm$  SD of the groups.  
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39 **Figure 4.** Immune responses to HLA-A24-restricted peptide epitopes derived from  
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41 tumor antigens in HLA-A24-positive patients treated with OK432-stimulated DCs  
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43 during TAE therapy (numbers 2, 5, 6, 7, 10 and 11) and HLA-A24-positive historical  
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45 controls treated with TAE without DC transfer (numbers t8, t14, t15, t19, t20 and t22).  
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47 PBMCs were obtained before (open bars) and one month after the infusion (solid bars),  
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49 pulsed with the peptides derived from SART2, SART3, MRP3, AFP and hTERT, and  
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51 IFN- $\gamma$  production was quantitated by ELISPOT. Negative controls consisted of an HIV  
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53 envelope-derived peptide (HIVenv<sub>584</sub>). Positive controls consisted of 10 ng/ml PMA or a  
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55 CMV pp65-derived peptide (CMVpp65<sub>328</sub>). The number of specific spots was  
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57 determined by subtracting the number of spots in the absence of antigen from the  
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59 number of spots in its presence. T lymphocyte responses to the peptide epitopes were  
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induced following TAE therapy, but no additional responses were observed after DC

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6 transfer. Numbers denote specific spots beyond the upper limit of y-axis. n.d., not  
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8 determined.  
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12 **Figure 5.** Cytokine and chemokine profiling and arginase activity in sera of patients  
13 treated with OK432-stimulated DCs during TAE therapy (TAE + OK-DC; n = 13) and  
14 the historical controls treated with TAE without DC transfer (TAE; n = 22). (A) Serum  
15 samples were examined for their content of a validated panel of cytokines and  
16 chemokines using the Bioplex assay. Percentage changes in serum levels two weeks  
17 after the treatments were calculated as follows: [(posttreatment level – pretreatment  
18 level) / pretreatment level] X 100. The data are means ± SEM of the groups. \**P* < 0.05  
19 when compared by the Mann-Whitney U test. (B) Serum samples were tested for  
20 arginase activity by conversion of L-arginine to L-ornithine, and for alanine  
21 aminotransferase (ALT) and lactic dehydrogenase (LDH) activities. While there was a  
22 trend for the arginase activity in the TAE + OK-DC group to decrease four weeks after  
23 treatment, the difference did not reach statistical significance (*P* > 0.05). Percentage  
24 changes in serum levels two weeks after the treatments were calculated as follows:  
25 [(posttreatment level – pretreatment level) / pretreatment level] X 100. The data indicate  
26 means ± SEM of the groups.  
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Table 1. Patient Characteristics

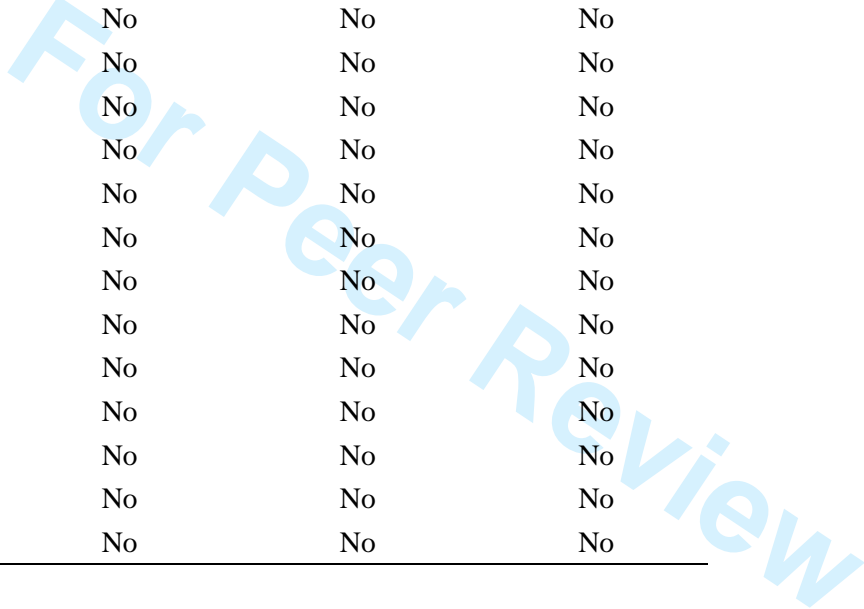
| Patient no. | Gender | Age (yrs) | HLA     | TNM stages | No. of tumors | Largest tumor (mm) | Child-Pugh | KPS | Post-TAE Rx |
|-------------|--------|-----------|---------|------------|---------------|--------------------|------------|-----|-------------|
| 1           | M      | 60        | A11 A33 | III        | 5             | 35                 | B          | 100 | RFA         |
| 2           | M      | 57        | A11 A24 | III        | 1             | 21                 | B          | 100 | RFA         |
| 3           | M      | 57        | A11 A31 | III        | 2             | 39                 | B          | 100 | RFA         |
| 4           | M      | 77        | A2 A24  | III        | 2             | 35                 | A          | 100 | RFA         |
| 5           | F      | 83        | A11 A24 | III        | 3             | 29                 | B          | 100 | RFA         |
| 6           | F      | 74        | A2 A24  | II         | 1             | 35                 | A          | 100 | RFA         |
| 7           | F      | 72        | A24 A33 | III        | 3             | 41                 | B          | 100 | RFA         |
| 8           | F      | 65        | A2 A11  | II         | 4             | 12                 | B          | 100 | RFA         |
| 9           | M      | 71        | A2 A11  | II         | 4             | 16                 | A          | 100 | RFA         |
| 10          | M      | 79        | A11 A24 | III        | 2             | 40                 | A          | 100 | RFA         |
| 11          | M      | 71        | A2 A24  | II         | 1             | 28                 | A          | 100 | RFA         |
| 12          | M      | 56        | A2 A26  | III        | 2             | 25                 | B          | 100 | RFA         |
| 13          | M      | 64        | A2 A33  | III        | 2             | 37                 | B          | 100 | RFA         |

TNM, tumor-node-metastasis; Child-Pugh, Child-Pugh classification; KPS, Karnofsky performance scores; TAE, transcatheter arterial embolization; Rx, Treatment; HCC, hepatocellular carcinoma; RFA, percutaneous radiofrequency ablation.

Table 2. Adverse Events

| Patient no. | Fever (days) | Vomiting | Abdominal Pain | Encephalopathy | Others* |
|-------------|--------------|----------|----------------|----------------|---------|
| 1           | 2            | No       | No             | No             | No      |
| 2           | 2            | No       | No             | No             | No      |
| 3           | 1            | No       | No             | No             | No      |
| 4           | 3            | No       | No             | No             | No      |
| 5           | 3            | No       | No             | No             | No      |
| 6           | 4            | No       | No             | No             | No      |
| 7           | 10           | No       | No             | No             | No      |
| 8           | No           | No       | No             | No             | No      |
| 9           | 2            | No       | No             | No             | No      |
| 10          | 1            | No       | No             | No             | No      |
| 11          | 2            | No       | No             | No             | No      |
| 12          | 2            | No       | No             | No             | No      |
| 13          | 1            | No       | No             | No             | No      |

\*Others of adverse events include myalgia, ascites, gastrointestinal disorder, bleeding, hepatic abscess and autoimmune diseases.



**Table 3. Clinical Characteristics of Patients Treated with TAE + OK-DC and TAE alone**

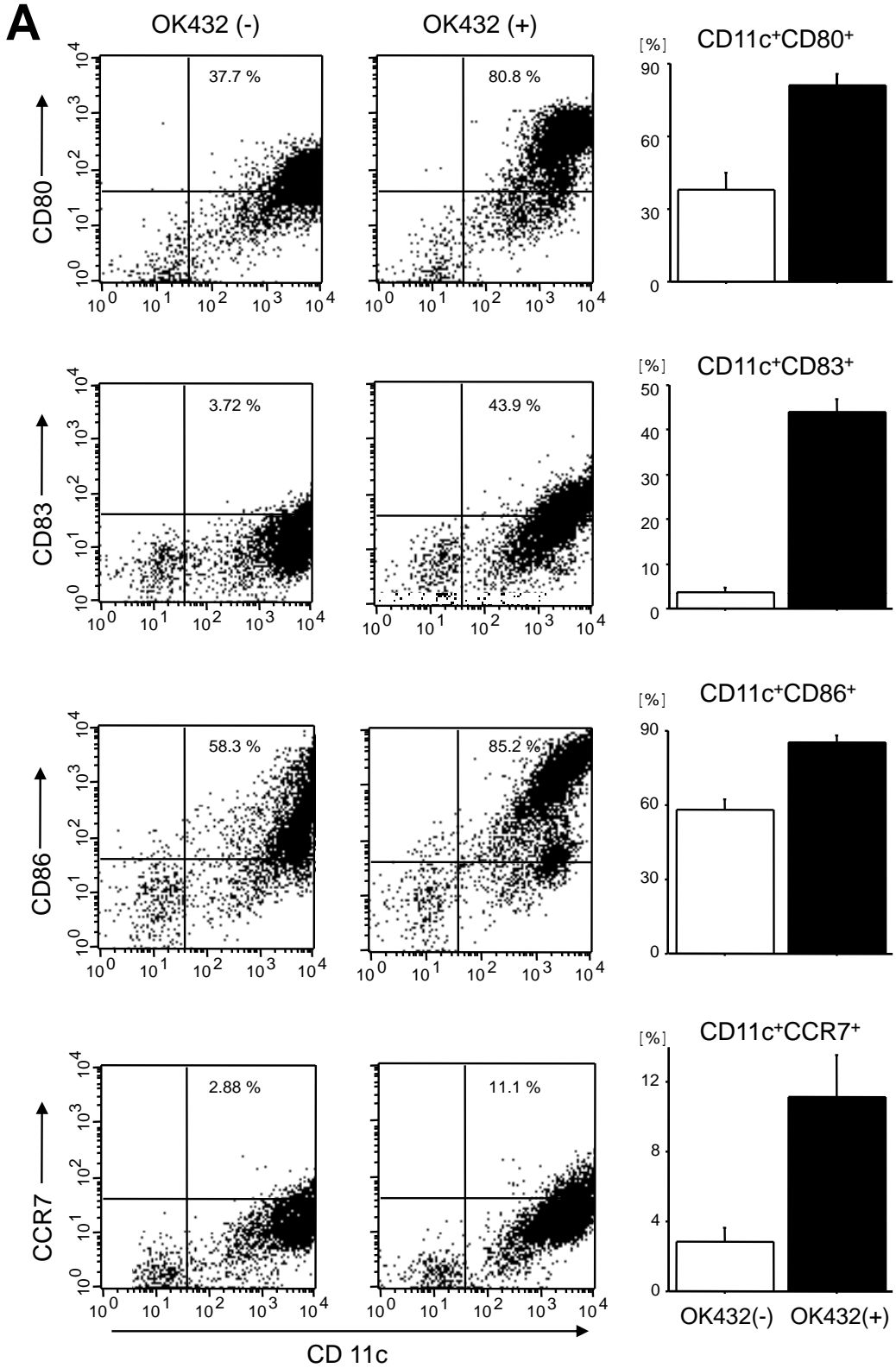
|   | TAE + OK-DC   | TAE           | <i>P</i> |
|---|---------------|---------------|----------|
| No. of patients                         | 13            | 22            |          |
| Age (yrs)                               | 68.2 ± 9.1    | 70.0 ± 7.6    | NS*      |
| Gender (M/F)                            | 9 / 4         | 13 / 9        | NS†      |
| White cell count (×10 <sup>2</sup> /μL) | 34.4 ± 11.6   | 41.4 ± 18.9   | NS*      |
| Lymphocytes (×10 <sup>2</sup> /μL)      | 10.4 ± 3.6    | 12.4 ± 4.7    | NS*      |
| Platelets (×10 <sup>4</sup> /μL)        | 11.5 ± 10.2   | 10.3 ± 5.8    | NS*      |
| Hepaplastin test (%)                    | 64.6 ± 11.6   | 75.5 ± 24.3   | NS*      |
| ALT (IU/L)                              | 56.7 ± 38.9   | 67.9 ± 44.6   | NS*      |
| Total bilirubin (mg/dL)                 | 1.3 ± 0.7     | 1.1 ± 0.6     | NS*      |
| Albumin (g/dL)                          | 3.4 ± 0.6     | 3.6 ± 0.4     | NS*      |
| Noncancerous liver parenchyma (no.)     |               |               |          |
| Chronic hepatitis                       | 0             | 8             |          |
| Cirrhosis (Child-Pugh A/B/C)            | 13 (5/8/0)    | 14 (6/8/0)    | NS†      |
| TNM stages (I/II/III/IV-A/IV-B)         | 0/4/9/0/0     | 3/8/11/0/0    | NS†      |
| No. of tumors                           | 2.5 ± 1.3     | 1.9 ± 1.3     | NS*      |
| Largest tumor (mm)                      | 30.2 ± 9.4    | 32.6 ± 15.2   | NS*      |
| AFP                                     | 204.8 ± 404.1 | 201.8 ± 544.2 | NS*      |

