

Anti-tumor effects of a nonsteroidal anti-inflammatory drug zaltoprofen on chondrosarcoma via activating peroxisome proliferator-activated receptor gamma and suppressing matrix metalloproteinase-2 expression.

| | |
|------------------------------|--|
| 著者 | Higuchi Takashi, Takeuchi Akihiko, Munesue Seiichi, Yamamoto Norio, Hayashi Katsuhiko, Kimura Hiroaki, Miwa Shinji, Inaki Hiroyuki, Shimozaki Shingo, Kato Takashi, Aoki Yu, Abe Kensaku, Taniguchi Yuta, Aiba Hisaki, Murakami Hideki, Harashima Ai, Yamamoto Yasuhiko, Tsuchiya Hiroyuki |
| 著者別表示 | 武内 章彦, 棟居 聖一, 山本 憲男, 林 克洋, 三輪 真嗣, 谷口 裕太, 原島 愛, 山本 靖彦, 土屋 弘行 |
| journal or publication title | Cancer Medicine |
| volume | 7 |
| number | 5 |
| page range | 1944-1954 |
| year | 2018-05 |
| URL | http://doi.org/10.24517/00053846 |

doi: 10.1002/cam4.1438



ORIGINAL RESEARCH

Anti-tumor effects of a nonsteroidal anti-inflammatory drug zaltoprofen on chondrosarcoma via activating peroxisome proliferator-activated receptor gamma and suppressing matrix metalloproteinase-2 expression

Takashi Higuchi¹ , Akihiko Takeuchi¹ , Seiichi Munesue², Norio Yamamoto¹, Katsuhiro Hayashi¹, Hiroaki Kimura¹, Shinji Miwa¹ , Hiroyuki Inatani¹, Shingo Shimosaki¹, Takashi Kato¹, Yu Aoki¹, Kensaku Abe¹ , Yuta Taniguchi¹, Hisaki Aiba¹, Hideki Murakami¹, Ai Harashima², Yasuhiko Yamamoto² & Hiroyuki Tsuchiya¹

¹Department of Orthopaedic Surgery, Kanazawa University Graduate School of Medical Sciences, 13-1 Takara-machi, Kanazawa 920-8641, Japan

²Department of Biochemistry and Molecular Vascular Biology, Kanazawa University Graduate School of Medical Sciences, 13-1 Takara-machi, Kanazawa 920-8640, Japan

Keywords

Chondrosarcoma, MMP2, nonsteroidal anti-inflammatory drugs, PPAR γ , zaltoprofen

Correspondence

Akihiko Takeuchi, Department of Orthopaedic Surgery, Kanazawa University Graduate School of Medical Sciences, 13-1 Takara-machi, Kanazawa 920-8641, Japan.
Tel: +81-76-265-2374;
Fax: +81-76-234-4261;
E-mail: a_take@med.kanazawa-u.ac.jp

Funding Information

Japan Society for the Promotion of Science (JP15K10435, JP17K16682).

Received: 1 December 2017; Revised: 15 February 2018; Accepted: 17 February 2018

Cancer Medicine 2018; **7**(5):1944–1954

doi: 10.1002/cam4.1438

Abstract

Surgical resection is the only treatment for chondrosarcomas, because of their resistance to chemotherapy and radiotherapy; therefore, additional strategies are crucial to treat chondrosarcomas. Peroxisome proliferator-activated receptor gamma (PPAR γ) is a ligand-activated transcription factor, which has been reported as a possible therapeutic target in certain malignancies including chondrosarcomas. In this study, we demonstrated that a nonsteroidal anti-inflammatory drug, zaltoprofen, could induce PPAR γ activation and elicit anti-tumor effects in chondrosarcoma cells. Zaltoprofen was found to induce expressions of PPAR γ mRNA and protein in human chondrosarcoma SW1353 and OUMS27 cells, and induce PPAR γ -responsible promoter reporter activities. Inhibitory effects of zaltoprofen were observed on cell viability, proliferation, migration, and invasion, and the activity of matrix metalloproteinase-2 (MMP2); these effects were dependent on PPAR γ activation and evidenced by silencing PPAR γ . Moreover, we showed a case of a patient with cervical chondrosarcoma (grade 2), who was treated with zaltoprofen and has been free from disease progression for more than 2 years. Histopathological findings revealed enhanced expression of PPAR γ and reduced expression of MMP2 after administration of zaltoprofen. These findings demonstrate that zaltoprofen could be a promising drug against the malignant phenotypes in chondrosarcomas via activation of PPAR γ and inhibition of MMP2 activity.

Introduction

Chondrosarcomas are the second largest group of malignant bone tumors following osteosarcoma, and they arise from various sites including extremity, trunk, spine, and head [1]. Because of their resistance to chemotherapy and radiotherapy, surgical resection is the only treatment for chondrosarcomas; however, they often recur after incomplete resection or metastasis [2]. Therefore, novel strategies are crucial to overcome the malignancy of chondrosarcomas.

Peroxisome proliferator-activated receptor gamma (PPAR γ) is a ligand-activated transcription factor that belongs to the nuclear hormone receptor superfamily [3]. PPAR γ plays a central role in the differentiation of adipocytes and is reported to be expressed in various malignant tumors [4]. Anti-tumorigenic effects through ligand activation of PPAR γ have been demonstrated in certain malignancies [5, 6]. Furthermore, in chondrosarcoma, PPAR γ has been considered as a possible therapeutic target [7]. We have recently reported a case of giant cell tumor

of bone, in which complete necrosis and high expression of PPAR γ appeared in the resected specimen after zaltoprofen administration [8]. Zaltoprofen is a propionic-acid derivative of nonsteroidal anti-inflammatory drugs (NSAIDs) [8]. Previous studies have shown that several NSAIDs could serve as exogenous ligands for PPAR γ [9–11]. We consider the possibility of anti-tumorigenic effects by zaltoprofen via the activation of PPAR γ in musculoskeletal neoplasms.

Matrix metalloproteinase-2 (MMP2) is a member of the MMP family and can degrade matrix collagen and basement membrane, which plays a critical role in tumor invasion and metastasis [12, 13]. Overexpression of MMP2 has been detected in various types of cancer, including chondrosarcomas [14]. Therefore, the inhibition of MMP2 is a potent strategy to suppress tumor progression in chondrosarcoma [14, 15]. Recent studies report that the activation of PPAR γ inhibited MMP2 production and tumor cell invasion in several types of cancer, indicating that downregulation of MMP2 by PPAR γ could represent an anti-tumorigenic procedure [16, 17].

The purpose of this study was to assess the anti-tumor effects of zaltoprofen via regulation of MMP2 and PPAR γ activity, and examine whether zaltoprofen could be a novel therapeutic agent in human chondrosarcoma.

Material and Methods

Cells and reagents

Human chondrosarcoma cell lines, OUMS27, and SW1353 were purchased from Japanese Collection of Research Bioresources Cell Bank (JCRB Cell Bank, Tokyo, Japan) and American Type Culture Collection (ATCC, Manassas, VA), respectively. They were cultured in Dulbecco's modified Eagle's medium (DMEM) (Wako Pure Chemical Industries, Ltd., Osaka, Japan) supplemented with 10% fetal bovine serum (FBS), penicillin (100 U/mL) and streptomycin (100 μ g/mL) at 37°C. Zaltoprofen and ARP100, a selective MMP2 inhibitor, were purchased from Santa Cruz (Santa Cruz Biotechnology, Inc., Santa Cruz, CA) and dissolved in dimethyl sulfoxide (DMSO).

