

# A New Synthetic Method for Dipeptides Containing $\alpha,\beta$ -Didehydroamino Acids Utilizing an $\alpha$ -Tosylglycine Residue

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The reaction of peptides containing an *N*-Boc- or *N*-Z- $\alpha$ -tosylglycine residue at the *N*-terminus with a variety of excess nitro compounds under basic conditions to give the nitro compound-adducts in good yields, followed by the elimination of nitrous acid from the adducts to afford the corresponding dipeptide containing  $\alpha,\beta$ -didehydroamino acids with (*Z*)-configuration predominantly. Treatment of the same starting materials with aldehydes in the presence of base and tributylphosphine also gave a dipeptide containing  $\alpha,\beta$ -didehydroamino acids with high (*Z*)-selectivity in satisfactory yields. The present method can be successfully applied to the synthesis of a protected leucine enkephalin analog.

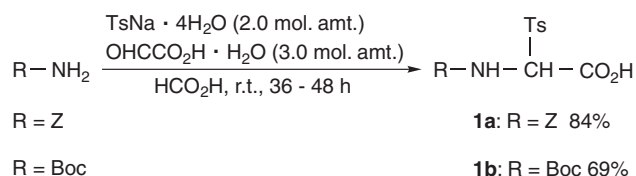
$\alpha,\beta$ -Didehydroamino acid residues are common components of naturally occurring peptides that remarkably affect the physiological activity of peptides,<sup>1</sup> as well as their conformations,<sup>2</sup> and their resistance to enzyme-catalyzed degradation.<sup>3</sup> Consequently, there are many reports related to methods for the syntheses of peptides containing  $\alpha,\beta$ -didehydroamino acid residues.<sup>4</sup>

In previous papers, we already reported that *N*-*t*-butoxycarbonyl (Boc)- or *N*-benzyloxycarbonyl (*Z*)- $\alpha$ -tosylglycine ester was reacted with either a variety of nitroalkanes in the presence of base<sup>5a</sup> or various kinds of aldehydes in the presence of base and tributyl phosphine<sup>5b</sup> to afford the corresponding  $\alpha,\beta$ -didehydroamino acid derivatives with high (*Z*)-selectivity in good yields. The latter procedure was successfully applied to the syntheses of optically active 4-hydroxyprolines.<sup>5b</sup>

In the light of our results mentioned above, we now wish to report a convenient method for the preparation of dipeptides **17–31** containing an  $\alpha,\beta$ -didehydroamino acid residue starting from dipeptides containing  $\alpha$ -tosylglycine residue at *N*-termi-

nus **2**. *Z*- and Boc- $\alpha$ -tosylglycines, **1a** and **1b**, were prepared by the reaction of sodium sulfinate, glyoxylic acid, and *Z*- or Boc-carbamate in 99% or 50% formic acid in 84% and 69% yields, respectively (Scheme 1).

First, dipeptide **2a** was obtained as a mixture of diastereomers in a 71% yield by the coupling reaction of methyl *L*-leucinate and *N*-Z- $\alpha$ -tosylglycine (**1a**) using the dicyclohexylcarbodiimide (DCC)-*N*-hydroxysuccinimide (HONSu) method. Dipeptides **2b–d** were obtained in moderate yields, respectively (Entries 2–4 in Table 1). Similarly, *N*-Boc protected dipeptides



Scheme 1. Preparation of *Z*- and Boc- $\alpha$ -tosylglycine.

Table 1. Preparation of Dipeptides **2a–i** Containing Tosylglycine Residue

Entry	R	AA	Additive	Product/%	Ratio of diastereomer <sup>a)</sup>
1	Z	Leu	HONSu	<b>2a</b> 71	56/44
2	Z	Ala	HOBt	<b>2b</b> 51	55/45
3	Z	Gly	HOBt	<b>2c</b> 65	
4	Z	Val	HOBt	<b>2d</b> 56	56/44
5	Boc	Leu	HOBt	<b>2e</b> 78	67/33
6	Boc	Ala	HOBt	<b>2f</b> 74	54/46
7	Boc	Gly	HOBt	<b>2g</b> 76	
8	Boc	Val	HONSu	<b>2h</b> 45	53/47
9	Boc	Ile	HOBt	<b>2i</b> 62	68/32

a) Determined by NMR spectrum.



Table 4. Preparation of Dipeptides Containing Dehydroamino Acid Residue

$$\begin{array}{ccc}
 \text{R-NH-CH-CO-AA-OMe} & & \text{R-NH} & \text{CO-AA-OMe} \\
 | & & | & | \\
 \text{R}^1-\text{C}-\text{R}^2 & \xrightarrow[\text{r.t., 1-2 d, CH}_2\text{Cl}_2]{\text{DBU (2.0-3.0 mol. amt.)}} & \text{C} & \\
 | & & | & | \\
 \text{NO}_2 & & \text{R}^1 & \text{R}^2 \\
 \mathbf{3-6, 9-16} & & \mathbf{17-28}
 \end{array}$$

Entry	Substrate	R	R <sup>1</sup>	R <sup>2</sup>	AA	Product	Yield/%	E/Z <sup>a)</sup>
1	<b>3</b>	Z	Me	Me	Leu	<b>17</b>	70	—
2	<b>4</b>	Z	Me	H	Leu	<b>18</b>	48	0/100
3	<b>5</b>	Z	Me	Me	Ala	<b>19</b>	53	—
4	<b>6</b>	Z	Me	Me	Gly	<b>20</b>	47	—
5	<b>9</b>	Boc	Me	Me	Leu	<b>21</b>	84	—
6	<b>10</b>	Boc	Me	H	Leu	<b>22</b>	66	0/100
7	<b>11</b>	Boc	Ph	H	Leu	<b>23</b>	34	0/100
8 <sup>b)</sup>	<b>11</b>	Boc	Ph	H	Leu	<b>23</b>	62	0/100
9	<b>12</b>	Boc	Et	Me	Leu	<b>24</b>	55	14/86
10	<b>13</b>	Boc	Me	Me	Ala	<b>25</b>	67	—
11	<b>14</b>	Boc	Me	Me	Gly	<b>26</b>	81	—
12	<b>15</b>	Boc	Me	Me	Val	<b>27</b>	70	—
13	<b>16</b>	Boc	Me	Me	Ile	<b>28</b>	91	—

a) Determined by NOE measurement. b) 7.0 equimolar amounts of 1,4-diazabicyclo[2.2.2]octane (DABCO) were used instead of DBU.