Results are expressed as means ± SD. \*Mann-Whitney U test. †Fisher's exact test. TAE, transcatheter arterial embolization; OK-DC, OK432-stimulated dendritic cells; ALT, alanine transaminase; TNM, tumor-node-metastasis; AFP, alpha-fetoprotein; NS, not significant.

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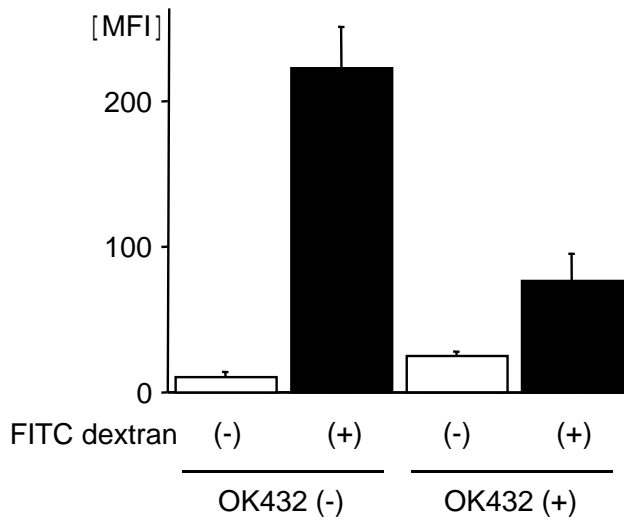
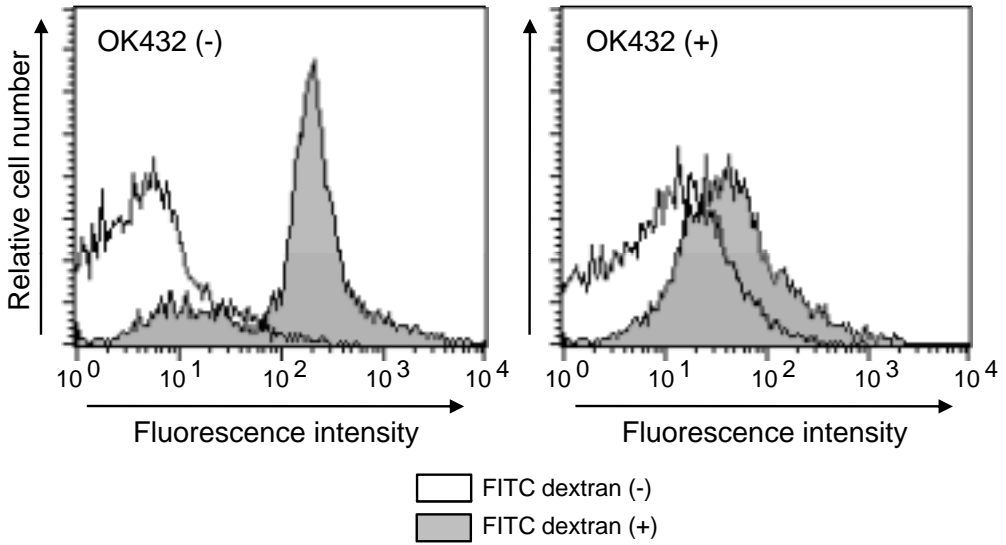
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Fig.1A

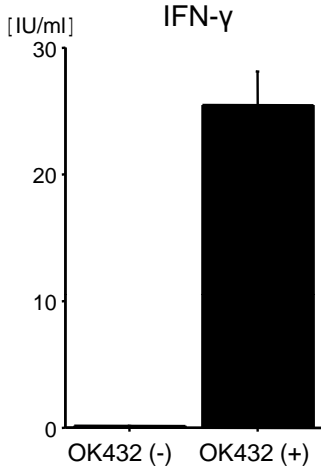
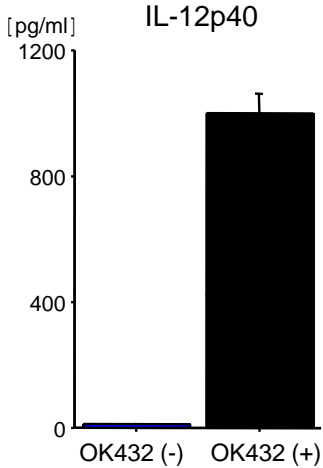


**B**

FITC dextran uptake



**C**



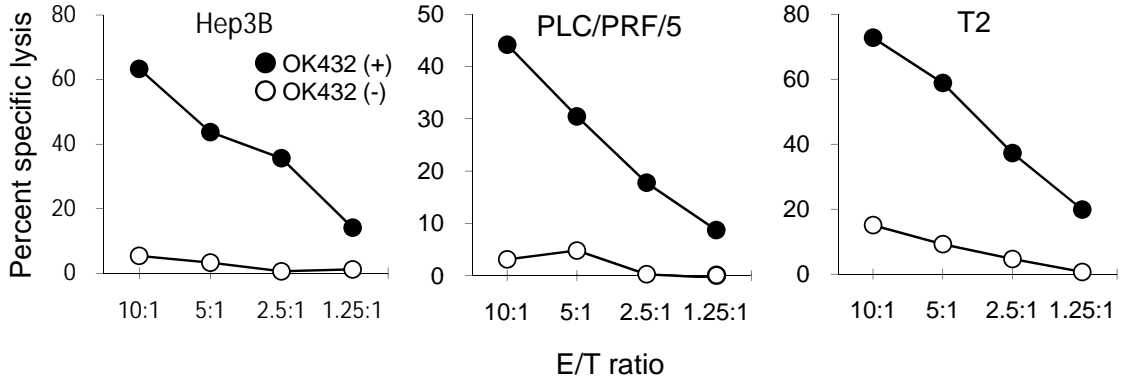
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Fig.2