Western blot analysis

OUMS27 and SW1353 cells treated with various concentrations of zaltoprofen for 24 h were lysed by RIPA buffer (150 mmol/L NaCl, 1% sodium deoxycholate, 1% Triton X-100, 0.1% SDS, 10 mmol/L Tris, pH 7.2, 100 μ mol/L sodium orthovanadate, and 50 mmol/L sodium fluoride) containing protease cocktail inhibitors (Sigma-Aldrich Co., Munich, Germany). The cell lysates

(100 μ g) were boiled in SDS sample buffer, resolved by SDS-PAGE (12.5%), run on an e-PAGE (ATTO Co., Tokyo, Japan), and then transferred onto PVDF membrane (Merck Millipore, Darmstadt, Germany). The membranes were blocked with Odyssey Blocking Buffer PBS (LI-COR Inc., Lincoln, NE) for 1 h and then reacted with a mouse monoclonal anti-PPAR γ antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, CA) at 4°C for 24 h. After washing with PBS with Tween-20, the membranes were reacted with IRDye 680 anti-mouse IgG (LI-COR Inc., Lincoln, NE) for 30 min, and immunoreacted bands were visualized using the Odyssey Infrared Imaging system (LI-COR Inc., Lincoln, NE). The glyceraldehyde-3-phosphate dehydrogenase (GAPDH) protein was used as a house-keeping protein and detected with a monoclonal anti-GAPDH antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, CA).

Quantitative real-time reverse transcription-polymerase chain reaction (qRT-PCR)

After treatment with zaltoprofen for 24 h, total RNA extracted from OUMS27 and SW1353 cells using Ambion TRIzol reagent (Invitrogen, Carlsbad, CA) was quantified using a NanoDrop Lite spectrophotometer (Thermo Fisher Scientific Inc., Waltham, MA). The RNA sample (100 ng) was reverse-transcribed into cDNA on a T100 Thermal Cycler (Bio-Rad Laboratories, Inc., Hercules, CA) using AffinityScript cDNA Synthesis Kit (Agilent Technologies Inc., Santa Clara, CA) in accordance with the manufacturer's instructions. The qRT-PCR was performed with 10 ng cDNA per well on a StepOne™ Real-time PCR System (Applied Biosystems, Waltham, MA) using the QuantiTect SYBR Green PCR Kits (Qiagen, Hilden, Germany). *GAPDH* was used as an internal control. Primers used for qRT-PCR were as follows: primer for PPAR γ was purchased by Qiagen (QuantiTect Primer Assay, PPAR γ). *GAPDH* forward, 5'-TGCACCA-CCAACTGC TTAGC-3'; reverse, 5'-GGCATGGACTGTGGTCATGA G-3'.

PPAR γ reporter assay

PPAR γ activity was assessed using a luciferase reporter gene assay kit (INDIGO Biosciences, Inc., State College, PA), according to the manufacturer's instructions [18]. Briefly, reporter cells were dispensed into wells and then immediately treated with zaltoprofen. Following 24 h incubation, treatment media were discarded and luciferase detection reagent was added. The intensity of light emission from each sample well was quantified using a plate-reading luminometer (TriStar LB 941 Multimode

Microplate Reader, Berthold Technologies, Bad Wildbad, Germany).

PPAR γ knockdown

SureSilencing short hairpin RNA (shRNA) plasmid for human PPAR γ (Qiagen, Hilden, Germany) was used to knockdown PPAR γ gene. PPAR γ -specific shRNA (shPPAR) or nontarget control shRNA (shNTC) plasmid were transfected in OUMS27 and SW1353 cells using Attractene transfection reagent (Qiagen, Hilden, Germany) according to the manufacturer's protocol [19]. Transfected cells were selected by resistance to 600 μ g/mL neomycin (G418 solution, Roche Diagnostics, Indianapolis, IN) and the knockdown efficiency was confirmed by qRT-PCR. Insert sequence of PPAR γ -specific shRNA is CCACGAGATCATTACACA AT, and nontarget control shRNA is GGAATCTCATTTCGAT GCATAC. Finally, we established PPAR γ knockdown cell lines (SW-1353^{shPPAR} and OUMS-27^{shPPAR}) and nontarget control cell lines (SW-1353^{shNTC} and OUMS-27^{shNTC}).

Cell proliferation assay

The cell proliferation assay was performed using a Cell Counting Kit-8 (Dojindo Laboratory, Mashiki-machi, Japan). Briefly, 5×10^3 cells/well were seeded in 96-well plates. After 24 h incubation at 37°C, indicated concentrations of zaltoprofen were added to the medium, and after further 72 h of incubation, the medium was removed and a Cell Counting Kit-8 reagent was added to the plates. After a final incubation at 37°C for 2 h, the absorbance was measured at 450 nm by a microplate reader (iMark Microplate Absorbance Reader, Bio-Rad Laboratories, Inc., Hercules, CA). In addition, 5-ethynyl-2'-deoxyuridine (EdU) proliferation assay was carried out using Click-iT[®] EdU Alexa Fluor[®] 555 Imaging Kit (Invitrogen, Carlsbad, CA) according to the manufacturer's protocol [20]. Briefly, OUMS27^{shPPAR}, OUMS27^{shNTC}, SW1353^{shPPAR}, and SW1353^{shNTC} were seeded on slide chambers and incubated overnight. After treatment with zaltoprofen (400 μ mol/L) or DMSO for 24 h, cells were treated with EdU (10 mmol/L) for 1 h. Then, they were fixed with 4% paraformaldehyde (Wako Pure Chemical Industries, Ltd., Osaka, Japan), washed with 3% bovine serum albumin in PBS, and permeabilized with 0.5% Triton X-100. Cells were then incubated with the Click-iT reaction cocktail, followed by Hoechst 33342 (NucBlue Live ReadyProbes Reagent, Invitrogen, Carlsbad, CA), and were observed using fluorescence microscope (BZ-9000, Keyence Co., Osaka, Japan). The number of EdU-positive cells and Hoechst 33342-positive cells was counted using Image J software according to the manufacturer's instructions [21].

Cell migration assay

Cell migration was evaluated by a monolayer denudation assay as described previously [22]. Briefly, OUMS27^{shPPAR}, OUMS27^{shNTC}, SW1353^{shPPAR}, and SW1353^{shNTC} were seeded at a density of 1×10^6 cells/well and grown to confluence in a 6-well plate. Cells were then wounded by denuding a strip of the monolayer approximately 1 mm in width with a 200 μ L pipette tip. After washing cells with serum-free DMEM, DMSO (0.1%), zaltoprofen (200 or 400 μ mol/L), a selective MMP2 inhibitor, and ARP100 (25 μ mol/L) were added to each well, and then cells were incubated for 30 h. The rate of wound closure was assessed in six separate fields using Image J software [23].

