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Cloning of two members of the calcitonin-family receptors from stingray, *Dasyatis akajei*: Possible physiological roles of the calcitonin family in osmoregulation

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Abbreviations: bp, base pair(s); cDNA, DNA complementary to RNA; CGRP, calcitonin gene-related peptide; calcitonin, CT; calcitonin receptor, CTR; calcitonin-receptor-like receptor, CRLR; mRNA, messenger RNA; PCR, polymerase chain reaction; RACE, rapid amplification of cDNA ends; RT-PCR, reverse transcription-polymerase chain reaction.

Abstract In cartilaginous fish, two cDNAs encoding calcitonin-family receptors were isolated for the first time from the stingray brain. The open reading frame of one receptor cDNA coded a 525-amino acid protein. The amino acid identity of this receptor to human calcitonin-receptor-like receptor (CRLR) is 64.5%, frog CRLR is 64.7%, and flounder CRLR is 61.2% and this was higher than to human calcitonin receptor (CTR) (46.1%), frog CTR (54.7%), and flounder CTR (48.9%). We strongly suggested that this receptor is a ray CRLR based on phylogenetic analysis. In case of the second receptor, amino acid identity among CRLRs (human 50.5%; frog 50.7%; flounder 48.0%) and CTRs (human 43.2%; frog 49.1%; flounder 41.8%) was similar. From phylogenetic analysis of both CRLRs and CTRs, we believe that this receptor is ray CTR. The expression of ray CRLR mRNA was predominantly detected in the nervous system (brain) and vascular system (atrium, ventricle, and gill), which reflects the similar localization of CGRP in the nervous and vascular systems as mammals. It was observed that the second receptor was expressed in several tissues, namely cartilage, brain, pituitary gland, gill, atrium, ventricle, pancreas, spleen, liver, gall bladder, intestine, rectal gland, kidney, testis and ovary. This localization pattern was very similar to flounder CTR. Both receptor mRNAs were strongly expressed in the gill. This suggests that the calcitonin-family members are involved in the osmoregulation of stingray as this fish is known to be euryhaline. When a stingray was transferred to diluted seawater (20% seawater), the expression of both receptors significantly decreased in the gill. Similar results were obtained in the kidney of the stingray. Thus, our cloning and isolation of both receptors in the stingray will be helpful for elucidation of their physiological role (s) such as osmoregulation including calcium metabolism of cartilaginous fish.

1. Introduction

Calcitonin gene-related peptide (CGRP) is a 37 amino acid hormone whose mRNA is co-encoded with calcitonin (CT) mRNA on a single gene (Amara et al., 1982). In mammals, the mRNA synthesis of two hormones is controlled by tissue-specific alternative splicing; CGRP precursor mRNA is synthesized in the neural tissues; CT precursor mRNA is synthesized in the thyroidal C-cells (Rosenfeld et al., 1983). Since CGRP exists widely in the central nervous system (Rosenfeld et al., 1983; Gibson et al., 1984), it is suggested that CGRP plays role as a neuromediator or neuromodulator (Lafont et al., 2007). In addition, CGRP is synthesized in the heart and blood vessels to control blood flow by its potent vasodilatory actions (Mulderry et al., 1985; Wimalawansa, 1997; Brain and Grant, 2004).

In teleosts, the presence of CGRP immunoreactive substances has been demonstrated in trout (Fouchereau-Peron et al., 1990). Jansz and Zandberg (1992) characterized the CGRP gene of salmon, suggesting that salmon CGRP is the product of the alternative splicing of the CT/CGRP gene, common to mammals. We also cloned a part of genomic DNA including the CGRP gene from flounder (*Paralichthys olivaceus*) and detected mRNA expression in the brain and heart (Suzuki et al., 2001), suggesting that CGRP acts as a neuropeptide and a vasodilator in fish as well as mammals. As CGRP immunoreactivity has been detected in the brain of small-spotted dogfish (Molist et al., 1995), and may be quite important in homeostasis in all fish including cartilaginous fish (Lafont et al., 2007). On the other hand, calcitonin receptor-like receptor (CRLR) functions to a receptor for CGRP when it is co-expressed with RAMP1 (McLatchie et al., 1998; Husmann et al., 2000; Sexton et al., 2001). The distribution and expression analysis of CRLR under various physiological and environmental conditions could contribute to the understanding of physiological functions of CGRP in cartilaginous fish.