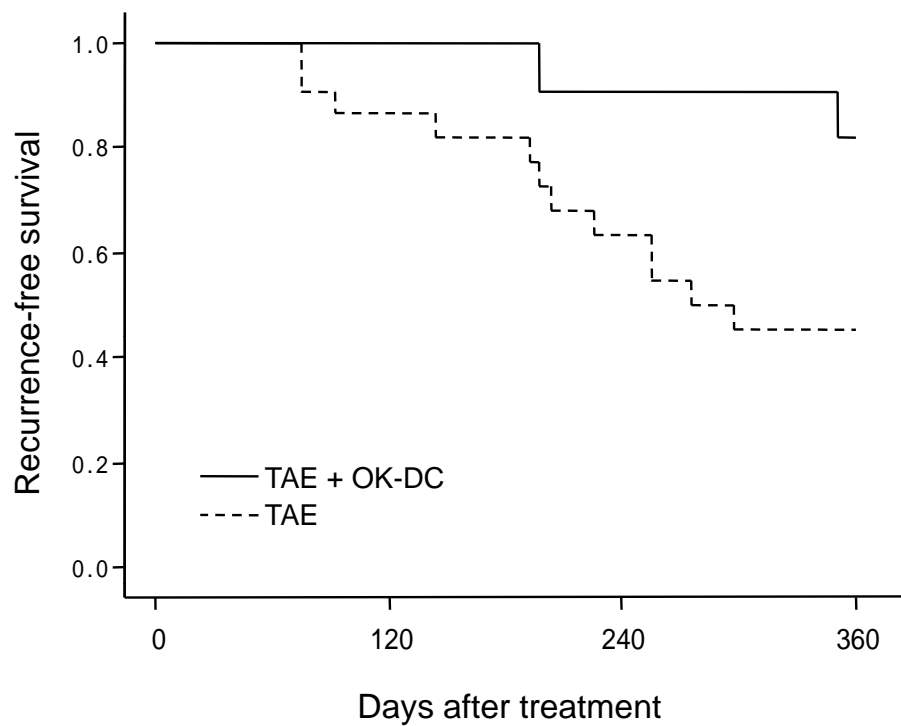


Fig.3

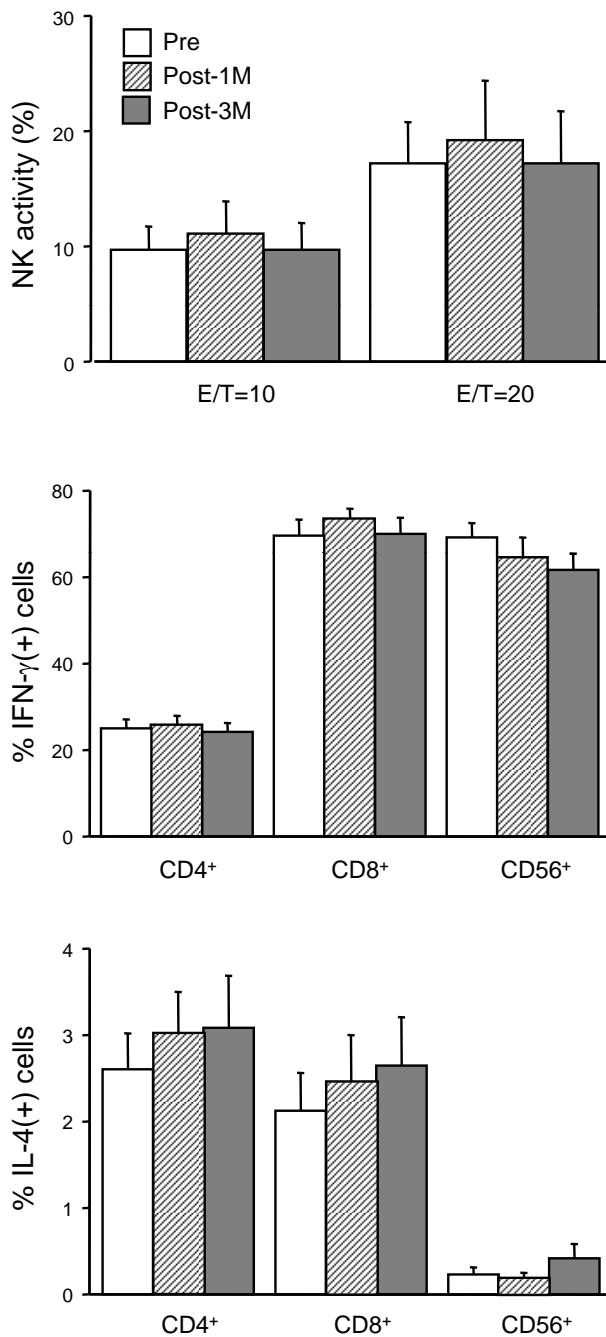
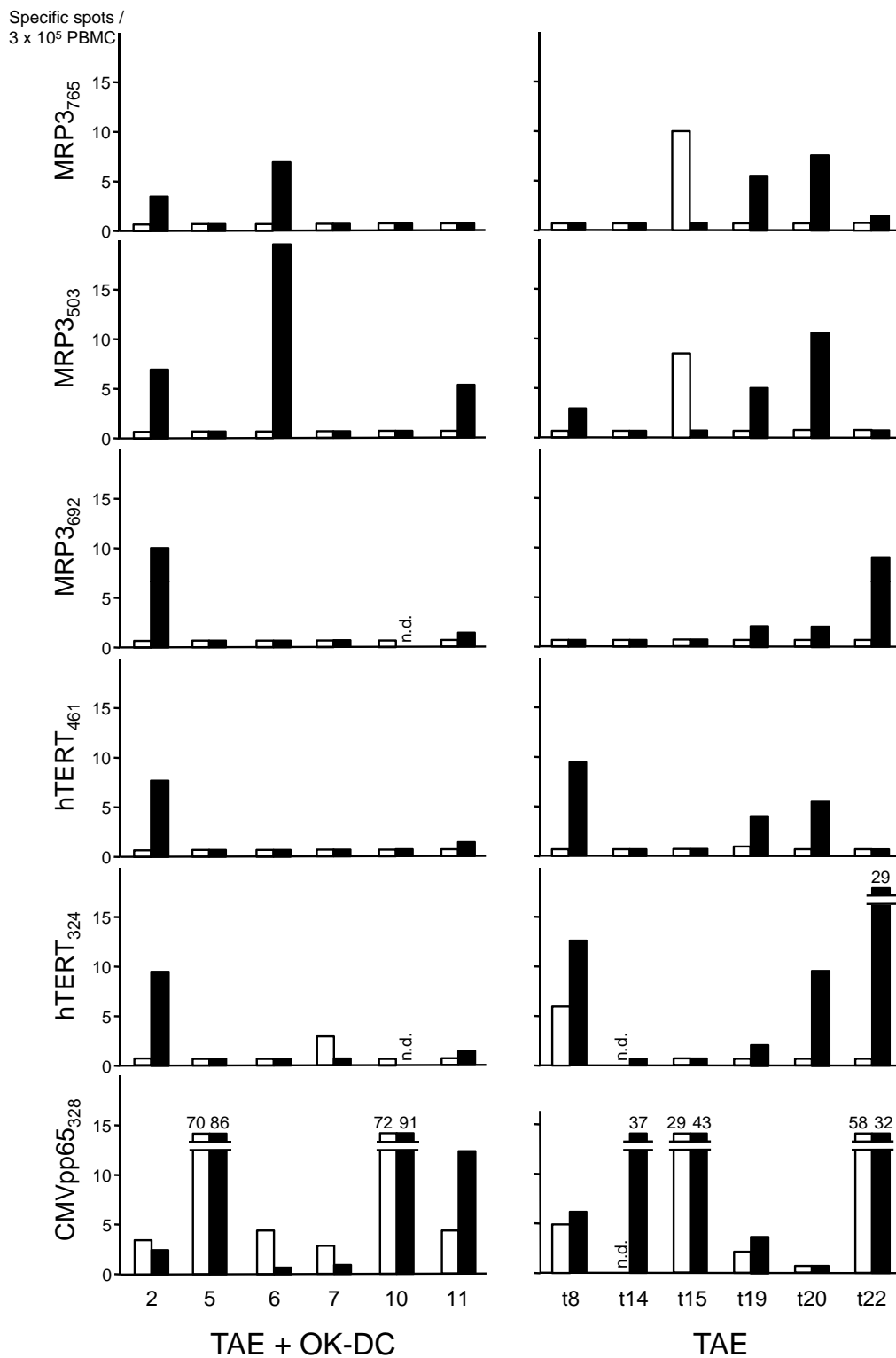
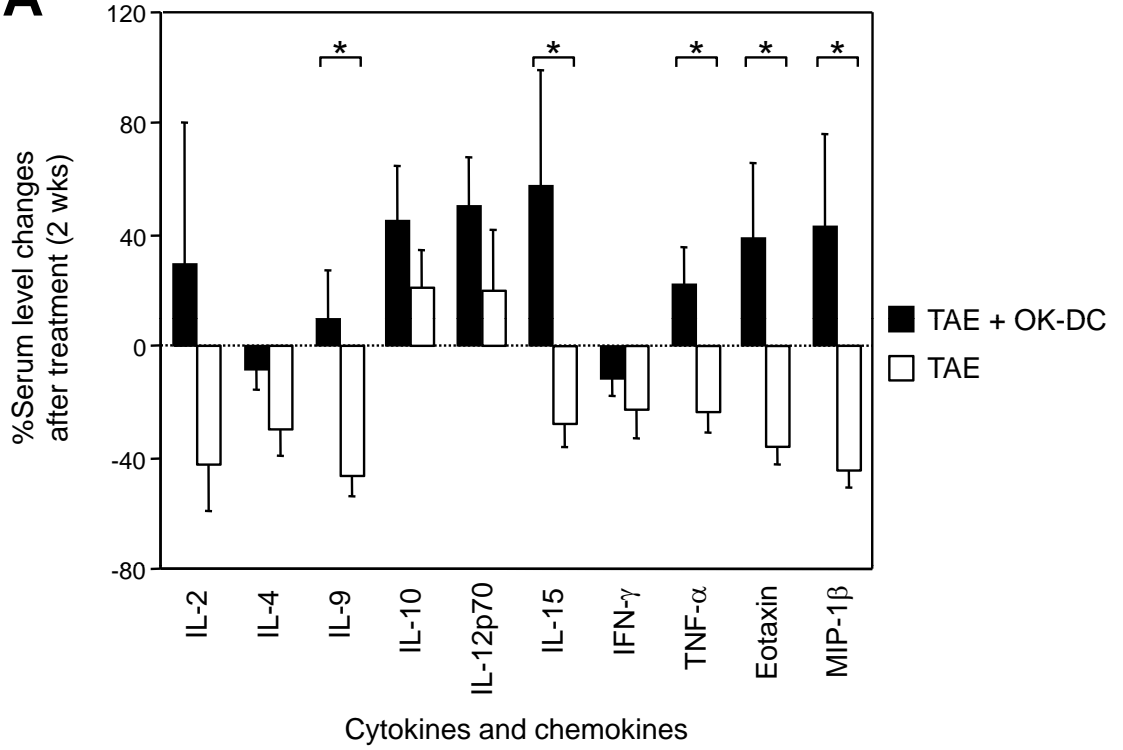
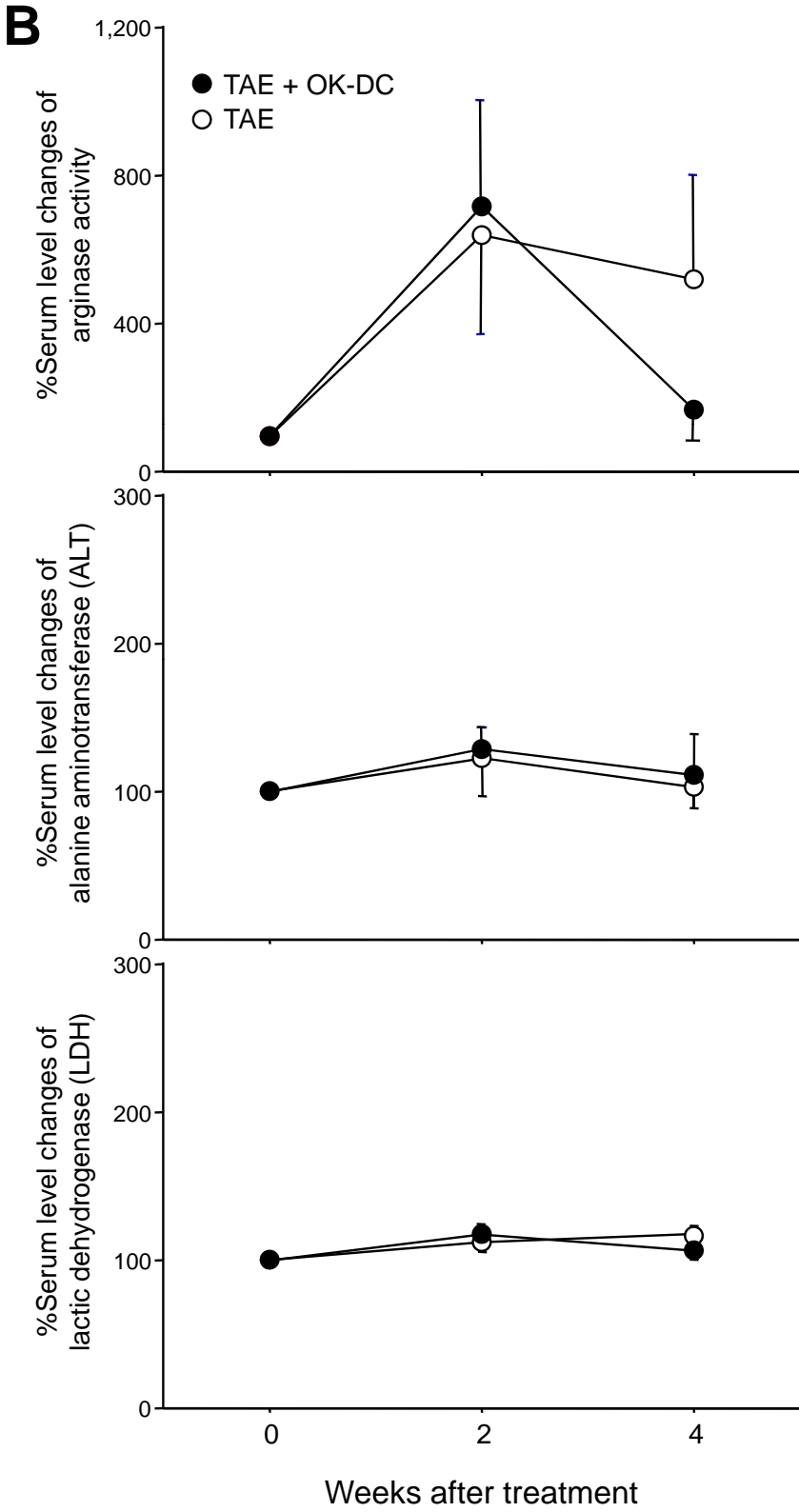


Fig.4



**A**





[Changes are UNDERLINED]

**Prolonged Recurrence-Free Survival following OK432-Stimulated Dendritic Cell  
Transfer into Hepatocellular Carcinoma during Transarterial Embolization**

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Short Title: OK432-Stimulated DC Transfer into HCC during TAE

Keywords: hepatocellular carcinoma, dendritic cells, immunotherapy,  
transcatheter hepatic arterial embolization, recurrence-free survival.