Cell invasion assay

The invasive potential of OUMS27^{shPPAR}, OUMS27^{shNTC}, SW1353^{shPPAR}, and SW1353^{shNTC} was assessed using Matrigel invasion chambers (24-well, 8 μ m pore size) (BD Biosciences, San Diego, CA) [24]. Cells were plated into the upper chamber at an initial density of 4×10^4 cell/ml in 0.5 mL serum-free DMEM, and 0.5 mL of DMEM supplemented with 10% FBS and fibronectin 500 μ g/mL was added to the bottom chambers as chemoattractant. After 24 h incubation in the presence of DMSO (0.1%), zaltoprofen (200 or 400 μ mol/L) and ARP100 (25 μ mol/L) into both the upper and bottom chambers, the noninvasive cells that remained at the top chambers were removed by scraping using a cotton swab, and the invasive cells at the bottom of the membranes were fixed with methanol. Cells were then washed twice with PBS and stained with 1% crystal violet for 30 min at RT. The invasive cells were counted in six separate fields using an optical microscope at 20 \times magnification [24].

Gelatin zymography analysis

The expression of activated MMPs in conditioned medium was assayed by gelatin zymography [25]. The cells (OUMS27^{shPPAR}, OUMS27^{shNTC}, SW1353^{shPPAR}, and SW1353^{shNTC}) were seeded at a density of 1×10^6 cells/well and grown to 70–80% confluency in a 6-well plate. Then the cells were cultured in serum-free DMEM containing DMSO (0.1%) or zaltoprofen (200 or 400 μ mol/L) for 24 h. Afterward, the supernatants were collected and mixed with 5 \times SDS sample buffer without reducing agent or heating. The samples were loaded onto a Gelatin-zymography Precast GEL (Cosmo Bio Co., Ltd., Tokyo, Japan) and subjected to electrophoresis. Following electrophoresis, the gel was washed twice with 2.5% Triton X-100, 10 mmol/L Tris-HCl (pH 8.0) for 30 min to remove SDS, and then incubated in 1 μ mol/L ZnCl₂,

10 mmol/L CaCl₂, 0.2 mol/L NaCl, and 50 mmol/L Tris-HCl (pH 8.0) for 28 h at 37°C. Enzyme activity was visualized as negative staining with Coomassie Brilliant Blue R250 and gel was washed with destaining solution until clear bands were visible.

A clinical case of chondrosarcoma with zaltoprofen administration

A 34-year-old man presented with grade 1 chondrosarcoma of cervical spine underwent tumor excision by his previous doctor. After 1 year from initial surgery, recurrent tumor was detected and excised by the same doctor, and the pathological diagnosis of the resected section was grade 2 chondrosarcoma. Although, adjuvant radiotherapy was conducted, the tumor had gradually enlarged. After 3 years from second surgery, he was referred to our hospital for severe pain and numbness of the left upper arm. There was bone destruction of the preserved vertebral arch and the tumor invaded the spinal canal, excluding the dura of cervical spine from anterolateral side of C5-6. We performed radiotherapy and tumor excision of the posterior side of C2-4. However, the tumor of C5-6 was unresectable; hence, 240 mg/day of zaltoprofen was administered to the patient against his persistent pain and numbness on the neck and the left upper arm. For approximately 2 years since zaltoprofen administration, the tumor remained within mild growth for grade 2 chondrosarcoma; however, instability and anteflexion of the cervical spine proceeded. Therefore, the patient underwent tumor excision and posterior fixation of the cervical spine. At the final follow-up, the patient was alive with disease without distant metastasis for more than 7 years after the initial diagnosis and 4 years after the administration of zaltoprofen (Fig. 5). The resection paraffin sections of two different treatment periods, the second surgery (preadministration specimen, Pre) and final surgery (postadministration specimen, Post), were used for hematoxylin-Eosin (HE) staining and PPAR γ and MMP2 immunofluorescent staining with anti-PPAR γ antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, CA) and anti-MMP2 antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, CA), respectively, in combination with IRDye 680 anti-mouse IgG (LI-COR Inc., Lincoln, NE) as a secondary antibody. Nuclear staining was performed with 4', 6-diamidino-2-phenylindole (DAPI).

Consent for use of human specimens

Declaration of Helsinki protocols were followed and the patient gave their written, informed consent, which was approved by the Ethics Committee of Kanazawa University.

Statistical analysis

Each experiment was repeated independently at least three times. All data are presented as mean \pm SEM. Student's *t*-test and ANOVA using the Tukey-Kramer post hoc test were used when mean differences were identified between the groups. All *P*-values were two-sided and *P*-values of 0.05 or less were considered statistically significant. All statistical analyses were performed with EZR (Saitama Medical Center, Jichi Medical University, Saitama, Japan) [26], which is a graphical user interface for R (The R Foundation for Statistical Computing, Vienna, Austria).

Results

Effects of zaltoprofen on the induction and activation of PPAR γ in chondrosarcoma cells

qRT-PCR demonstrated a significant upregulation of PPAR γ mRNA in OUMS27 and SW1353 cells after 24 h-treatment with 400 μ mol/L zaltoprofen (Fig. 1A). Moreover, zaltoprofen treatment dose-dependently increased expression levels of PPAR γ protein in both cell lines as observed by western blotting (Fig. 1B), indicating that zaltoprofen induces PPAR γ expression in chondrosarcoma cells. We next investigated the activity of PPAR γ using a luciferase reporter assay system. The zaltoprofen treatment could significantly and dose-dependently upregulate PPAR γ -responsible promoter activities, and the EC₅₀ (effective concentration) was found to be 47.3 μ mol/L (Fig. 1C); this suggests a strong induction of PPAR γ activity, with a relatively mild upregulation of PPAR γ expression by zaltoprofen.

Inhibitory effects of zaltoprofen on the viability and proliferation of chondrosarcoma cells

We next investigated whether zaltoprofen could reduce the viability of OUMS27 and SW1353 chondrosarcoma cell lines in vitro. When cells were treated with various concentrations (0–400 μ mol/L) of zaltoprofen for 72 h, cell viability of OUMS27 and SW1353 cells was significantly and dose-dependently lowered by zaltoprofen treatment (Fig. 2A). In addition, cell proliferation assay was performed to evaluate EdU incorporation into the nucleus. The number of EdU-incorporated cells per Hoechst 33342-positive nuclei ratio was calculated and the involvement of PPAR γ in the cell proliferation was analyzed with or without downregulation of PPAR γ by a specific shRNA knockdown system (Fig. 2B). As a result, the EdU-positive cell ratio was significantly and dose-dependently decreased by the treatment of zaltoprofen in both OUMS27 and SW1353 cells, when compared to controls (Fig. 2C and D). Moreover, the downregulation