To study the physiological role of CGRP in cartilaginous fish, the full coding region of CRLR was sequenced from the brain of stingray and the mRNA expression in the various tissues was analyzed. In the process of CRLR cloning, a second member of the calcitonin-family receptors was isolated and cloned. Based on the data of multiple alignment, phylogenetic analysis and tissue distribution pattern, it is apparent that this receptor is equivalent to the stingray calcitonin receptor (CTR). In addition, it was found that the expression of two receptors was decreased in the gill and kidney when the rays were transferred to diluted seawater. Thus, this is the first study that identifies two members of calcitonin-family receptors in cartilaginous fish and indicates the possible physiological roles of CT and CGRP in the osmoregulation of stingray.

2. Materials and methods

2.1. Animals

The brain of male stingray (*Dasyatis akajei*) was used for PCR amplification. Male and female stingrays were used for the analysis of tissue-specific expression and adaptation in seawater. The original concentration of seawater (100%) and diluted seawater (20%) were used for this study. The animals were anesthetized with ethyl 3-amino-benzoate, methanesulfonic acid salt (Sigma-Aldrich Inc., MO, USA) and then dissected for tissue preparation.

All experimental procedures were conducted in accordance with the Guide for the Care and Use of Laboratory Animals prepared by Kanazawa University.

2.2. PCR amplification

Total RNAs were prepared using a total RNA isolation kit (Nippon Gene, Tokyo,

Japan) from the brains of stingray. RT-PCR was performed using Oligotex-dT 30 Super (Takara, Kusatsu, Japan) as an oligo dT primer to prevent genomic DNA contamination (Suzuki et al., 1997). The PCR primers were designed at the well-conserved region of the mammalian CRLR and CTR cDNA sequences (Figure 1). The primer sequences were: 5'A-GYAAAYMGXACXTGGGAYGG; 5'B-GAYTAYTTYCCXGAYTTYGA; 3'A-CCYTCRCAXARCATCCARAA; 3'B:CATCCARAARTARTTRCA (Suzuki et al., 2000). The 1st and the nested-PCR (1st PCR: 5'A/3'A primer set; 2nd PCR: 5'B/3'B primer set) were performed using a *Taq* polymerase and an additional buffer (Takara Bio Inc., Otsu, Japan). The PCR parameters were 35 cycles of 96°C for 30s, 45°C for 1 min, and 72°C for 2 min. The sequencing of the PCR products of the expected length (422 bp) was performed by Dragon Genomics Center, Takara Bio Inc. (Mie, Japan).

2.3. RACE amplification of receptors cDNAs in stingray

Total RNA (1 µg) was obtained from the stingray brain. The complete sequences of the two receptors were obtained by 5'- and 3'-RACE method using a kit 5' RACE System, Version 2.0 and 3' RACE System for Rapid Amplification of cDNA Ends, respectively (Invitrogen, CA, USA). The primer locations for two receptors were described in supplementary figures. RACE was performed using the respective gene-specific primer and the primer of adaptor sequences in the kit. The products were sequenced on an ABI PRISM 3130 sequencer (Applied Biosystems, CA, USA) using BigDye terminators.

2.4. Expression analysis

Total RNAs were prepared from the cartilage, brain, pituitary gland, gill atrium, ventricle, pancreas, spleen, liver, gall bladder, intestine, rectal gland, kidney, and testis of

male stingray, as well as the ovary of female stingray using a total RNA isolation kit (Nippon Gene). RT-PCR was performed using the primer set (CRLR 5':AGACTTCGATCCATCAGA; CRLR 3': AGTCAATGCTGTCTTTA; CTR-like 5': CACGGAAAAGGTCACC; CTR-like 3': AGTGAACAAAGAGTAATAT). The conditions for PCR amplification were 40 cycles of denaturation for 0.5 min at 96°C, annealing for 1 min at 60°C, and extension for 2 min at 72°C, followed by a single cycle at 72°C for 30 min. In case of the β -actin cDNA (AB675482), the amplification using a primer set (5': TTGGCAATGAGCGATTCAGA; 3': CACAGGATTCCATACCCAAGAAA) were consisted of 25 cycles of denaturation for 0.5 min at 96°C, annealing for 1 min at 60°C, and extension for 2 min at 72°C, followed by a single cycle at 72°C for 15 min. The PCR products were analyzed on 2.5% NuSive GTG agarose gel (FMC Bioproducts, ME, USA) and stained with ethidium bromide.