## SUMMARY

Despite curative locoregional treatments for hepatocellular carcinoma (HCC), tumor recurrence rates remain high. The current study was designed to assess the safety and bioactivity of infusion of dendritic cells (DCs) stimulated with OK432, a Streptococcus-derived anticancer immunotherapeutic agent, into tumor tissues following transcatheter hepatic arterial embolization (TAE) treatment in patients with HCC. DCs were derived from peripheral blood monocytes of patients with hepatitis C virus-related cirrhosis and HCC in the presence of interleukin (IL)-4 and granulocyte-macrophage colony-stimulating factor and stimulated with 0.1 KE/ml OK432 for two days. Thirteen patients were administered with  $5 \times 10^6$  of DCs through arterial catheter during the procedures of TAE treatment on day 7. The immunomodulatory effects and clinical responses were evaluated in comparison with a group of 22 historical controls treated with TAE but without DC transfer. OK432 stimulation of immature DCs promoted their maturation towards cells with activated phenotypes, high expression of a homing receptor, fairly well-preserved phagocytic capacity, greatly enhanced cytokine production and effective tumoricidal activity. Administration of OK432-stimulated DCs to patients was found to be feasible and safe. Kaplan-Meier analysis revealed prolonged recurrence-free survival of patients treated in this manner compared with the historical controls ( $P = 0.046$ , log-rank test). The bioactivity of the transferred DCs was reflected in higher serum concentrations of the cytokines IL-9, IL-15 and tumor necrosis factor- $\alpha$  and the chemokines CCL4 and CCL11. Collectively, this study suggests that a DC-based, active immunotherapeutic strategy in combination with locoregional treatments exerts beneficial antitumor effects against liver cancer.

## INTRODUCTION

Many locoregional therapeutic approaches including surgical resection, radiofrequency ablation (RFA) and transcatheter hepatic arterial embolization (TAE) have been taken in the search for curative treatments of hepatocellular carcinoma (HCC). Despite these efforts, tumor recurrence rates remain high [1,2], probably because active hepatitis and cirrhosis in the surrounding non-tumor liver tissues causes de novo development of HCC [3,4]. One strategy to reduce tumor recurrence is to enhance antitumor immune responses that may induce sufficient inhibitory effects to prevent tumor cell growth and survival [5,6]. Dendritic cells (DCs) are the most potent type of antigen-presenting cells in the human body, and are involved in the regulation of both innate and adaptive immune responses [7]. DC-based immunotherapies are believed to contribute to the eradication of residual and recurrent tumor cells.

To enhance tumor antigen presentation to T lymphocytes, DCs have been transferred with major histocompatibility complex (MHC) class I and class II genes [8] and costimulatory molecules, e.g., CD40, CD80 and CD86 [9,10], and loaded with tumor-associated antigens including tumor lysates, peptides and RNA transfection [11]. To induce natural killer (NK) and natural killer T (NKT) cell activation, DCs have been stimulated and modified to produce larger amounts of cytokines, e.g., interleukin (IL)-12, IL-18 and type I interferons [10,12]. Furthermore, DC migration into secondary lymphoid organs could be induced by expression of chemokine genes, e.g., C-C chemokine receptor-7 (CCR7) [13], and by maturation using inflammatory cytokines [14], matrix metalloproteinases and Toll-like receptor (TLR) ligands [15].

DCs stimulated with OK432, a penicillin-inactivated and lyophilized preparation of *Streptococcus pyogenes*, were recently suggested to produce large amounts of T helper-1 (Th1)-type cytokines including IL-12 and interferon- $\gamma$  (IFN- $\gamma$ )



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6 and enhance cytotoxic T lymphocyte activity compared to a standard mixture of  
7 cytokines [tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), IL-1 $\beta$ , IL-6, and prostaglandin E2 (PGE2)]  
8 [16]. Furthermore, because OK432 modulates DC maturation through TLR4 and the  $\beta_2$   
9 integrin system [16,17] and TLR4-stimulated DCs can abrogate the activity of  
10 regulatory T cells [18], OK432-stimulated DCs may contribute to the induction of  
11 antitumor immune responses partly by reducing the activity of suppressor cells.  
12 Recently, in addition to the orchestration of immune responses, OK432-activated DCs  
13 have been shown themselves to mediate strong, specific cytotoxicity toward tumor cells  
14 via CD40 / CD40 ligand interactions [19].  
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26 Recently, we have reported that combination therapy using TAE together with  
27 immature DC infusion is safe for patients with cirrhosis and HCC [20]. DCs were  
28 infused precisely into tumor tissues and contributed to the recruitment and activation of  
29 immune cells in situ. However, this approach by itself yielded limited antitumor effects  
30 probably due to insufficient stimulation of immature DCs [the preparation of which  
31 seems closely related to therapeutic outcome [21,22]]. The current study was designed  
32 to assess the safety and bioactivity of OK432-stimulated DC infusion into tumor tissues  
33 following TAE treatment in patients with cirrhosis and HCC. In addition to  
34 documenting the safety of this approach, we found that patients treated with  
35 OK432-stimulated DCs displayed unique cytokine and chemokine profiles, and, most  
36 importantly, experienced prolonged recurrence-free survival.  
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## PATIENTS AND METHODS

### Patients.

Inclusion criteria were a radiological diagnosis of primary HCC by computed tomography (CT) angiography, hepatitis C virus (HCV)-related HCC, a Karnofsky score of  $\geq 70$  %, an age of  $\geq 20$  years, informed consent, and the following normal baseline hematological parameters (within 1 week before DC administration): hemoglobin  $\geq 8.5$  g/dl; white cell count  $\geq 2,000$  / $\mu$ l; platelet count  $\geq 50,000$  / $\mu$ l; creatinine  $< 1.5$ mg/dl, and liver damage A or B [23].

Exclusion criteria included severe cardiac, renal, pulmonary, hematological or other systemic disease associated with a discontinuation risk; human immunodeficiency virus (HIV) infection; prior history of other malignancies; history of surgery, chemotherapy or radiation therapy within four weeks; immunological disorders including splenectomy and radiation to the spleen; corticosteroid or antihistamine therapy; current lactation; pregnancy; history of organ transplantation; or difficulty in follow-up.

Thirteen patients (4 women and 9 men) presenting at Kanazawa University Hospital between March 2004 and June 2006 were enrolled in the study, with an age range from 56 to 83 years (**Table 1**). Patients with verified radiological diagnoses of HCC stage II or more were eligible and enrolled in this study. In addition, a group of twenty-two historical controls (9 women and 13 men) treated with TAE without DC administration between July 2000 and September 2007 was included in this study. All patients received RFA therapy to increase the locoregional effects one week later [24]. They underwent ultrasound, CT scan or magnetic resonance imaging (MRI) of the abdomen about one month after treatment and at a minimum of once every three months thereafter, and tumor recurrences were followed for up to 360 days. The Institutional

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6 Review Board reviewed and approved the study protocol. This study complied with  
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8 ethical standards outlined in the Declaration of Helsinki. Adverse events were monitored  
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10 for one month after the DC infusion in terms of fever, vomiting, abdominal pain,  
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12 encephalopathy, myalgia, ascites, gastrointestinal disorder, bleeding, hepatic abscess  
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14 and autoimmune diseases.  
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### 18 **Preparation and injection of autologous DCs.**

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20 DCs were generated from blood monocyte precursors as previously reported  
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22 [25]. Briefly, peripheral blood mononuclear cells (PBMCs) were isolated by  
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24 centrifugation in Lymphoprep™ Tubes (Nycomed, Roskilde, Denmark). For generating  
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26 DCs, PBMCs were plated in 6-well tissue culture dishes (Costar, Cambridge, MA) at  
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28  $1.4 \times 10^7$  cells in 2-mL per well and allowed to adhere to plastic for 2 h. Adherent cells  
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30 were cultured in serum-free media (GMP CellGro® DC Medium; CellGro, Manassas,  
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32 VA) with 50 ng/ml recombinant human IL-4 (GMP grade; CellGro®), and 100 ng/ml  
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34 recombinant human granulocyte-macrophage colony-stimulating factor (GM-CSF)  
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36 (GMP grade; CellGro®) for 5 days to generate immature DC, and matured for a further 2  
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38 days in 0.1 KE/ml OK432 (Chugai Pharmaceuticals, Tokyo, Japan) to generate OK-DC.  
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40 On day 7, the cells were harvested for injection,  $5 \times 10^6$  cells were suspended in 5 ml  
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42 normal saline containing 1 % autologous plasma, mixed with absorbable gelatin sponge  
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44 (Gelfoam; Pharmacia & Upjohn, Peapack, NJ) and infused through an arterial catheter  
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46 following Lipiodol (iodized oil) (Lipiodol Ultrafluide, Laboratoire Guerbet,  
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48 Aulnay-Sous-Bois, France) injection during selective TAE therapy. Release criteria for  
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50 DCs were viability >80%, purity >30%, negative gram stain and endotoxin polymerase  
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52 chain reaction (PCR), and negative in process cultures from samples sent 48 h before  
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54 release. All products met all release criteria, and the DCs had a typical phenotype of  
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56  $CD14^-$  and human leukocyte antigen (HLA)-DR<sup>+</sup>.  
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### **Flow cytometry analysis.**

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6 The DC preparation was assessed by staining with the following monoclonal  
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9 chlorophyll protein (PerCP) (L243), anti-CCR7-phycoerythrin (PE) (3D12) (BD  
10 Pharmingen, San Diego, CA), anti-CD80-PE (MAB104), anti-CD83-PE (HB15a) and  
11 anti-CD86-PE (HA5.2B7) (Beckman Coulter, Fullerton, CA). Cells were analyzed on a  
12 FACSCalibur™ flow cytometer. Data analysis was performed with CELLQuest™  
13 software (Becton Dickinson, San Jose, CA).  
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#### 24 **DC phagocytosis.**