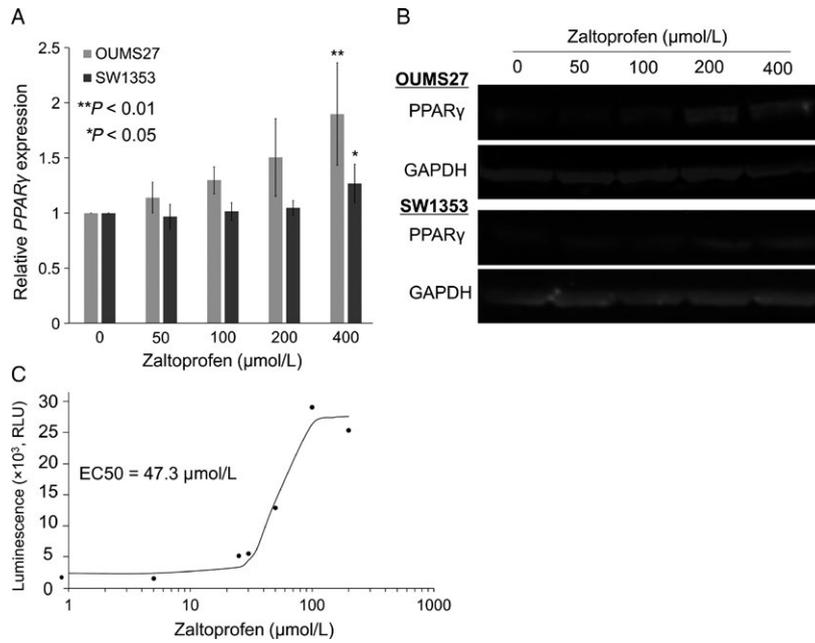


Figure 1. (A) Effects of zaltoprofen on PPARγ mRNA expression in OUMS27 and SW1353 cells by qRT-PCR. Values represented as the mean ± SEM. Asterisks denote a statistically significant difference (**P* < 0.05 and ***P* < 0.01) between cells with and without treatment of zaltoprofen (400 μmol/L). (B) Effects of zaltoprofen on PPARγ protein expression in OUMS27 and SW1353 cells by western blotting. GAPDH was used as an internal control. (C) PPARγ activity was determined by luciferase reporter gene assay. Reporter cells were treated with zaltoprofen (0–200 μmol/L) for 24 h, followed by measurement of luciferase activities.

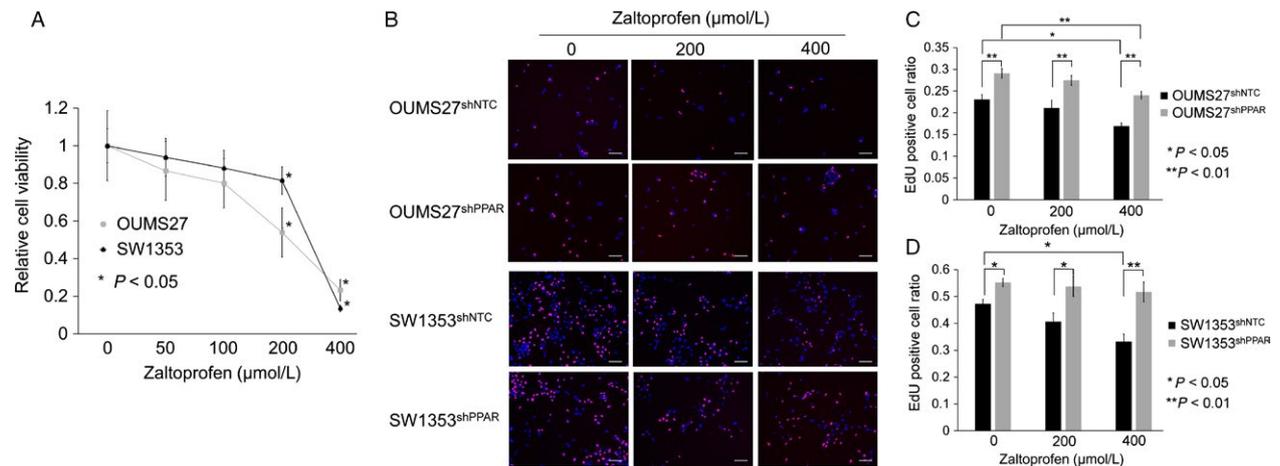


Figure 2. (A) Effects of zaltoprofen on the viability of OUMS27 and SW1353 cells. The cells were treated with or without zaltoprofen for 72 h, followed by WST assay. Asterisks denote a statistically significant difference (**P* < 0.05) between cells with and without treatment of zaltoprofen (200 or 400 μmol/L). (B) Representative photos of 5-ethynyl-2'-deoxyuridine (EdU) proliferation assay in PPARγ knockdown cell lines, OUMS27^{shPPAR} and SW1353^{shPPAR}, and the control cell lines, OUMS27^{shNTC} and SW1353^{shNTC} with or without the treatment of zaltoprofen for 24 h. Scale bar = 100 μm. (C, D) The EdU-positive cell ratio [EdU-positive nuclei (red)/Hoechst 33342-positive nuclei (blue)] in OUMS27 (C) and SW1353 cells (D). Values represented as the mean ± SEM. Asterisks denote a statistically significant difference (**P* < 0.05 and ***P* < 0.01).

of the EdU-positive cell ratio by zaltoprofen was significantly inhibited by PPARγ silencing in chondrosarcoma cells, OUMS27^{shPPAR} and SW1353^{shPPAR} cells (Fig. 2C and D). These data suggest that inhibitory effects on cell viability and proliferation by zaltoprofen were via PPARγ.

Inhibitory effects of zaltoprofen on cell migration and invasion in chondrosarcoma cells

To examine whether zaltoprofen contributes to tumor malignancy in terms of metastatic potentials, we performed in

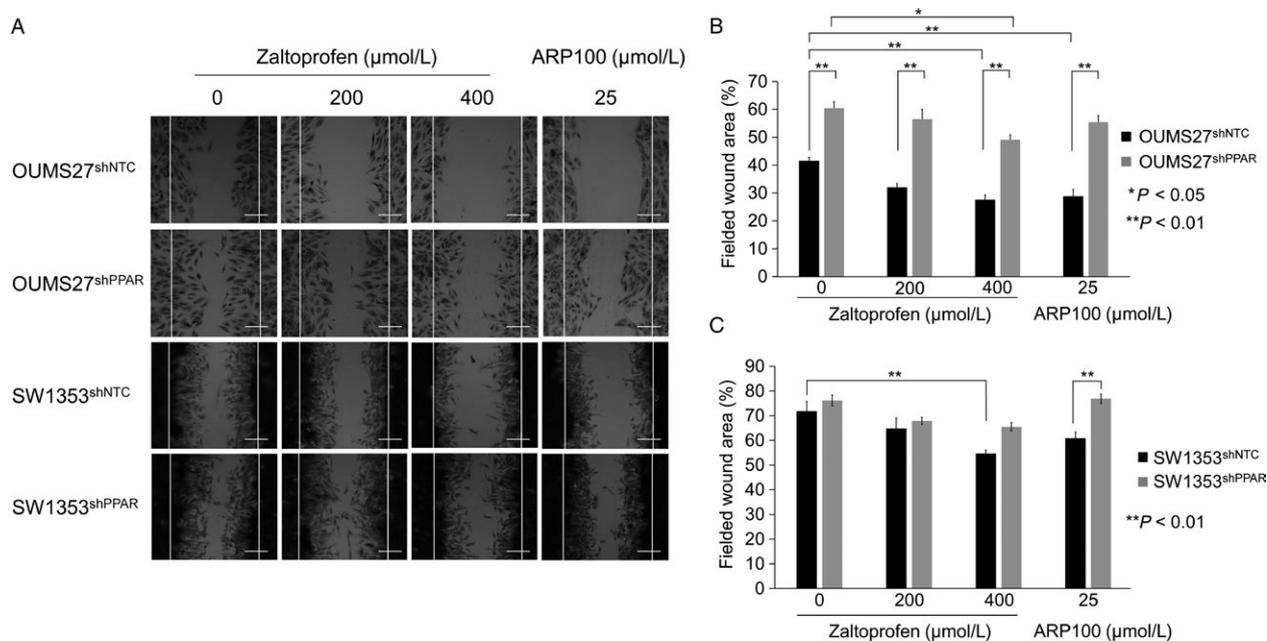


Figure 3. (A) Cell migration assay was carried out using PPAR γ knockdown cell lines, OUMS27^{shPPAR} and SW1353^{shPPAR}, and the control cell lines, OUMS27^{shNTC} and SW1353^{shNTC} with or without the treatment of zaltopfen. A selective inhibitor of MMP2, ARP100, was used at 25 $\mu\text{mol/L}$. Scale bar = 100 μm . (B, C) The fielded wound area (%) was calculated in OUMS27 (B) and SW1353 cells (C). Values represented as the mean \pm SEM. Asterisks denote a statistically significant difference (* $P < 0.05$ and ** $P < 0.01$).