2.5. Phylogenetic analysis

The amino acid sequences were aligned using the CLUSTAL program (Higgins and Sharp 1988). After removing gaps, the verified alignments were used to construct phylogenetic trees. The trees were calculated using MEGA program based on the neighbor-joining method (Saitou and Nei 1987; Kumar et al. 2001). The sequences used as follow: Human-CRLR, NP_005786; Pig-CRLR, Q867C1; Dog-CRLR, XP_545560; Mouse-CRLR, NP_061252; Xenopus-CRLR, NP_001016893; Fugu-CRLR2, BAE45313; Salmon-CRLR, CAD48406; Flounder-CRLR, AB035314; Fugu-CRLR1, BAE45312; Flounder-CTR, AB035315; Chicken-CTR, XP_425985; Bullfrog-CTR, Q28DX2; Fugu-CTR, BAE76018; Medaka-CTR, AAI19273; Pig-CTR, AAA31023; Mouse-CTR, AAI19273; Human-CTR, NP_001733; Dog-CTR, XP_539423; Ascidian-CTR,

BAI63096; Human-CRHR1, NP_004373.

2.6. Analysis of the mRNA expression of both ray CRLR and CTR-like receptors in the gill and kidney after transfer to diluted seawater.

Seawater adapted-stingrays (n = 8, both sexes) were gradually transferred into diluted seawater. In the first day, the stingrays were transferred to 80% seawater and then kept for 6 hours followed by putting them into 60% seawater for remaining of the day. In the next day, the stingray were kept in 40% seawater for 6 hours, and then transferred into 20% seawater. After keeping the stingrays in 20% seawater for 12 hours, the stingrays were anesthetized with ethyl 3-aminobenzoate, methanesulfonic acid salt (Sigma-Aldrich). The gill and kidney were collected from stingray under proper anesthetic condition. Also, the seawater-adapted stingrays (n = 8, both sexes) were anesthetized with ethyl 3-aminobenzoate, methanesulfonic acid salt (Sigma-Aldrich). The gill and kidney were removed from the seawater stingrays.

Total RNA was prepared using a total RNA isolation kit for fibrous tissue and complementary DNA synthesis was performed (RNase Easy Fibrous Mini-Kit, Qiagen GmbH, Hilden, Germany). The real-time PCR amplification was analyzed with a BIO-RAD iCycler (BIO-RAD, Hercules, CA) using the primers for CRLR (5': GGAGAAGCTAAAGACAGCATTGACT; 3': CAGCAGCGAAGCCACTGATA') and for CTR-like (5': GGAAAAGGTCACCAAGATTTGC; 3': AGTCCATGTTTCGGTTGCTCTCT). The annealing temperature in the amplification of both CRLR and CTR-like cDNAs was 63°C. The amplified PCR products have been verified by sequencing to confirm its sequences. The detailed conditions of PCR were described in our previous study (Takahashi et al., 2007). The CRLR and CTR-like mRNA

levels were normalized to the β -actin mRNA level measured by above described primer set (5': TTGGCAATGAGCGATTCAGA; 3': CACAGGATTCCATACCCAAGAAA).

2.7. Statistical analysis

Real-time PCR was performed and the data were analyzed using the Student's *t*-test. The significance level was $P < 0.05$.

3. Results and discussion

3.1. PCR amplification of two receptors in the brain of stingray

Sequence analysis indicated that two types of cDNA fragments are amplified by PCR primer sets at the conserved regions of mammalian CRLR and CTR cDNA sequences. One cDNA fragment (422 bp) had high identity to flounder CRLR (73.4%) and human CRLR (81.3%) and to a lesser extent to flounder CTR (53.9%) and human CTR (56.3%). The second cDNA (422 bp) had high identity to both CRLR (to human 57.0%; to flounder 48.3%) and CTR (to human 59.4%; to flounder 63.3%). Thus, we believe that two receptors belong to the calcitonin-family receptors.