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26 Immature DCs and OK432-stimulated DCs were incubated with 1 mg/ml FITC  
27 dextran (Sigma-Aldrich, St. Louis, MO) for 30 min at 37°C and the cells were washed  
28 three times in FACS buffer before cell acquisition using a FACSCalibur™ cytometer.  
29 Control DCs (not incubated with FITC dextran) were acquired at the same time to allow  
30 background levels of fluorescence to be determined.  
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#### 38 **Enzyme-linked immunosorbent assay (ELISA).**

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40 DCs were seeded at 200,000 cells/ml, and supernatant collected after 48 h.  
41 IL-12p40 and IFN- $\gamma$  were detected using matched paired antibodies (BD Pharmingen)  
42 following standard protocols.  
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#### 48 **Cytotoxicity assays.**

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50 The ability of DCs to exert cytotoxicity was assessed in a standard <sup>51</sup>Cr release  
51 assay [19]. We used the HCC cell lines Hep3B and PLC/PRF/5 (ATCC, Manassas, VA)  
52 and a lymphoblastoid cell line T2 that expresses HLA-A\*0201 (ATCC) as target cells.  
53 Target cells were labeled with <sup>51</sup>Cr. In a 96-well plate, 2.5 x 10<sup>3</sup> target cells per well  
54 were incubated with DCs for 8 h at different effector / target (E / T) ratios in triplicates.  
55 Percent specific lysis was calculated as follows: (experimental release - spontaneous  
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6 release) / (maximum release - spontaneous release) x 100. Spontaneous release was  
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8 always < 20 % of the total.  
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### 10 11 12 **NK cell activity.**

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14 NK cell cytotoxicity against K562 erythroleukemia target cells was measured  
15 by using <sup>51</sup>Cr-release assay according to previously published methods [26] with  
16 PBMCs obtained from the patients. All experiments were performed in triplicate.  
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18 Percent cytotoxicity was calculated as follows: [(experimental cpm – spontaneous cpm)  
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20 / (total cpm – spontaneous cpm)] X 100.  
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### 26 27 **Intracellular cytokine expression.**

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29 Freshly isolated PBMCs were stimulated with 25 ng/ml phorbol 12-myristate  
30 13-acetate (PMA, Sigma-Aldrich) and 1 µg/ml ionomycin (Sigma-Aldrich) at 37 °C in  
31 humidified 7 % CO<sub>2</sub> for 4 h. To block cytokine secretion, brefeldin A (Sigma, St. Louis,  
32 MO) [27] was added to a final concentration of 10 µg/mL. After addition of stimuli, the  
33 surface staining was performed with anti-CD4-PC5 (13B8.2), anti-CD8-PerCP (SK1)  
34 and anti-CD56-PC5 (N901) (Beckman Coulter). Subsequently, the cells were  
35 permeabilized, stained for intracellular IFN-γ and IL-4 using the FastImmune™ system  
36 (BD Pharmingen), resuspended in PBS containing 1 % paraformaldehyde (PFA), and  
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38 analyzed on a flow cytometer (≈10,000 gated events acquired per sample).  
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### 48 49 **IFN-γ enzyme-linked immunospot (ELISPOT) assay.**

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51 ELISPOT assays were performed as previously described with the following  
52 modifications [28-30]. HLA-A24 restricted peptide epitopes, squamous cell carcinoma  
53 antigen recognized by T cells 2 (SART2)<sub>899</sub> (SYTRLFLIL), SART3<sub>109</sub>  
54 (VYDYNCHVDL), multidrug resistance protein 3 (MRP3)<sub>765</sub> (VYSDADIFL), MRP3<sub>503</sub>  
55 (LYAWEPSFL), MRP3<sub>692</sub> (AYVPQQAWI), alpha-fetoprotein (AFP)<sub>403</sub> (KYIQESQAL),  
56  
57 AFP<sub>434</sub> (AYTKKAPQL), AFP<sub>357</sub> (EYSRRHPQL), human telomerase reverse  
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6 transcriptase (hTERT)<sub>167</sub> (AYQVCGPPL) (unpublished), hTERT<sub>461</sub> (VYGFVRACL),  
7  
8 hTERT<sub>324</sub> (VYAETKHFL), were used in this study. Negative controls consisted of an  
9  
10 HIV envelope-derived peptide (HIVenv<sub>584</sub>). Positive controls consisted of 10 ng/ml  
11  
12 PMA (Sigma) or a CMV pp65-derived peptide (CMVpp65<sub>328</sub>). The colored spots were  
13  
14 counted with a KS ELISpot Reader (Zeiss, Tokyo, Japan). The number of specific spots  
15  
16 was determined by subtracting the number of spots in the absence of antigen from the  
17  
18 number of spots in its presence. Responses were considered positive if more than 10  
19  
20 specific spots were detected and if the number of spots in the presence of antigen was at  
21  
22 least two fold greater than the number of spots in the absence of antigen.  
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### 26 **Cytokine and chemokine profiling.**

27  
28 Serum cytokine and chemokine levels were measured using the Bioplex assay  
29  
30 (Bio-Rad, Hercules, CA). Briefly, frozen serum samples were thawed at room  
31  
32 temperature, diluted 1 : 4 in sample diluents, and 50  $\mu$ l aliquots of diluted sample were  
33  
34 added in duplicate to the wells of 96-well microtiter plate containing the coated beads  
35  
36 for a validated panel of 27 human cytokines and chemokines (cytokine 27-plex antibody  
37  
38 bead kit) according to the manufacturer's instructions. These included IL-1 $\beta$ , IL-1Ra,  
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40 IL-2, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12p70, IL-13, IL-15, IL-17, basic FGF,  
41  
42 eotaxin, G-CSF, GM-CSF, IFN- $\gamma$ , IP-10, MCP-1, MIP-1 $\alpha$ , MIP-1 $\beta$ , PDGF-BB,  
43  
44 RANTES, TNF- $\alpha$ , and VEGF. Eight standards (ranging from 2 to 32,000 pg/mL) were  
45  
46 used to generate calibration curves for each cytokine. Data acquisition and analysis were  
47  
48 done using Bio-Plex Manager software version 4.1.1.  
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### 52 **Arginase activity.**

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54 Serum samples were tested for arginase activity by conversion of L-arginine to  
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56 L-ornithine [31] using a kit supplied the manufacturer (BioAssay Systems, Hayward,  
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58 CA). Briefly, sera were treated with a membrane filter (Millipore, Billerica, MA) to  
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60 remove urea, combined with the sample buffer in wells of 96-well plate, and incubated

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6 at 37°C for 2 h. Subsequently, the urea reagent was added to stop the arginase reaction.  
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8 The color produced was read at 520 nm using a microtiter plate reader.  
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### 10 11 **Statistical analysis.**

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14 Results are expressed as means  $\pm$  SD. Differences between groups were  
15 analyzed for statistical significance by the Mann-Whitney U test. Qualitative variables  
16 were compared by means of Fisher's exact test. The estimated probability of tumor  
17 recurrence-free survival was determined using the Kaplan-Meier method. The Mantel  
18 Cox log-rank test was used to compare curves between groups. Any *P* values less than  
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0.05 were considered statistically significant. All statistical tests were two-sided.

## RESULTS

### Preparation of OK432-stimulated DCs.

Adherent cells isolated from PBMCs of patients with cirrhosis and HCC (**Table 1**) were differentiated into DCs in the presence of IL-4 and GM-CSF. The cells were stimulated with 0.1 KE/ml OK432 for three days.  $54.6 \pm 9.5$  % (mean  $\pm$  SD; n = 13) of OK432-stimulated cells showed high levels of MHC class II (HLA-DR) and the absence of lineage markers including CD3, CD14, CD16, CD19, CD20 and CD56, in which  $30.9 \pm 14.2$  % were CD11c-positive (myeloid DC subset) and  $14.8 \pm 11.2$  were CD123-positive (plasmacytoid DC subset), consistent with our previous observations [20]. As reported [32,33], greater proportions of the cells developed high levels of expression of the costimulatory molecules B7-1 (CD80) and B7-2 (CD86) and an activation marker (CD83) compared to DCs prepared without OK432 stimulation (**Fig. 1A**). Furthermore, the chemokine receptor CCR7 which leads to homing to lymph nodes [13,34] was also induced following OK432 stimulation.

To evaluate the endocytic and phagocytic ability of the OK432-stimulated cells, uptake of FITC-dextran was quantitated by flow cytometry (**Fig. 1B**). The cells showed lower levels of uptake due to maturation compared to DCs prepared without OK432 stimulation, while the OK432-stimulated cells derived from HCC patients preserved a moderate uptake capacity. As expected, the OK432-stimulated cells produced large amounts of the cytokines IL-12 and IFN- $\gamma$  (**Fig. 1C**). In addition, they displayed high cytotoxic activity against HCC cell lines (Hep3B and PLC/PRF/5) and a lymphoblastoid cell line (T2) although DCs without OK432 stimulation lysed none of the target cells to any great degree (**Fig. 1D**). Taken together, these results demonstrate that OK432 stimulation of IL-4 and GM-CSF-induced immature DCs derived from HCC patients promoted their maturation towards cells with activated phenotypes, high expression of a homing receptor, fairly well-preserved phagocytic capacity, greatly enhanced cytokine



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6 production and effective tumoricidal activity, consistent with previous observations  
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8 [16,19].  
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### 10 11 **Safety of OK432-stimulated DC administration.** 12

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14 Prior to the administration of OK432-stimulated DCs to patients, the cells were  
15 confirmed to be safe in athymic nude mice to which 100-fold cell numbers / weight  
16 were injected subcutaneously (data not shown). Subsequently, OK432-stimulated DC  
17 administration was performed during TAE therapy in humans, in which DCs were  
18 mixed together with absorbable gelatin sponge (Gelfoam) and infused through an  
19 arterial catheter following iodized oil (Lipiodol) injection as reported previously [20].  
20 Adverse events were monitored clinically and biochemically after DC infusion (**Table**  
21 **2**). A larger proportion (12 / 13) of the patients were complicated with high fever  
22 compared to those previously treated with immature DCs (5 / 10) [20], probably due to  
23 the proinflammatory responses induced by OK432-stimulated DCs. However, there  
24 were no grade III or IV National Cancer Institute Common Toxicity Criteria adverse  
25 events including vomiting, abdominal pain, encephalopathy, myalgia, ascites,  
26 gastrointestinal disorders, bleeding, hepatic abscess or autoimmune diseases associated  
27 with DC infusion and TAE in this study. There was also no clinical or serological  
28 evidence of hepatic failure or autoimmune response in any patients. Thus, concurrent  
29 treatment with OK432-stimulated DC infusions can be performed safely at the same  
30 time as TAE in patients with cirrhosis and HCC.  
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### 50 **Recurrence-free survival following DC infusion.** 51

52 A further objective of this study was to determine clinical response following  
53 DC infusion. A group of historical controls treated with TAE without DC administration  
54 was reviewed for this study (**Table 3**). The clinical characteristics including tumor  
55 burden and hepatic reserve were comparable between patients treated with TAE and  
56 OK432-stimulated DC transfer (n = 13) and those historical controls with TAE but  
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6 without DC administration (n = 22). We compared the recurrence-free survival between  
7 these patient groups. Kaplan-Meier analysis indicated that patients treated with TAE and  
8 OK432-stimulated DC transfer had prolonged recurrence-free survival compared with  
9 the historical controls that had been treated with TAE alone (recurrence rates 360 days  
10 after the treatments; 2 / 13 and 12 / 22, respectively;  $P = 0.046$ , log-rank test) (**Fig. 2**).  
11 The results demonstrated that OK432-stimulated DC transfer during TAE therapy  
12 reduces tumor recurrence in HCC patients.  
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### 22 **NK cell activity and intracellular cytokine responses in PBMCs.**