in vitro cell migration assays and cell invasion assay using transwell inserts coated with a reconstituted basement membrane barrier (Matrigel). First, in vitro cell migration assays were performed and fielded wound area (%) was calculated (Fig. 3). Zaltopfen at a concentration of 400 $\mu\text{mol/L}$ and 200 $\mu\text{mol/L}$ significantly and partially inhibited cell migration of both OUMS27 and SW1353 cells, respectively (Fig. 3). Positive control ARP100 (25 $\mu\text{mol/L}$), a selective MMP2 inhibitor, also showed a significant inhibitory effect on cell migration (Fig. 3). PPAR γ silencing in chondrosarcoma cells, OUMS27^{shPPAR} and SW1353^{shPPAR}, exhibited a significant inhibition of cell migration even in the nontreated control and following cancelation of the inhibitory effect by zaltopfen, as well as ARP100 (Fig. 3). Second, we employed a Matrigel cell invasion assay with zaltopfen and ARP100. Zaltopfen significantly and dose-dependently inhibited cell invasion of SW1353 cells (Fig. 4A and B). ARP100 significantly inhibited cell invasion of SW1353 cells at a concentration of 25 $\mu\text{mol/L}$ (Fig. 4A and B). In the absence of zaltopfen or ARP100, PPAR γ silencing SW1353^{shPPAR} cells showed a significant increase in cell invasion when compared to control SW1353^{shNTC} cells, suggesting a PPAR γ -dependent inhibition of cell invasion, which was on the contrary to obtained data on cell migration (Fig. 4A and B). However, zaltopfen treatment significantly blocked invasion of SW1353^{shPPAR} cells, speculating the involvement of a PPAR γ -independent effect of zaltopfen (Fig. 4A and B).

Inhibitory effects of zaltopfen on MMP2 expression in chondrosarcoma cells

The data on the selective MMP2 inhibitor, ARP100, led us to investigate the relationship between MMP2 and zaltopfen. Gelatin zymography showed downregulation of MMP2 enzyme activities by zaltopfen at 400 $\mu\text{mol/L}$ in OUMS27^{shNTC} and SW1353^{shNTC} cells (Fig. 4C). However, in OUMS27^{shPPAR} and SW1353^{shPPAR} cells, the downregulation of MMP2 activities by zaltopfen was inhibited by PPAR γ silencing (Fig. 4C), thus, suggesting that a zaltopfen-induced PPAR γ activation could reduce MMP2 activities in chondrosarcoma cells.

A clinical case of chondrosarcoma treated with zaltopfen

We experienced a case of chondrosarcoma with unintentional zaltopfen treatment. (Fig. 5). A specimen was obtained from the patient with grade 2 chondrosarcoma before administration of zaltopfen (preadministration, Pre), and the microscopic data showed several nuclei with irregular shape and mitoses (Fig. 6A). In contrast, in the postadministration (Post) specimens, less nuclei with irregular shape or mitoses were found, and sporadic myxoid degeneration and necrotic portions were observed (Fig. 6A). Immunofluorescent staining of PPAR γ and

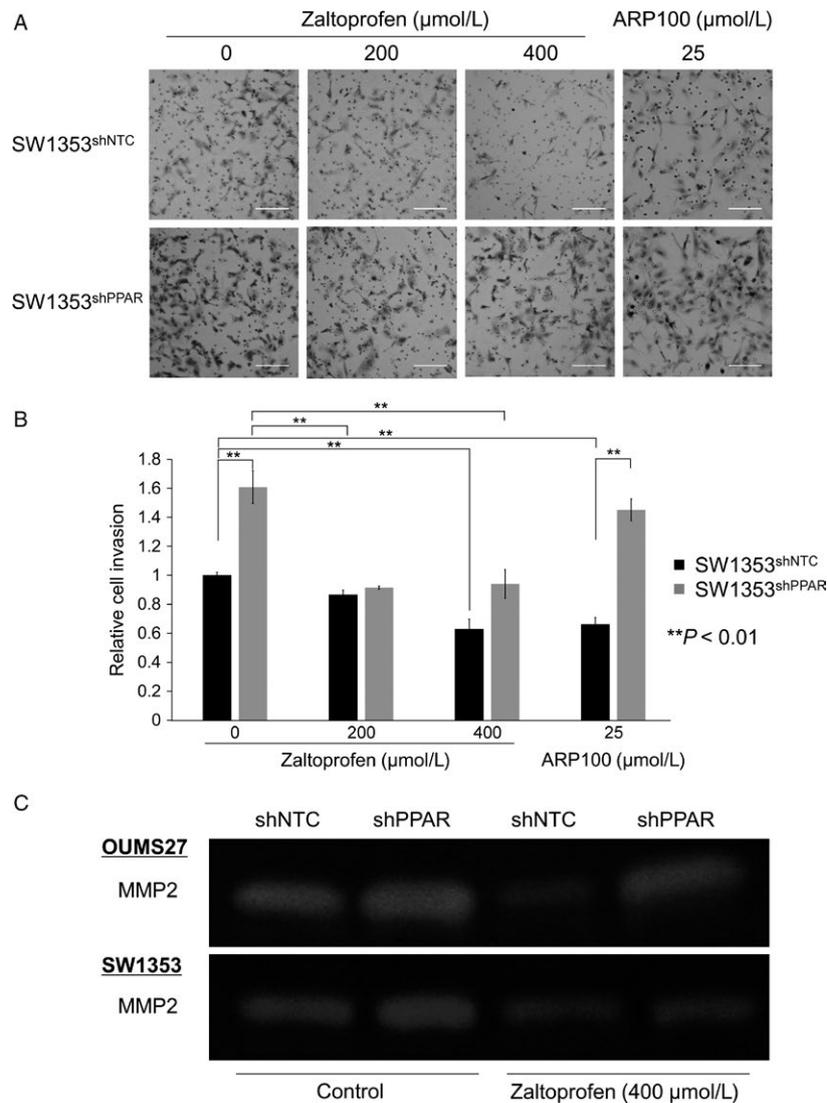


Figure 4. (A) Cell invasion assay was carried out using a PPAR γ knockdown cell line SW1353^{shPPAR} and the control cell line SW1353^{shNTC} with or without the treatment of zaltoprofen. A selective inhibitor of MMP2, ARP100, was used at 25 μ mol/L. Scale bar = 100 μ m. (B) Relative invaded cell number was calculated. Values represented as the mean \pm SEM. Asterisks denote a statistically significant difference (** P < 0.01). (C) Gelatin zymography. Effects of zaltoprofen (400 μ mol/L) on MMP2 activities in PPAR γ knockdown cell lines, OUMS27^{shPPAR} and SW1353^{shPPAR}, and the control cell lines, OUMS27^{shNTC} and SW1353^{shNTC}.

MMP2 demonstrated that the expression of PPAR γ was enhanced, but that of MMP2 was reduced, after the administration of zaltoprofen (Fig. 6B and C).