3.2. RACE cloning and sequencing of two calcitonin-family receptors

By RACE cloning, the full sequences of the two calcitonin-family receptors were determined. The open reading frame of one receptor cDNA coded a 525-amino acid protein (see, supplementary data). The amino acid identity of this receptor to CRLR was highest with human CRLR (64.5%), frog (64.7%), and flounder (61.2%) and this was higher than the CTR of human (46.1%), frog (54.7%), or flounder (48.9%). This strongly suggested that the obtained cDNA encodes CRLR in stingray.

The amino acid identity of the second stingray receptor (see, supplementary data) was similar to CRLRs (human 50.5%; frog 50.7%; flounder 48.0%) and CTRs of other species (human 43.2%; frog 49.1%; flounder 41.8%). After alignment of flounder CRLR, flounder CTR, human CRLR, human CTR, and two stingray receptors, three potential sites (Asn-X-Ser/Thr) for N-linked glycosylation, which are conserved in mammalian CRLR and CTR, were identified in stingray receptors before the first putative transmembrane domain. Furthermore, 12 cysteines were conserved among CRLRs, CTRs, and stingray receptors. These glycosylation sites and cysteines are suggested to be important for ligand binding (Qi et al., 1997; Ho et al., 1999). Using BLAST against the DDBJ/NCBI protein database, furthermore, we confirmed that the sequences are most similar to CRLR and CTR. Thus, we concluded that two receptors belong to the calcitonin-family receptors.

3.3. Phylogenetic analysis of calcitonin-family receptors

The phylogenetic tree was determined for the known calcitonin-family receptors together with stingray receptors (see, supplementary data). As a result, the stingray receptors were located with CRLR and CTR in other animals. The first stingray receptor branched from teleost CRLR and composed of a clade of mammalian and xenopus CRLR. This result strongly supports that the amino acid sequence was that of stingray CRLR (ray CRLR). The second stingray receptor, co-located with flounder CTR, and was placed between the CRLRs clade and the non-mammalian CTRs clade (see, supplementary data).

We previously reported amino acid sequences of both CT and CGRP in flounder (Suzuki et al., 2001). The amino acid sequence of flounder CT was more similar to teleosts (78–100%), salamander (81%), reptiles (84%), and chicken (84%) than those of stingray (72%) and frogs (56–78%), and it is largely different from that of mammals

(31–50%). However, the predicted amino acid sequence of flounder CGRP was more conserved than CT among vertebrates and showed 78%, 78%, 78%, 81%, and 73–78% identity to salmon, cod, frog, chicken, and mammalian CGRPs, respectively. Among vertebrates, CGRP is a well-conserved molecule during evolution while CT is quite varied. Corresponding to sequence changes in ligand (CT), the receptor (CTR) seems to be varied largely. Thus, it is highly likely that the second receptor is the stingray equivalent of CTR (ray CTR-like).

3.4. Expression of ray CRLR and ray CTR-like mRNAs in different tissues

Specific PCR products of ray CRLR mRNA were amplified from brain, gill, atrium and ventricle and detected in cartilage, pituitary gland, spleen, liver, gall-bladder, intestine, rectal gland, kidney, testis and ovary (Fig. 2). Pancreas did not show any expression with the present conditions (Fig. 2). The ray CRLR mRNA expression was found in brain, heart and intestine and this same as seen in mammals (Eysselein et al., 1991; Poyner, 1992), suggesting similar functions in those tissues. In spleen, relatively high expression of ray CRLR was detected. Arlot-Bonnemains et al. (1991) reported that CGRP-specific binding in trout tissues is high in the spleen although the function has not yet been elucidated. In the early development of gut in zebrafish, the number of nerve cell bodies and fibers gradually increased and CGRP was detected in these nerve cells (Olsson et al., 2008). In another study, cod CGRP (10^{-9} – 10^{-7} M) inhibited the motility of spontaneously active ring preparations from the cod intestine (Shahbazi et al., 1998) suggesting that CGRP plays a role in fish gut development and physiology. The localization of CGRP in the stingray gut suggests that CGRP may have significant roles in its gut.