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24 To assess systemic immunomodulatory effects of OK432-stimulated DC  
25 transfer, PBMCs were isolated one and three months after treatment and NK cell  
26 cytotoxicity against K562 erythroleukemia target cells measured using the  $^{51}\text{Cr}$ -release  
27 assay (**Fig. 3**). The level of NK cell was unaltered following treatment. In addition,  
28 cytokine production capacity of lymphocyte subsets was quantitated by measuring  
29 intracellular IFN- $\gamma$  and IL-4 using flow cytometry. There were also no significant  
30 changes in terms of cytokine production capacity in the CD4 $^{+}$ , CD8 $^{+}$  and CD56 $^{+}$  subsets  
31 in the patients treated with OK432-stimulated DCs.  
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### 42 **Immune responses to peptide epitopes derived from tumor antigens.**

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44 To assess the effects on T cell responses to tumor antigens, PBMCs were  
45 obtained 4 weeks after DC infusion, pulsed with peptides derived from AFP, MRP3,  
46 SART2, SART3 and hTERT. IFN- $\gamma$  production was then quantitated in an ELISPOT  
47 assay. Cells producing IFN- $\gamma$  in response to stimulation with HLA-A24 [the most  
48 common HLA-A antigen (58.1%) in Japanese populations [35]]-restricted peptide  
49 epitopes derived from tumor antigens MRP3 and hTERT were induced in 3 of 6  
50 HLA-A24-positive patients (numbers 2, 6 and 11) after treatment with TAE and  
51 OK432-stimulated DCs (**Fig. 4**). To understand the immunological and clinical  
52 significance of the T lymphocyte responses, PBMCs obtained from the historical control  
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6 patients who had been treated with TAE without DC administration were also evaluated  
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8 by ELISPOT. Similarly, positive reactions were observed in 4 (numbers t8, t19, t20 and  
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10 t22) of 6 HLA-A24-positive patients. These data indicate that T lymphocyte responses  
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12 to HLA-A24 restricted peptide epitopes of tumor antigens were induced following the  
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14 TAE therapy, but no additional responses were observed as a result of  
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16 OK432-stimulated DC transfer in the current study.  
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### 20 **Serum levels of cytokines, chemokines and arginase activity.**

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22 To screen for immunobiological responses induced following  
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24 OK432-stimulated DC transfer, serum levels of cytokines and chemokines were  
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26 measured simultaneously using the Bio-Plex multiplex suspension array system. The  
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28 results were compared with the historical control patients treated with TAE without DC  
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30 administration. Interestingly, serum concentrations of IL-9, IL-15 and TNF- $\alpha$  were  
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32 greatly increased after OK432-stimulated DC infusion, in contrast to their reduction  
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34 following TAE treatment alone (**Fig. 5A**). Furthermore, the chemokines eotaxin  
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36 (CCL11) and MIP-1 $\beta$  (CCL4) were markedly induced after DC transfer although they  
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38 were also decreased after TAE alone. These data indicate that transfer of  
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40 OK432-stimulated DC during TAE therapy induced unique immune responses that may  
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42 be mediated by the cytokines IL-9, IL-15 and TNF- $\alpha$  and the chemokines eotaxin and  
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44 MIP-1 $\beta$ .  
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51 In addition, serum arginase activity was reported to reflect numbers of  
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53 myeloid-derived suppressor cells (MDSCs) that may inhibit T lymphocyte responses in  
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55 cancer patients [36]. Therefore, serum arginase activity was measured after  
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57 OK432-stimulated DC infusion, and it was found that it was increased 6- or 7-fold in  
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59 patients treated with TAE. However, this increase was independent of the presence or  
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activity was decreased again 4 weeks after treatment with both TAE and

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6 OK432-stimulated DC transfer but tended to be maintained at a high level in patients  
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8 treated with TAE without DC transfer. However, these differences did not reach  
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10 statistical significance ( $P > 0.05$ ). Because arginase activity is known to be relatively  
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12 high in liver and HCC cells [37], the influence of tissue injury was assessed  
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14 biochemically by measuring serum levels of alanine aminotransferase (ALT) and lactic  
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16 dehydrogenase (LDH) activities. We did not observed ALT or LDH elevation, indicating  
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18 that the increase of arginase activity was not due to tissue damage following treatment.  
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20 Collectively, these results demonstrate that infusion of OK432-stimulated DCs during  
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22 TAE treatment may reduce the immunosuppressive activities of MDSCs, and assist in  
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24 developing a favorable environment for the induction of antitumor immunity.  
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## DISCUSSION

Although many novel strategies including immunotherapies have been developed in an attempt to suppress tumor recurrence after curative treatments for HCC, recurrence rates and survival times have not been improved significantly [38]. In the current study, we first established that OK432-stimulated DC administration during TAE therapy did not cause critical adverse events in patients with cirrhosis and HCC. Most importantly, DC transfer resulted in prolonged recurrence-free survival after combination therapy with TAE and OK432-stimulated DC administration. In terms of the immunomodulatory effects of DC transfer, although NK cell activity, intracellular cytokine production, and T lymphocyte-mediated immune responses were not altered in PBMCs from treated patients, serum levels of IL-9, IL-15 and TNF- $\alpha$  and the chemokines eotaxin and MIP-1 $\beta$  were markedly enhanced after DC transfer. In addition, serum levels of arginase activity were decreased following DC transfer. Collectively, this study demonstrated the feasibility, safety, and beneficial antitumor effects of OK432-stimulated DC infusion into tumor tissues for patients with cirrhosis and HCC, suggesting the ability of an active immunotherapeutic strategy to reduce tumor recurrence after locoregional treatment of HCC.

DCs were stimulated with OK432 prior to infusion into tumor tissues through an arterial catheter. OK432 was reported to activate DCs through its binding to TLR-2 and -4 [16,39] that can be used for cancer therapy [33]. The current results indicate that OK432 stimulation of immature DCs from HCC patients promoted their maturation processes while preserving antigen uptake capacity and enhancing tumoricidal activity, consistent with previous observations [16,19] and supporting the current strategy in which OK432-stimulated DCs were directly infused into tumor tissues. Since the tumoricidal activity of unstimulated DCs was not observed in in vitro experiments, OK432 stimulation obviously altered the cytotoxic properties of DCs. One of the

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6 mechanisms of DC killing was reported to be CD40 / CD40 ligand interaction [19]. The  
7 further studies are needed to determine the killing mechanisms of DCs derived from  
8 HCC patients in a direct (TNF, TRAIL, Fas ligand, NO and perforin / granzyme) and  
9 indirect (MHC-restricted) manner [40-43]. Although the main mechanism by which  
10 OK432-stimulated DCs prolonged the recurrence-free survival was not elucidated, the  
11 tumoricidal activity of mature DCs was implicated in in vivo enhancement of antigen  
12 presentation, costimulation and inflammatory cytokine production.  
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22 Very recent reports document injection of OK432-stimulated DCs into patients  
23 with cancer of the gastrointestinal tract or pancreas [44,45], but their antitumor effects  
24 were not clearly defined. The current study shows for the first time that  
25 OK432-stimulated DCs induce beneficial antitumor responses when transferred into  
26 tumor tissues during TAE therapy. The antitumor responses may have been enhanced as  
27 a result of optimal activation of the DCs with OK432 or combining infusion of  
28 stimulated DCs with TAE therapy. Inappropriately activated DCs may be unable to  
29 generate sufficient numbers of properly activated effector T lymphocytes [46]. As  
30 shown in **Fig. 1**, all of these alterations could contribute to the further enhancement of  
31 antitumor effects compared to those in our previous study with immature DCs [20].  
32 Furthermore, the tumor cell death-promoting therapies, e.g. chemotherapy [47] and TAE  
33 [48], can be expected to enhance the effects of therapeutic cancer vaccines by redressing  
34 the immunosuppressive tumor environment.  
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50 NK cell activity and intracellular cytokine responses in CD4<sup>+</sup> and CD8<sup>+</sup> T  
51 lymphocytes and CD56<sup>+</sup> NK cell subsets in PBMCs were not significantly changed in  
52 patients treated with OK432-stimulated DCs. Furthermore, we did not observe tumor  
53 antigen-specific T lymphocyte responses clearly associated with DC administration. The  
54 data therefore suggest that the immune responses induced by the therapy applied here  
55 were not systemically detectable. Because cytotoxic T lymphocyte responses were  
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6 enhanced in patients receiving  $> 3 \times 10^7$  cells [49,50], the numbers of transferred  
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8 OK432-stimulated DCs were apparently not sufficient to induce responses detectable in  
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10 the peripheral blood, but were enough to exert beneficial antitumor effects. In addition,  
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12 many studies have concluded that cytotoxic T lymphocyte responses rarely predict  
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14 clinical outcomes of DC-based immunotherapies [51,52], and that in many cases, also  
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16 including our own studies [28,30], tumor-specific effector T lymphocytes coexist with  
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18 the tumors. Consistent with these observations, the current results suggest that cytotoxic  
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20 T lymphocyte responses in PBMCs are not reliable predictors of beneficial antitumor  
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22 effects in patients treated with the current OK432-stimulated DC strategy.  
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26 Serum levels of the cytokines IL-9, IL-15 and TNF- $\alpha$  and the chemokines  
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28 eotaxin and MIP-1 $\beta$  were increased following OK432-stimulated DC transfer, but  
29  
30 decreased after TAE therapy without DC administration. IL-9 and IL-15 belong to the  
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32 cytokine receptor common gamma chain ( $\gamma_c$ ; CD132) family, a member of the Type I  
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34 cytokine receptor family expressed on most lymphocyte populations [53]. IL-9 exerts  
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36 pleiotropic activities on T and B lymphocytes, mast cells, monocytes and hematopoietic  
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38 progenitors [54,55]. IL-15 and TNF- $\alpha$  are known to prime T lymphocytes and NK cells  
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40 when secreted by DCs [56] and to induce antitumor immune responses [57]. Eotaxin is  
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42 known to selectively recruit eosinophils also contributing to antitumor effects [58,59],  
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44 and MIP-1 $\beta$  is a chemoattractant for NK cells, monocytes and a variety of other  
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46 immune cells [60]. In addition, serum levels of arginase tended to decrease after DC  
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48 transfer. Because serum arginase activity reflects the numbers of MDSCs that inhibit T  
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50 lymphocyte responses in cancer patients [36], the patients treated with  
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52 OK432-stimulated DCs might have developed lower levels of suppressor cells.  
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54 Collectively, the results suggest that infusion of OK432-stimulated DCs may orchestrate  
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56 the immune environment in the whole body that could enhance beneficial antitumor  
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58 effects, although the precise molecular and cellular mechanisms associated with the  
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60 actions of these cytokines and chemokines were not clearly defined in the current

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analysis.