Discussion

In the present study, we found that zaltoprofen could dose-dependently upregulate PPAR γ activities and significantly, but mildly, induce PPAR γ expression in human chondrosarcoma OUMS27 and SW1353 cells (Fig. 1). Zaltoprofen inhibited cell viability and proliferation as well as cell migration in a PPAR γ -dependent manner in

OUMS27 and SW1353 cells (Figs. 2 and 3). Although cell invasion was significantly reduced by zaltoprofen treatment, PPAR γ -knockdown could not cancel the effects of zaltoprofen (Fig. 4A and B), suggesting two possibilities including an inhibitory action of PPAR γ against cell invasion and a PPAR γ -independent effect of zaltoprofen on cell invasion. In addition, the downregulation of MMP2 activity by zaltoprofen was dependent on PPAR γ expression (Fig. 4C).

It has been reported that PPAR γ is closely associated with malignant tumors [4]. Recent studies suggest that PPAR γ ligands can be useful anti-tumor agents in cancer

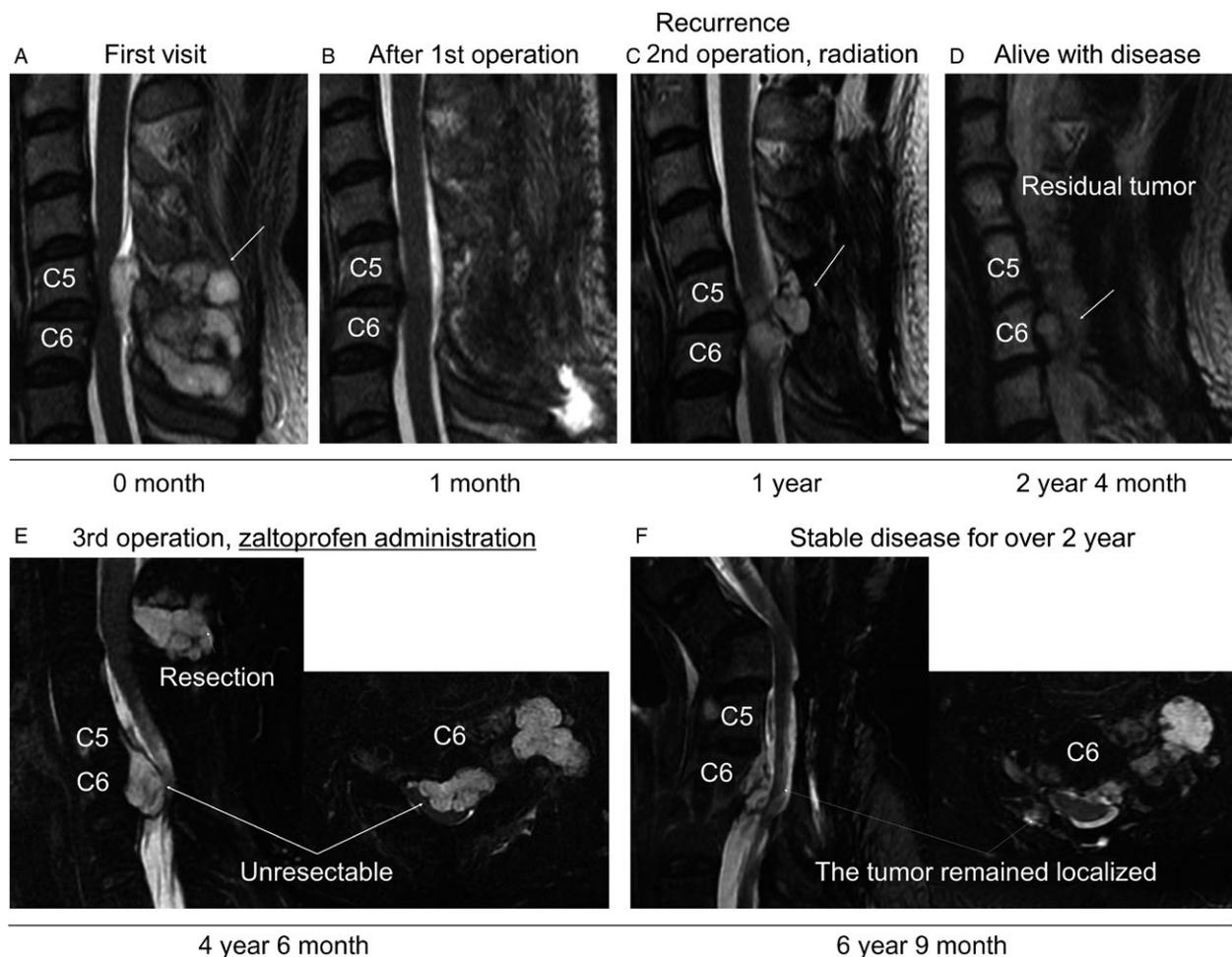


Figure 5. (A, B) Sagittal, MRI T2 weighted image of cervical spine at first visit on the initial hospital before (A) and after (B) the surgery. The tumor (arrow) was resected (1st operation), which markedly compressed the spinal cord. (C) Local recurrence was detected at C5-6 and tumor excision (2nd operation) was then performed after 1 year from the initial diagnosis. An arrow indicates the tumor. (D) The residual tumor (arrow). (E) Sagittal, MRI T2 fat suppression image. The tumors markedly enlarged and severe pain occurred in his left hand. The tumor of C2-4 was resected, while the tumor of C5-6 was unresectable. Zaltoprofen (240 mg/day) was then administered for the pain. (F) Before the final operation for severe cervical kyphosis, MRI T2 fat suppression image showed the localized tumor of C5-C6 remained.

cells [5, 6]. Endogenous and exogenous ligands for PPAR γ include fatty acids, eicosanoids, fibrates, thiazolidine derivatives, angiotensin receptor blockers, and NSAIDs [3, 8, 27]. Among them, NSAIDs such as indomethacin, diclofenac, oxaprozin, and zaltoprofen have been reported as potent ligands for PPAR γ [8] and anti-tumor agents against colorectal and lung cancers [28, 29]. This is the first report of anti-cancer effects of zaltoprofen against malignant musculoskeletal tumors. Activation of PPAR γ is reported to drive the positive feedback loop upregulating the *PPARG* gene *per se* via direct binding of PPAR γ /RXR α heterodimers to the *PPARG* promoter [30]. However, the involvement of other transcriptional activation factors for *PPARG*, including CEBPs and KLFs, is still unclear in the PPAR γ upregulation by zaltoprofen. It is also

interesting to examine whether zaltoprofen can selectively activate not only PPAR γ , but also PPAR α or PPAR δ , in terms of the anti-tumor effects of zaltoprofen. Further studies are required to elucidate the overall mechanisms of action of zaltoprofen on the anti-cancer activity.