Recently, involvement of CGRP in the outgrowth of the gubernaculum has been noted,

and it has been suggested to play a role in testicular descent in mammals (Chan et al., 2009). CGRP released from the genitofemoral nerve causes maximal mitosis in the gubernacular bulb of mammals (Chan et al., 2009). In stingray, this hormone may involve in testicular development that regulating the balance between cell proliferation and apoptosis.

Specific PCR products of ray CTR-like mRNA were amplified from cartilage, brain, pituitary gland, gill, atrium, ventricle, pancreas, spleen, liver, gall bladder, intestine, rectal gland, kidney, testis, and ovary (Fig. 2). In the cartilage of stingray, we detected ray CTR-like mRNA expression. It has been reported that CT acts on mammalian cartilage and promotes synthesis (Karsdal et al., 2006; Sondergaard et al., 2006). In the stingray, thus, CT may function to the growth of cartilage. CT-immunoreactive cells were found in the intestine of goldfish and appeared to control absorption of nutrients (Okuda et al., 1999). Therefore, the presence of ray CTR-like mRNA in intestine corresponds well with this physiological role of CT in fish. Previously mRNA expression of salmon CTR was detected in brain and pituitary gland and this was consistent with another study in mammalian tissues (Azria, 1989) where it was suggested that CT might act as a neurotransmitter.

In female sharks, it had been shown that plasma CT levels were elevated during reproductive periods (Nichols et al., 2003). We previously reported that estrogen-specific-binding and estrogen receptor mRNA were present in the stingray ultimobranchial gland (CT secreting organ in sub-mammalian vertebrates) (Yamamoto et al., 1996). The possible involvement of CT in stingray reproductive physiology is implied because of the presence of this receptor in the ovary and testis of stingray in the present study.

Ray CRLR and ray CTR-like mRNAs were found to be expressed in the excretory

organs such as gill, kidney and rectal gland. As this fish is euryhaline (de Vlaming and Sage, 1973; Janech et al., 2003; Evans et al., 2010) and CGRP functions to osmoregulation in teleosts (Lamharzi et al., 1996; Suzuki et al., 2002; Lafont et al., 2007), we examined ray CRLR and ray CTR-like mRNA expression in the gill and kidney when stingrays were transferred to the diluted seawater.

3.5. Changes in mRNA expression of both ray CRLR and CTR-like receptors in the gill and kidney when transferred to diluted seawater

The results are shown in Fig. 3. The ray CTR-like mRNA expression in the gill and kidney was higher than the ray CRLR mRNA. These results coincided with the tissue specific expressions (Fig. 2). When stingrays were transferred to the diluted seawater (20% seawater), both receptors mRNA expression significantly decreased in the gill. Similar results were obtained in the stingray kidney. When rainbow trout were adapted to seawater, their plasma CGRP levels and CGRP binding sites in the gill increased (Lamharzi et al., 1996). Similarly when flounder were moved from seawater to freshwater, CRLR mRNA expression decreased (Suzuki et al., 2002). Thus, CGRP may play a role in osmoregulation in teleosts (Lafont et al., 2007).

Interestingly, the plasma level of CT did not change when freshwater adapted eels were moved to 100% seawater (Suzuki et al., 1999). This was reflected in the study where the flounder were transferred from seawater to freshwater and CTR mRNA expression in the gill did not alter (Suzuki et al., 2002). Therefore, CT may play a role in osmoregulation in cartilaginous fish and not in teleosts, as CT has some functions in osmoregulation as the CTR-like mRNA expression in both stingray gill and kidney was decreased when the fish were transferred from seawater to freshwater. Further study is required to elucidate the

possible roles of CT and CGPR in cartilaginous fish.

3.6. Conclusions

(1) cDNA encoding CRLR was cloned from the stingray brain. Tissue-expression analysis, using RT-PCR, indicated that ray CRLR mRNA was detected mainly in the nervous system (brain) and vascular system (atrium, ventricle, and gill), indicating that CGRP may play roles in the nervous and vascular systems as it does in mammals.