For Peer Review



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**DISCLOSURE**

The authors have declared that no conflict of interest exists.

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## REFERENCES

1. Omata M, Tateishi R, Yoshida H, Shiina S. Treatment of hepatocellular carcinoma by percutaneous tumor ablation methods: Ethanol injection therapy and radiofrequency ablation. *Gastroenterology* 2004; 127:S159-66.
2. Belghiti J. Resection and liver transplantation for HCC. *J Gastroenterol* 2009; 44 Suppl 19:132-5.
3. Nakamoto Y, Guidotti LG, Kuhlen CV, Fowler P, Chisari FV. Immune pathogenesis of hepatocellular carcinoma. *J Exp Med* 1998; 188:341-50.
4. Ercolani G, Grazi GL, Ravaioli M, et al. Liver resection for hepatocellular carcinoma on cirrhosis: univariate and multivariate analysis of risk factors for intrahepatic recurrence. *Ann Surg* 2003; 237:536-43.
5. Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ, Schreiber RD. IFN $\gamma$  and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature* 2001; 410:1107-11.
6. Vulink A, Radford KJ, Melief C, Hart DN. Dendritic cells in cancer immunotherapy. *Adv Cancer Res* 2008; 99:363-407.
7. Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu YJ, Pulendran B, Palucka K. Immunobiology of dendritic cells. *Annu Rev Immunol* 2000; 18:767-811.
8. Lemos MP, Esquivel F, Scott P, Laufer TM. MHC class II expression restricted to CD8 $\alpha$ <sup>+</sup> and CD11b<sup>+</sup> dendritic cells is sufficient for control of *Leishmania major*. *J Exp Med* 2004; 199:725-30.
9. Ni K, O'Neill HC. The role of dendritic cells in T cell activation. *Immunol Cell Biol* 1997; 75:223-30.
10. Andrews DM, Andoniou CE, Scalzo AA, van Dommelen SL, Wallace ME, Smyth MJ, Degli-Esposti MA. Cross-talk between dendritic cells and natural killer cells in

- 1  
2  
3  
4  
5  
6  
7 viral infection. *Mol Immunol* 2005; 42:547-55.
- 8  
9 11. Heiser A, Coleman D, Dannull J, et al. Autologous dendritic cells transfected with  
10 prostate-specific antigen RNA stimulate CTL responses against metastatic prostate  
11 tumors. *J Clin Invest* 2002; 109:409-17.
- 12  
13 12. Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature*  
14 1998; 392:245-52.
- 15  
16 13. Forster R, Schubel A, Breitfeld D, Kremmer E, Renner-Muller I, Wolf E, Lipp M.  
17 CCR7 coordinates the primary immune response by establishing functional  
18 microenvironments in secondary lymphoid organs. *Cell* 1999; 99:23-33.
- 19  
20 14. MartIn-Fontecha A, Sebastiani S, Hopken UE, Ugucioni M, Lipp M,  
21 Lanzavecchia A, Sallusto F. Regulation of dendritic cell migration to the draining  
22 lymph node: impact on T lymphocyte traffic and priming. *J Exp Med* 2003;  
23 198:615-21.
- 24  
25 15. Ratzinger G, Stoitzner P, Ebner S, et al. Matrix metalloproteinases 9 and 2 are  
26 necessary for the migration of Langerhans cells and dermal dendritic cells from  
27 human and murine skin. *J Immunol* 2002; 168:4361-71.
- 28  
29 16. Nakahara S, Tsunoda T, Baba T, Asabe S, Tahara H. Dendritic cells stimulated with  
30 a bacterial product, OK-432, efficiently induce cytotoxic T lymphocytes specific to  
31 tumor rejection peptide. *Cancer Res* 2003; 63:4112-8.
- 32  
33 17. Okamoto M, Oshikawa T, Tano T, et al. Mechanism of anticancer host response  
34 induced by OK-432, a streptococcal preparation, mediated by phagocytosis and  
35 Toll-like receptor 4 signaling. *J Immunother* 2006; 29:78-86.
- 36  
37 18. Pasare C, Medzhitov R. Toll pathway-dependent blockade of CD4+CD25+ T  
38 cell-mediated suppression by dendritic cells. *Science* 2003; 299:1033-6.
- 39  
40 19. Hill KS, Errington F, Steele LP, et al. OK432-activated human dendritic cells kill  
41 tumor cells via CD40/CD40 ligand interactions. *J Immunol* 2008; 181:3108-15.
- 42  
43 20. Nakamoto Y, Mizukoshi E, Tsuji H, et al. Combined therapy of transcatheter  
44  
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4  
5  
6  
7 hepatic arterial embolization with intratumoral dendritic cell infusion for  
8  
9 hepatocellular carcinoma: clinical safety. *Clin Exp Immunol* 2007; 147:296-305.
- 10  
11 21. Steinman RM, Banchereau J. Taking dendritic cells into medicine. *Nature* 2007;  
12  
13 449:419-26.
- 14  
15 22. Tacken PJ, de Vries IJ, Torensma R, Figdor CG. Dendritic-cell immunotherapy:  
16  
17 from ex vivo loading to in vivo targeting. *Nat Rev Immunol* 2007; 7:790-802.
- 18  
19 23. Makuuchi M. General rules for the clinical and pathological study of primary liver  
20  
21 cancer, 2nd English ed. Tokyo, Japan: Kanehara & Co., Ltd., 2003.
- 22  
23 24. Veltri A, Moretto P, Doriguzzi A, Pagano E, Carrara G, Gandini G. Radiofrequency  
24  
25 thermal ablation (RFA) after transarterial chemoembolization (TACE) as a  
26  
27 combined therapy for unresectable non-early hepatocellular carcinoma (HCC). *Eur*  
28  
29 *Radiol* 2006; 16:661-9.
- 30  
31 25. Dhodapkar MV, Steinman RM, Sapp M, et al. Rapid generation of broad T-cell  
32  
33 immunity in humans after a single injection of mature dendritic cells. *J Clin Invest*  
34  
35 1999; 104:173-80.
- 36  
37 26. Orange JS, Brodeur SR, Jain A, et al. Deficient natural killer cell cytotoxicity in  
38  
39 patients with IKK-gamma/NEMO mutations. *J Clin Invest* 2002; 109:1501-9.
- 40  
41 27. Klausner RD, Donaldson JG, Lippincott-Schwartz J. Brefeldin A: insights into the  
42  
43 control of membrane traffic and organelle structure. *J Cell Biol* 1992; 116:1071-80.
- 44  
45 28. Mizukoshi E, Nakamoto Y, Marukawa Y, et al. Cytotoxic T cell responses to human  
46  
47 telomerase reverse transcriptase in patients with hepatocellular carcinoma.  
48  
49 *Hepatology* 2006; 43:1284-94.
- 50  
51 29. Mizukoshi E, Nakamoto Y, Tsuji H, Yamashita T, Kaneko S. Identification of  
52  
53 alpha-fetoprotein-derived peptides recognized by cytotoxic T lymphocytes in  
54  
55 HLA-A24+ patients with hepatocellular carcinoma. *Int J Cancer* 2006;  
56  
57 118:1194-204.
- 58  
59 30. Mizukoshi E, Honda M, Arai K, Yamashita T, Nakamoto Y, Kaneko S. Expression  
60

- 1  
2  
3  
4  
5  
6  
7 of multidrug resistance-associated protein 3 and cytotoxic T cell responses in  
8 patients with hepatocellular carcinoma. *J Hepatol* 2008; 49:946-54.
- 9  
10  
11 31. Rodriguez PC, Quiceno DG, Zabaleta J, et al. Arginase I production in the tumor  
12 microenvironment by mature myeloid cells inhibits T-cell receptor expression and  
13 antigen-specific T-cell responses. *Cancer Res* 2004; 64:5839-49.
- 14  
15  
16  
17 32. Itoh T, Ueda Y, Okugawa K, et al. Streptococcal preparation OK432 promotes  
18 functional maturation of human monocyte-derived dendritic cells. *Cancer Immunol*  
19 *Immunother* 2003; 52:207-14.
- 20  
21  
22  
23 33. Kuroki H, Morisaki T, Matsumoto K, Onishi H, Baba E, Tanaka M, Katano M.  
24 Streptococcal preparation OK-432: a new maturation factor of monocyte-derived  
25 dendritic cells for clinical use. *Cancer Immunol Immunother* 2003; 52:561-8.
- 26  
27  
28  
29 34. Gunn MD, Kyuwa S, Tam C, Kakiuchi T, Matsuzawa A, Williams LT, Nakano H.  
30 Mice lacking expression of secondary lymphoid organ chemokine have defects in  
31 lymphocyte homing and dendritic cell localization. *J Exp Med* 1999; 189:451-60.
- 32  
33  
34  
35 35. Imanishi T, Akaza T, Kimura A, Tokunaga K, Gojobori T. HLA 1991, Proceeding of  
36 the Eleventh International Histocompatibility Workshop and Conference. Tokyo,  
37 Japan: Oxford University Press, 1992.
- 38  
39  
40  
41 36. Zea AH, Rodriguez PC, Atkins MB, et al. Arginase-producing myeloid suppressor  
42 cells in renal cell carcinoma patients: a mechanism of tumor evasion. *Cancer Res*  
43 2005; 65:3044-8.
- 44  
45  
46  
47 37. Chrzanowska A, Krawczyk M, Baranczyk-Kuzma A. Changes in arginase  
48 isoenzymes pattern in human hepatocellular carcinoma. *Biochem Biophys Res*  
49 *Commun* 2008; 377:337-40.
- 50  
51  
52  
53 38. Caldwell S, Park SH. The epidemiology of hepatocellular cancer: from the  
54 perspectives of public health problem to tumor biology. *J Gastroenterol* 2009; 44  
55 *Suppl* 19:96-101.
- 56  
57  
58  
59 39. Okamoto M, Oshikawa T, Tano T, et al. Involvement of Toll-like receptor 4  
60