MMP2, the 72 kDa gelatinase A/Type IV collagenase, plays an important role in the tumorigenic processes, such as tumor invasion and metastasis [12, 31]. Therefore, the inhibition of MMP2 activation has been suggested to be a potent strategy for the prevention of cancer cell metastasis [31]. To date, several MMP2 inhibitors have been developed for clinical use. In chondrosarcomas, the expression of MMP2 has been confirmed and demonstrated a significant correlation with the tumor histological grade linking to the prognosis [14]. Therefore, MMP2 could be a possible

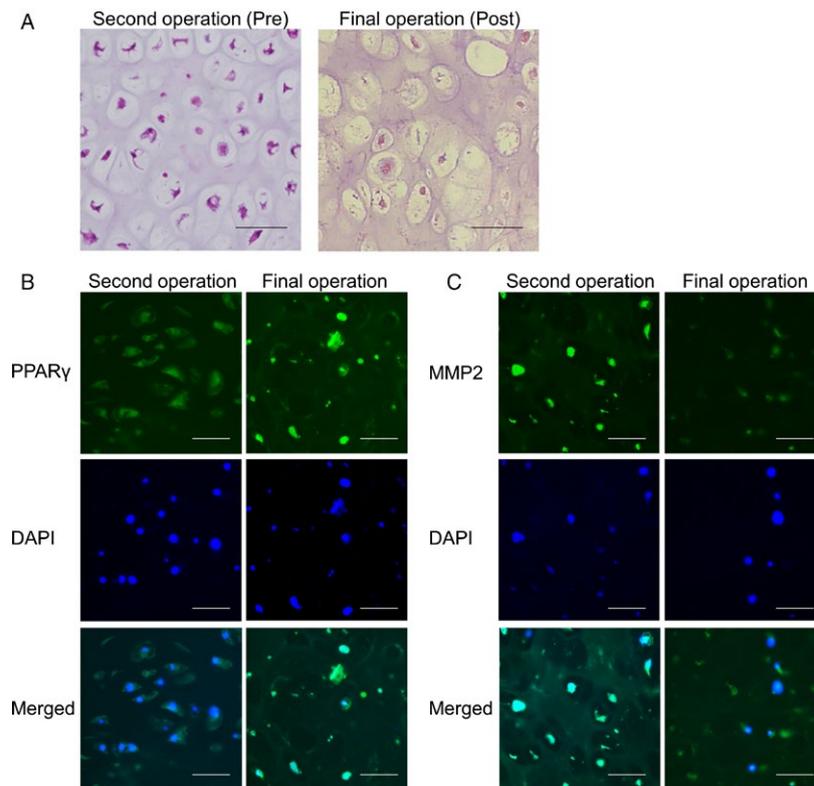


Figure 6. Histopathological findings of the tumors at the second operation (preadministration of zaltoprofen, Pre) and the final operation (postadministration of zaltoprofen, Post). Scale bar = 25 μ m. (A) HE staining. (B, C) Immunofluorescent stainings for PPAR γ (B) and MMP2 (C). DAPI, a nuclear staining.

therapeutic target to reduce local spread and invasion in chondrosarcomas. Lai et al. reported that alendronate, a second-generation bisphosphonate, inhibited MMP2 production and the invasion of a chondrosarcoma cell line [15]. Recent studies have also demonstrated that PPAR γ activation could inhibit cell migration and invasion through the downregulation of MMP2 in cancer cells [16, 17]. In this study, we first discovered the downregulation of MMP2 activity in a PPAR γ -dependent fashion by zaltoprofen.

We experienced a case of chondrosarcoma with unintentional zaltoprofen treatment. (Fig. 5). It is known that grade 2 chondrosarcoma progresses destructively and metastasizes. Local recurrence and metastasis rates have been reported to be 27% and 50%, respectively, in grade 2 chondrosarcoma, and 70% of chondrosarcoma cases with local recurrence have developed metastasis [32]. Interestingly, for approximately 2 years during administration of zaltoprofen, the tumor remained localized without distant metastasis even though the tumor was grade 2 chondrosarcoma. Histopathological findings showed upregulation of PPAR γ and downregulation of MMP2 after the administration of zaltoprofen in the patient (Fig. 6). We believe that a long-term administration of zaltoprofen could reduce the local spreading and metastatic abilities of the tumor cells.

In conclusion, this study showed that zaltoprofen activated PPAR γ and subsequently decreased MMP2 activity in human chondrosarcoma cells, thereby contributing to anti-tumor effects against cell viability, proliferation, migration, and invasion. A beneficial effect of zaltoprofen was also demonstrated in a patient with grade 2 chondrosarcoma. Therefore, these findings suggest that zaltoprofen could be a novel promising remedy against chondrosarcomas via PPAR γ activation and MMP2 inhibition. Future studies using orthotopic xenograft mouse models will be required to show the in vivo effect of zaltoprofen on chondrosarcomas, based on previous reports of osteosarcoma [33, 34], rhabdomyosarcoma [35], and undifferentiated pleomorphic sarcoma [36–38] with various anti-cancer drugs. In addition, large prospective cohort studies will be required to draw a final conclusion about the efficacy of zaltoprofen and to evaluate the effect of clinical applications in patients with chondrosarcoma.

Acknowledgment

The authors greatly appreciate Kazuhiro Tanaka M.D., for valuable contribution.

Conflict of Interest

None declared.