(2) Another receptor cDNA was cloned from the brain of stingray. Based on the data of multiple alignments, phylogenetic analysis and tissue distribution pattern, this receptor is CTR equivalent in stingray.

(3) When stingray transferred to the diluted seawater (20% seawater), mRNA expression of both receptors were significantly decreased in the stingray gill. Similar results were obtained in the kidney. Thus, identification of two members of the calcitonin-family receptors in the stingray will assist in the elucidation of the physiological roles of these receptors and their ligands in cartilaginous fish.

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Figure Legends

Fig. 1. Location and sequence of primers. The CRLR and CTR molecules have four extracellular domains (Ecd) 1-4, seven transmembrane domains (Tmd 1-7), and for intracellular domains (Icd 1-4). The PCR primers were designed to the highly conserved region of the mammalian CRLR and CTR cDNA sequences.

Fig. 2. Tissue specific expression of ray CRLR and ray CTR-like mRNAs by RT-PCR. After PCR amplification, the PCR products were analyzed on 2.5% NuSive GTG agarose gel (FMC Bioproducts, ME, USA) and stained with ethidium bromide.

Fig. 3. Changes in mRNA expression of both ray CRLR and CTR-like receptors in the gill (A) and kidney (B) when transferred to diluted (20%) seawater. * indicates statistically significant difference at $P < 0.05$ from the values in the control. The data expressed as mean \pm SEM (n = 8).

Supplementary data of Legends

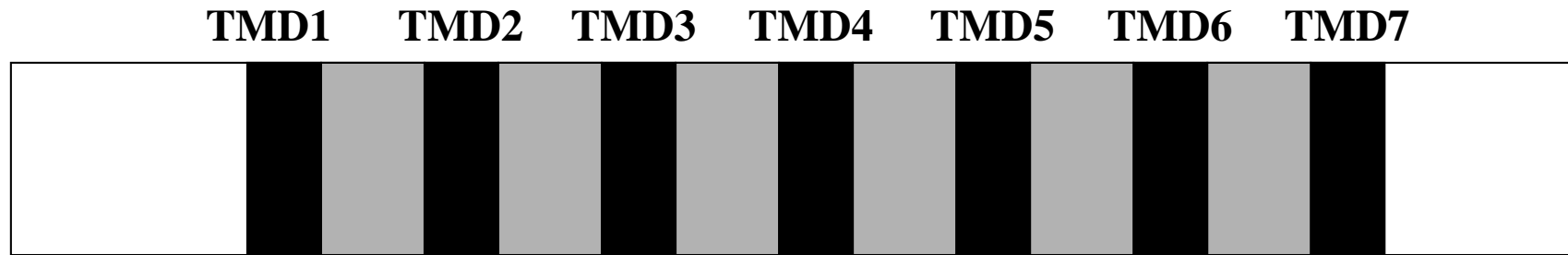
Fig. 1. Nucleotide and deduced amino-acid sequences of ray CRLR cDNA. The nucleotide sequence obtained by PCR using a 5'B/3'B primer set is shown in bold. The boxes and underlining indicate the location of 5' RACE primers and 3' RACE primers, respectively. The stop codon TGA is indicated by the asterisk. This data has been available under Genbank accession no. AB665754.

Fig. 2. Nucleotide and deduced amino-acid sequences of ray CTR-like cDNA. The nucleotide sequence obtained by PCR using a 5'B/3'B primer set is shown in bold. The boxes and underlining indicate the location of 5' RACE primers and 3' RACE primers, respectively. The stop codon TGA is indicated by the asterisk. This data has been available under Genbank accession no. AB665755.

Fig. 3. Multiple alignment of the amino-acid sequences of flounder CRLR (fCRLR), flounder CTR (fCTR), human CRLR (hCRLR), human CTR (hCTR), stingray CRLR (ray CRLR), and stingray CTR-like (ray CTR-like). Deletion of amino-acid residues is indicated by dashes. Conserved amino-acid residues at least three sequences are indicated by shaded areas. Underlining indicate putative transmembrane domains I to VII. # and * indicate potential N-linked glycosylation sites and cysteine residues commonly to these receptors, respectively. DDBJ/EMBL/GenBank accession numbers are as follow: fCRLR (AB035314), fCTR (AB035315), hCRLR (NP_005786), hCTR (NP_001733), ray CRLR (AB665754), and ray CTR-like (AB665755).