- 1  
2  
3  
4  
5  
6  
7 signaling in interferon-gamma production and antitumor effect by streptococcal  
8 agent OK-432. *J Natl Cancer Inst* 2003; 95:316-26.
- 9  
10  
11 40. Liu S, Yu Y, Zhang M, Wang W, Cao X. The involvement of TNF-alpha-related  
12 apoptosis-inducing ligand in the enhanced cytotoxicity of IFN-beta-stimulated  
13 human dendritic cells to tumor cells. *J Immunol* 2001; 166:5407-15.
- 14  
15  
16  
17 41. Lu G, Janjic BM, Janjic J, Whiteside TL, Storkus WJ, Vujanovic NL. Innate direct  
18 anticancer effector function of human immature dendritic cells. II. Role of TNF,  
19 lymphotoxin-alpha(1)beta(2), Fas ligand, and TNF-related apoptosis-inducing  
20 ligand. *J Immunol* 2002; 168:1831-9.
- 21  
22  
23  
24  
25 42. Nicolas A, Cathelin D, Larmonier N, et al. Dendritic cells trigger tumor cell death  
26 by a nitric oxide-dependent mechanism. *J Immunol* 2007; 179:812-8.
- 27  
28  
29 43. Stry G, Bangert C, Tauber M, Strohal R, Kopp T, Stingl G. Tumoricidal activity of  
30 TLR7/8-activated inflammatory dendritic cells. *J Exp Med* 2007; 204:1441-51.
- 31  
32  
33 44. West E, Morgan R, Scott K, et al. Clinical grade OK432-activated dendritic cells: in  
34 vitro characterization and tracking during intralymphatic delivery. *J Immunother*  
35 2009; 32:66-78.
- 36  
37  
38  
39 45. Hirooka Y, Itoh A, Kawashima H, et al. A combination therapy of gemcitabine with  
40 immunotherapy for patients with inoperable locally advanced pancreatic cancer.  
41 *Pancreas* 2009; 38:e69-74.
- 42  
43  
44  
45 46. Melief CJ. Cancer immunotherapy by dendritic cells. *Immunity* 2008; 29:372-83.
- 46  
47  
48 47. Zitvogel L, Apetoh L, Ghiringhelli F, Kroemer G. Immunological aspects of cancer  
49 chemotherapy. *Nat Rev Immunol* 2008; 8:59-73.
- 50  
51  
52 48. Ayaru L, Pereira SP, Alisa A, et al. Unmasking of alpha-fetoprotein-specific CD4(+)  
53 T cell responses in hepatocellular carcinoma patients undergoing embolization. *J*  
54 *Immunol* 2007; 178:1914-22.
- 55  
56  
57  
58 49. Thurner B, Haendle I, Roder C, et al. Vaccination with mage-3A1 peptide-pulsed  
59 mature, monocyte-derived dendritic cells expands specific cytotoxic T cells and  
60

- 1  
2  
3  
4  
5  
6 induces regression of some metastases in advanced stage IV melanoma. *J Exp Med*  
7 1999; 190:1669-78.  
8  
9
- 10 50. Banchereau J, Palucka AK, Dhodapkar M, et al. Immune and clinical responses in  
11 patients with metastatic melanoma to CD34(+) progenitor-derived dendritic cell  
12 vaccine. *Cancer Res* 2001; 61:6451-8.  
13  
14  
15 51. Engell-Noerregaard L, Hansen TH, Andersen MH, Thor Straten P, Svane IM.  
16 Review of clinical studies on dendritic cell-based vaccination of patients with  
17 malignant melanoma: assessment of correlation between clinical response and  
18 vaccine parameters. *Cancer Immunol Immunother* 2009; 58:1-14.  
19  
20  
21 52. Itoh K, Yamada A, Mine T, Noguchi M. Recent advances in cancer vaccines: an  
22 overview. *Jpn J Clin Oncol* 2009; 39:73-80.  
23  
24  
25 53. Sugamura K, Asao H, Kondo M, Tanaka N, Ishii N, Nakamura M, Takeshita T. The  
26 common gamma-chain for multiple cytokine receptors. *Adv Immunol* 1995;  
27 59:225-77.  
28  
29  
30 54. Temann UA, Geba GP, Rankin JA, Flavell RA. Expression of interleukin 9 in the  
31 lungs of transgenic mice causes airway inflammation, mast cell hyperplasia, and  
32 bronchial hyperresponsiveness. *J Exp Med* 1998; 188:1307-20.  
33  
34  
35 55. McMillan SJ, Bishop B, Townsend MJ, McKenzie AN, Lloyd CM. The absence of  
36 interleukin 9 does not affect the development of allergen-induced pulmonary  
37 inflammation nor airway hyperreactivity. *J Exp Med* 2002; 195:51-7.  
38  
39  
40 56. de Saint-Vis B, Fugier-Vivier I, Massacrier C, et al. The cytokine profile expressed  
41 by human dendritic cells is dependent on cell subtype and mode of activation. *J*  
42 *Immunol* 1998; 160:1666-76.  
43  
44  
45 57. Shanmugham LN, Petrarca C, Frydas S, et al. IL-15 an immunoregulatory and  
46 anti-cancer cytokine. Recent advances. *J Exp Clin Cancer Res* 2006; 25:529-36.  
47  
48  
49 58. Kataoka S, Konishi Y, Nishio Y, Fujikawa-Adachi K, Tominaga A. Antitumor  
50 activity of eosinophils activated by IL-5 and eotaxin against hepatocellular  
51  
52  
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57  
58  
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60



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7 carcinoma. *DNA Cell Biol* 2004; 23:549-60.

- 8  
9 59. Simson L, Ellyard JI, Dent LA, Matthaai KI, Rothenberg ME, Foster PS, Smyth MJ,  
10 Parish CR. Regulation of carcinogenesis by IL-5 and CCL11: a potential role for  
11 eosinophils in tumor immune surveillance. *J Immunol* 2007; 178:4222-9.  
12  
13 60. Bystry RS, Aluvihare V, Welch KA, Kallikourdis M, Betz AG. B cells and  
14 professional APCs recruit regulatory T cells via CCL4. *Nat Immunol* 2001;  
15 2:1126-32.  
16  
17  
18  
19  
20  
21  
22  
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For Peer Review

## FIGURE LEGENDS

**Figure 1.** Effects of OK432 stimulation on the properties of dendritic cells (DCs) generated from blood monocyte precursors in patients with cirrhosis and hepatocellular carcinoma (HCC) (n = 13). **(A)** lin<sup>+</sup>HLA-DR<sup>-</sup> subsets with [OK432(+)] and without [OK432(-)] stimulation were analyzed for surface expression of CD80, CD83, CD86 and CCR7. Dot plots of a representative case are shown in the left-hand panel. Mean percentages ( $\pm$  SD) of positive cells are indicated in the right-hand panel. OK432 stimulation resulted in the expression of high levels of CD80, CD83, CD86 and CCR7 in the lin<sup>+</sup>HLA-DR<sup>-</sup> DC subset. **(B)** DC subsets with and without OK432 stimulation were incubated with fluorescein isothiocyanate (FITC) dextran for 30 min and the uptake was determined by flow cytometry. A representative analysis is shown in the upper panel. Mean fluorescence intensities (MFIs) ( $\pm$  SD) of the positive cells are indicated in the lower panel. OK432-stimulated cells showed lower levels of uptake due to maturation. **(C)** DC supernatants were harvested and the concentrations of IL-12 and IFN- $\gamma$  measured by ELISA. OK432-stimulated cells produced large amounts of the cytokines. The data indicate means  $\pm$  SD of the groups with and without the stimulation. All comparisons in A - C [OK432(+) vs. OK432(-)] were statistically significant by the Mann-Whitney U test ( $P < 0.005$ ). **(D)** Tumoricidal activity of DCs assessed by incubation with <sup>51</sup>Cr-labeled Hep3B, PLC/PRF/5 and T2 targets for 8 h at the indicated E / T (effector / target) cell ratios. OK432-stimulated cells displayed high cytotoxic activity against the target cells. The results are representative of the cases studied.

**Figure 2.** Recurrence-free survival of patients treated with transcatheter hepatic arterial embolization (TAE) with (TAE + OK-DC; n = 13) and without (TAE: historical controls; n = 22) OK432-stimulated DC administration. Time zero is the date of TAE. All patients underwent ultrasound, CT scan or MRI of the abdomen about 1 month after treatment and at a minimum of once every 3 months thereafter. Kaplan-Meier analysis

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6 indicated that TAE + OK-DC treatment prolonged recurrence-free survival compared  
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8 with the TAE-alone group (recurrence rates 360 days after the treatments; 2 / 13 and 12  
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10 / 22, respectively;  $P = 0.046$ , log-rank test).  
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14 **Figure 3.** Natural killer (NK) cell activity and intracellular cytokine production in  
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16 PBMCs of patients treated with OK432-stimulated DCs during TAE therapy (n = 13).  
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18 PBMCs were isolated before and one and three months after treatment and used for the  
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20 analyses. Upper panel, NK cell cytotoxicity against K562 erythroleukemia target cells  
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22 was evaluated at the E / T cell ratios shown. NK cell activities were not changed  
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24 following treatment. Middle and lower panels, PBMCs were stimulated with phorbol  
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26 12-myristate 13-acetate (PMA) and ionomycin, stained for CD4, CD8 and CD56  
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28 expression, permeabilized, and stained for intracellular IFN- $\gamma$  and IL-4. Percentages of  
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30 cytokine-positive cells were quantitated by flow cytometry. There were no significant  
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32 changes in terms of cytokine production capacity in the CD4<sup>+</sup>, CD8<sup>+</sup> and CD56<sup>+</sup> subsets  
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34 following the treatments. The data are given as means  $\pm$  SD of the groups.  
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39 **Figure 4.** Immune responses to HLA-A24-restricted peptide epitopes derived from  
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41 tumor antigens in HLA-A24-positive patients treated with OK432-stimulated DCs  
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43 during TAE therapy (numbers 2, 5, 6, 7, 10 and 11) and HLA-A24-positive historical  
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45 controls treated with TAE without DC transfer (numbers t8, t14, t15, t19, t20 and t22).  
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47 PBMCs were obtained before (open bars) and one month after the infusion (solid bars),  
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49 pulsed with the peptides derived from SART2, SART3, MRP3, AFP and hTERT, and  
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51 IFN- $\gamma$  production was quantitated by ELISPOT. Negative controls consisted of an HIV  
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53 envelope-derived peptide (HIVenv<sub>584</sub>). Positive controls consisted of 10 ng/ml PMA or a  
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55 CMV pp65-derived peptide (CMVpp65<sub>328</sub>). The number of specific spots was  
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57 determined by subtracting the number of spots in the absence of antigen from the  
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59 number of spots in its presence. T lymphocyte responses to the peptide epitopes were  
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induced following TAE therapy, but no additional responses were observed after DC

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6 transfer. Numbers denote specific spots beyond the upper limit of y-axis. n.d., not  
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12 **Figure 5.** Cytokine and chemokine profiling and arginase activity in sera of patients  
13 treated with OK432-stimulated DCs during TAE therapy (TAE + OK-DC; n = 13) and  
14 the historical controls treated with TAE without DC transfer (TAE; n = 22). (A) Serum  
15 samples were examined for their content of a validated panel of cytokines and  
16 chemokines using the Bioplex assay. Percentage changes in serum levels two weeks  
17 after the treatments were calculated as follows: [(posttreatment level – pretreatment  
18 level) / pretreatment level] X 100. The data are means ± SEM of the groups. \**P* < 0.05  
19 when compared by the Mann-Whitney U test. (B) Serum samples were tested for  
20 arginase activity by conversion of L-arginine to L-ornithine, and for alanine  
21 aminotransferase (ALT) and lactic dehydrogenase (LDH) activities. While there was a  
22 trend for the arginase activity in the TAE + OK-DC group to decrease four weeks after  
23 treatment, the difference did not reach statistical significance (*P* > 0.05). Percentage  
24 changes in serum levels two weeks after the treatments were calculated as follows:  
25 [(posttreatment level – pretreatment level) / pretreatment level] X 100. The data indicate  
26 means ± SEM of the groups.  
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