References

- Dorfman, H. D., and B. Czerniak. 1995. Bone cancers. *Cancer* 75:203–210.
- Lee, F. Y., H. J. Mankin, G. Fondren, M. C. Gebhardt, D. S. Springfield, A. E. Rosenberg, et al. 1999. Chondrosarcoma of bone: an assessment of outcome. *J. Bone Joint Surg. Am.* 81A:326–338.
- Kersten, S., B. Desvergne, and W. Wahli. 2000. Roles of PPARs in health and disease. *Nature* 405:421–424.
- Koeffler, H. P. 2003. Peroxisome proliferator-activated receptor gamma and cancers. *Clin. Cancer Res.* 9:1–9.
- Fröhlich, E., and R. Wahl. 2015. Chemotherapy and chemoprevention by thiazolidinediones. *Biomed. Res. Int.* 2015:845340.
- Robbins, G. T., and D. Nie. 2012. PPAR gamma, bioactive lipids, and cancer progression. *Front. Biosci. (Landmark Ed.)* 17:1816–1834.
- Nishida, K., T. Kunisada, Z. N. Shen, Y. Kadota, K. Hashizume, and T. Ozaki. 2008. Chondrosarcoma and peroxisome proliferator-activated receptor. *PPAR Res.* 2008:250568.
- Takeuchi, A., N. Yamamoto, H. Nishida, H. Kimura, H. Ikeda, and H. Tsuchiya. 2013. Complete necrosis of a giant cell tumor with high expression of PPAR γ : a case report. *Anticancer Res.* 33:2169–2174.
- Yamazaki, R., N. Kusunoki, T. Matsuzaki, S. Hashimoto, and S. Kawai. 2002. Nonsteroidal anti-inflammatory drugs induce apoptosis in association with activation of peroxisome proliferator-activated receptor gamma in rheumatoid synovial cells. *J. Pharmacol. Exp. Ther.* 302:18–25.
- Knopfová, L., and J. Smarda. 2010. The use of Cox-2 and PPAR γ signaling in anti-cancer therapies. *Exp. Ther. Med.* 1:257–264.
- Ghanqhas, P., S. Jain, C. Rana, and S. N. Sanyal. 2016. Chemopreventive action of non-steroidal anti-inflammatory drugs on the inflammatory pathways in colon cancer. *Biomed. Pharmacother.* 78:239–247.
- Egeblad, M., and Z. Werb. 2002. New functions for the matrix metalloproteinases in cancer progression. *Nat. Rev. Cancer* 2:161–174.
- Bjorklund, M., and E. Koivunen. 2005. Gelatinase-mediated migration and invasion of cancer cells. *Biochim. Biophys. Acta* 1755:37–69.
- Sakamoto, A., Y. Oda, Y. Iwamoto, and M. Tsuneyoshi. 1999. Expression of membrane type 1 matrix metalloproteinase, matrix metalloproteinase 2 and tissue inhibitor of metalloproteinase 2 in human cartilaginous tumors with special emphasis on mesenchymal and dedifferentiated chondrosarcoma. *J. Cancer Res. Clin. Oncol.* 125:541–548.
- Lai, T. J., S. F. Hsu, T. M. Li, H. C. Hsu, J. G. Lin, C. J. Hsu, et al. 2007. Alendronate inhibits cell invasion and MMP-2 secretion in human chondrosarcoma cell line. *Acta Pharmacol. Sin.* 28:1231–1235.
- Li, Y., D. W. Zhang, D. Q. Lin, and L. Q. Cao. 2015. Peroxisome proliferator-activated receptor- γ inhibits pancreatic cancer cell invasion and metastasis via regulating MMP-2 expression through PTEN. *Mol. Med. Rep.* 12:6255–6260.
- Chuang, C. H., C. L. Yeh, S. L. Yeh, E. S. Lin, L. Y. Wang, and Y. H. Wang. 2016. Quercetin metabolites inhibit MMP-2 expression in A549 lung cancer cells by PPAR- γ associated mechanisms. *J. Nutr. Biochem.* 33:45–53.
- Shah, B. S., M. Chen, T. Suzuki, M. Embree, K. Kong, C. H. Lee, et al. 2017. Pyrintegrin induces soft tissue formation by transplanted or endogenous cells. *Sci. Rep.* 27:36402.
- Noutsios, G. T., P. Silveyra, F. Bhatti, and J. Floros. 2013. Exon B of human surfactant protein A2 mRNA, alone or within its surrounding sequences, interacts with 14-3-3; role of cis-elements and secondary structure. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 304: 722–735.
- Peters, A. A., P. T. Simpson, J. J. Bassett, J. M. Lee, L. Da Silva, L. E. Reid, et al. 2012. Calcium channel TRPV6 as a potential therapeutic target in estrogen receptor-negative breast cancer. *Mol. Cancer Ther.* 11:2158–2168.
- Grishagin, I. V. 2015. Automatic cell counting with Image. *J. Anal. Biochem.* 473:63–65.
- Yarrow, J. C., Z. E. Perlman, N. J. Westwood, and T. J. Mitchison. 2004. A high-throughput cell migration assay using scratch wound healing, a comparison of image based readout methods. *BMC Biotechnol.* 4:21.
- Silva Nunes, J. P., and A. A. Dias. 2017. ImageJ macros for the user-friendly analysis of soft-agar and wound-healing assays. *Biotechniques* 62:175–179.
- Takeuchi, A., Y. Yamamoto, S. Munesue, A. Harashima, T. Watanabe, H. Yonekura, et al. 2013. Low molecular weight heparin suppresses receptor for advanced glycation end products-mediated expression of malignant phenotype in human fibrosarcoma cells. *Cancer Sci.* 104:740–749.
- Sato, H., T. Takino, Y. Okada, J. Cao, A. Shinagawa, E. Yamamoto, et al. 1994. A matrix metalloproteinase expressed on the surface of invasive tumour cells. *Nature* 370:61–65.
- Kanda, Y. 2013. Investigation of the freely-available easy-to-use software “EZ R” (Easy R) for medical statistics. *Bone Marrow Transplant.* 48:452–458.

27. Kroker, A. J., and J. B. Bruning. 2015. Review of the structural and dynamic mechanisms of PPAR γ partial agonism. *PPAR Res.* 2015:816856.
28. Moon, C. M., J. H. Kwon, J. S. Kim, S. H. Oh, K. Jin Lee, J. J. Park, et al. 2014. Nonsteroidal anti-inflammatory drugs suppress cancer stem cells via inhibiting PTGS2 (cyclooxygenase 2) and NOTCH/HES1 and activating PPAR γ in colorectal cancer. *Int. J. Cancer* 134:519–529.
29. Kato, T., H. Fujino, S. Oyama, T. Kawashima, and T. Murayama. 2011. Indomethacin induces cellular morphological change and migration via epithelial-mesenchymal transition in A549 human lung cancer cells: a novel cyclooxygenase-inhibition-independent effect. *Biochem. Pharmacol.* 82:1781–1791.
30. Wakabayashi, K., M. Okamura, S. Tsutsumi, N. S. Nishikawa, T. Tanaka, I. Sakakibara, et al. 2009. The peroxisome proliferator-activated receptor gamma/retinoid X receptor alpha heterodimer targets the histone modification enzyme PR-Set7/Setd8 gene and regulates adipogenesis through a positive feedback loop. *Mol. Cell. Biol.* 29:3544–3555.
31. Coussens, L. M., B. Fingleton, and L. M. Matrisian. 2002. Matrix metalloproteinase inhibitors and cancer: trials and tribulations. *Science* 295:2387–2392.
32. Fiorenza, F., A. Abudu, R. J. Grimer, S. R. Carter, R. M. Tillman, K. Ayoub, et al. 2002. Risk factors for survival and local control in chondrosarcoma of bone. *J. Bone Joint Surg. Br.* 84B:93–99.
33. Igarashi, K., K. Kawaguchi, T. Murakami, T. Kiyuna, K. Miyake, S. D. Nelson, et al. 2017. Intra-arterial administration of tumor-targeting Salmonella typhimurium A1-R regresses a cisplatin-resistant relapsed osteosarcoma in a patient-derived orthotopic xenograft (PDOX) mouse model. *Cell Cycle* 16:1164–1170.
34. Igarashi, K., T. Murakami, K. Kawaguchi, T. Kiyuna, K. Miyake, Y. Zhang, et al. 2017. A patient-derived orthotopic xenograft (PDOX) mouse model of a cisplatin-resistant osteosarcoma lung metastasis that was sensitive to temozolomide and trabectedin: implications for precision oncology. *Oncotarget* 8:62111–62119.
35. Igarashi, K., K. Kawaguchi, T. Kiyuna, T. Murakami, S. Miwa, S. D. Nelson, et al. 2017. Patient-derived orthotopic xenograft (PDOX) mouse model of adult rhabdomyosarcoma invades and recurs after resection in contrast to the subcutaneous ectopic model. *Cell Cycle* 16:91–94.
36. Igarashi, K., K. Kawaguchi, T. Murakami, T. Kiyuna, K. Miyake, N. Yamamoto, et al. 2017. A novel anionic-phosphate-platinum complex effectively targets an undifferentiated pleomorphic sarcoma better than cisplatin and doxorubicin in a patient-derived orthotopic xenograft (PDOX). *Oncotarget* 8:63353–63359.
37. Igarashi, K., K. Kawaguchi, T. Murakami, T. Kiyuna, K. Miyake, A. S. Singh, et al. 2017. High efficacy of pazopanib on an undifferentiated spindle-cell sarcoma resistant to first-line therapy is identified with a patient-derived orthotopic xenograft (PDOX) nude mouse model. *J. Cell. Biochem.* 118:2739–2743.
38. Igarashi, K., K. Kawaguchi, T. Kiyuna, T. Murakami, S. Miwa, S. D. Nelson, et al. 2017. Temozolomide combined with irinotecan caused regression in an adult pleomorphic rhabdomyosarcoma patient-derived orthotopic xenograft (PDOX) nude-mouse model. *Oncotarget* 8:75874–75880.