Fig. 4. Phylogenetic tree of CRLRs and CTRs constructed by the neighbor-joining method. The number beside each branch indicates the percentage of times that a node was supported in 1000 bootstrap pseudoreplications. The scale bar indicates an evolutionary distance of 0.1 amino acid substitutions per protein.

Location of primer sets



ECD1

ICD1

ECD2

ICD2

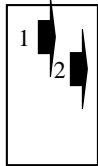
ECD3

ICD3

ECD4

ICD4

Forward primer



Reverse primer



TMD: transmembrane domain

ECD: extracellular domain

ICD: intracellular domain

Forward primer

1:GYAAYMGXACXTGGGAYGG

2:GAYTAYTTYCCXGAYTTYGA

Reverse primer

1:CCYTCRCAXARCATCCARAA

2:CATCCARAARTARTTRCA

Figure 1 Suzuki et al.

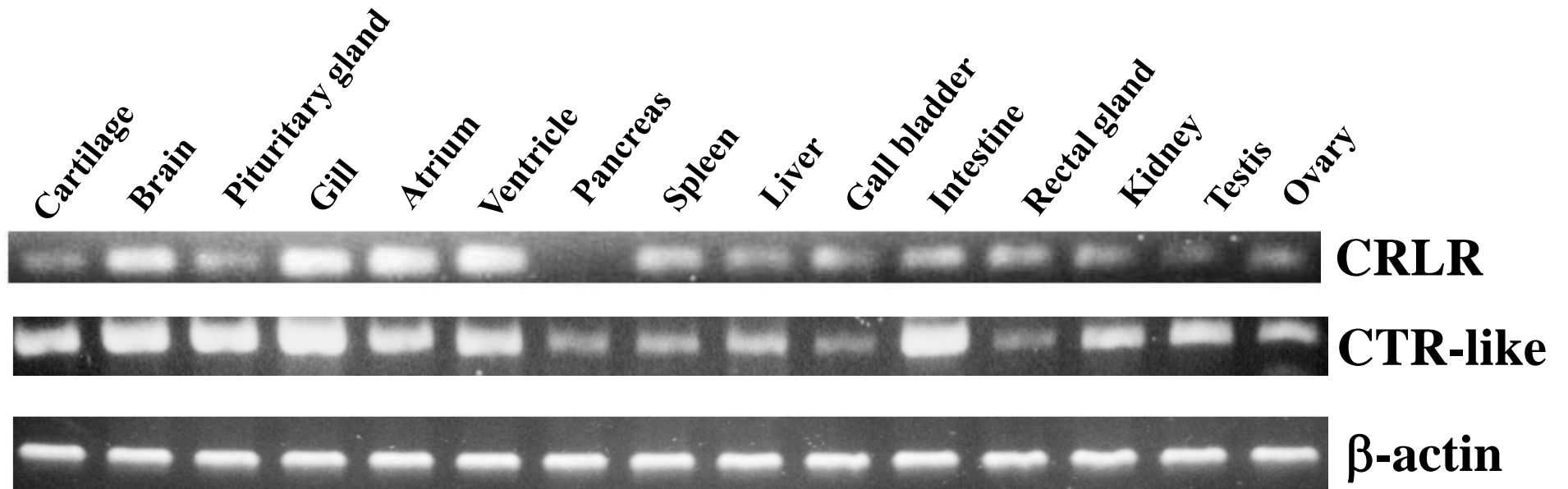
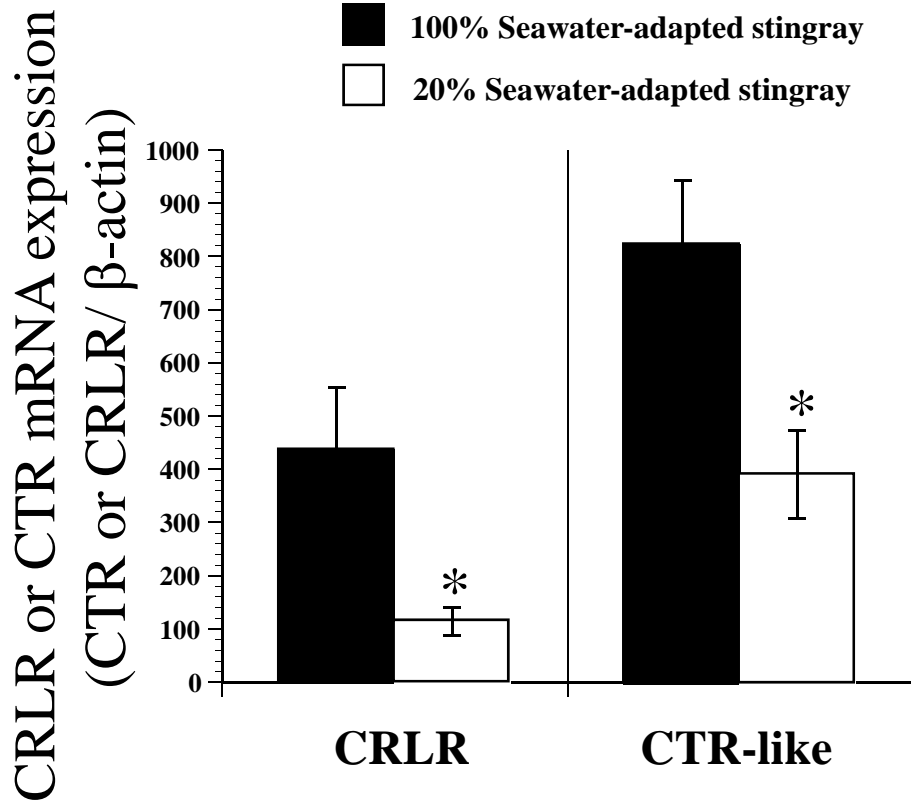


Figure 2 Suzuki et al.

A) Gill



B) Kidney

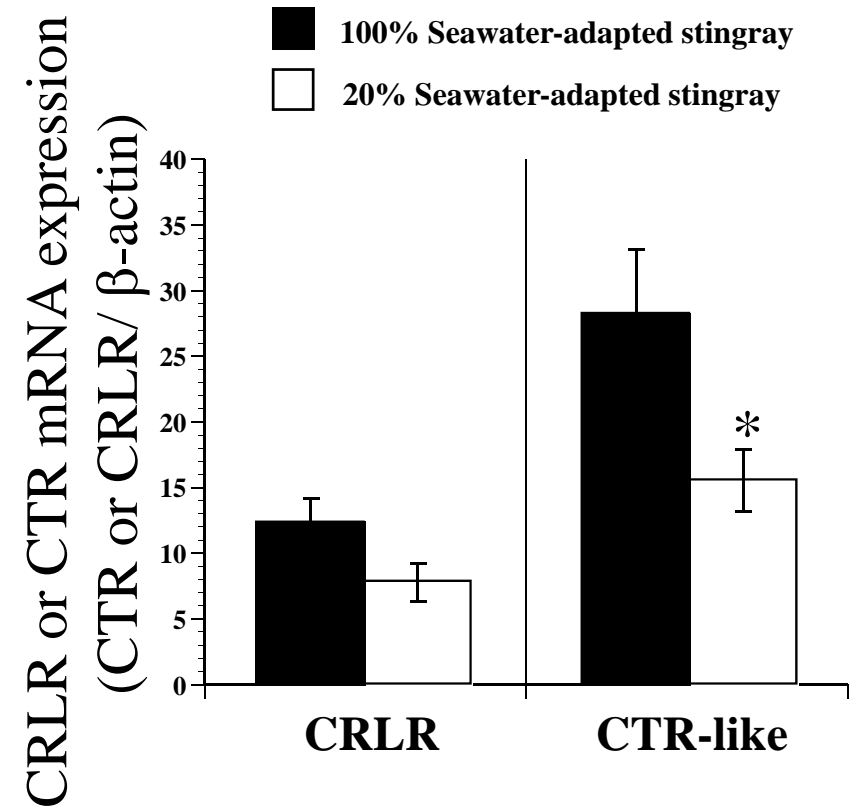


Figure 3 Suzuki et al.