Table 5. Reaction of Dipeptides Containing Tosylglycine Residue with Aldehydes

$$\begin{array}{ccc}
 \text{Ts} & & \text{BocNH} & \text{CO-AA-OMe} \\
 | & & | & | \\
 \text{BocNH-CH-CO-AA-OMe} & \xrightarrow[\text{N}_2, 0^\circ\text{C} \rightarrow \text{r.t., 1.5-2.5 h, THF}]{\text{RCHO (4.0 mol. amt.), Bu}_3\text{P (1.5 mol. amt.)}, \text{DBU (1.2 mol. amt.)}} & \text{C} & \\
 \mathbf{2e, 2g} & & | & | \\
 & & \text{R}' & \text{H}
 \end{array}$$

Entry	Substrate	AA	RCHO	R'	Product	Total yield/%	E/Z
1	<b>2e</b>	Leu	MeCHO	Me	(Z)- <b>22</b> (E)- <b>22'</b>	89	5/95 <sup>a)</sup>
2	<b>2e</b>	Leu	PhCHO	Ph	<b>23</b>	64	0/100 <sup>a)</sup>
3	<b>2e</b>	Leu	EtCHO	Et	(Z)- <b>29</b> (E)- <b>29'</b>	97	10/90 <sup>a)</sup>
4	<b>2g</b>	Gly	MeCHO	Me	(Z)- <b>30</b> (E)- <b>30'</b>	82	6/94 <sup>b)</sup>
5	<b>2g</b>	Gly	EtCHO	Et	(Z)- <b>31</b> (E)- <b>31'</b>	quant.	6/94 <sup>a)</sup>

a) Determined by NOE measurement. b) The major product was assigned to be (Z)-isomer by comparison with the authentic sample.

nate (**28**) in 91% yield utilizing 2 molar amounts of DBU in dichloromethane at room temperature for 2 days under nitrogen. Although the yield in Entry 7 was not so high, it was improved by using 7 molar amounts of 1,4-diazabicyclo[2.2.2]octane (DABCO) instead of DBU (Entry 8 in Table 4). In the cases of Entries 2 and 6–8 (Table 4), the products **18**, **22**, and **23** were obtained with only (Z)-configuration. The elimination reaction of the nitrous acid proceeded smoothly with high (Z)-selectivity to afford the products **17–28** in good yields (Table 4). The stereochemistry was assigned by NOE measurement.

We furthermore examined the direct conversion of a dipeptide containing an  $\alpha$ -tosylglycine residue to a dipeptide containing an  $\alpha,\beta$ -didehydroamino acid residue. Namely, compounds **2e** and **2g** were treated with 4 equimolar amounts of aldehyde in the presence of 1.5 molar amounts of tributylphosphine

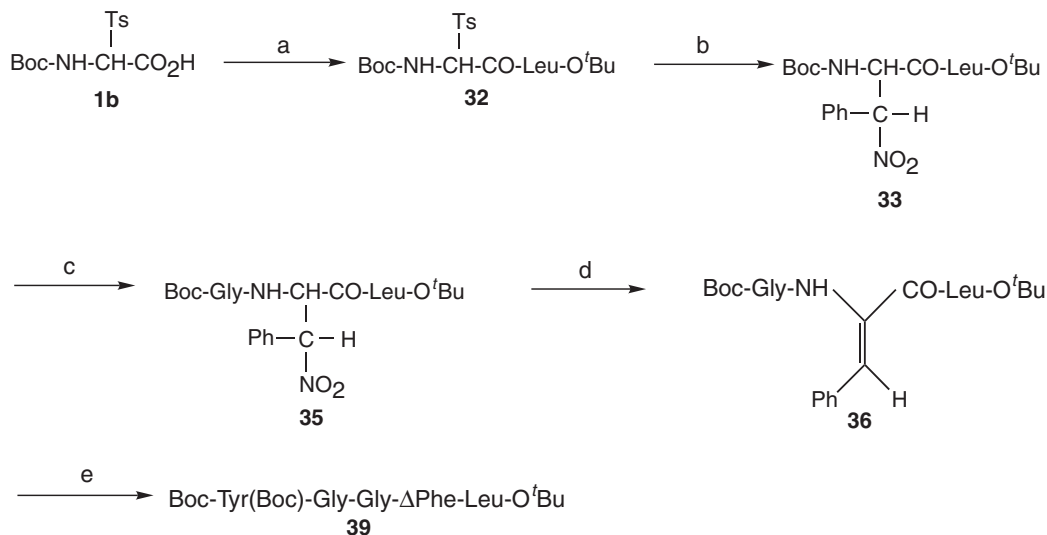
(Bu<sub>3</sub>P) and 1.2 molar amounts of DBU. Accordingly, the reaction proceeded through a Wittig-type reaction<sup>5b</sup> to afford dipeptides; the (Z)-isomer of **22**, **23**, and **29–31** as major products in good yields accompanied by a small amount of (E)-isomer, except for Entry 2 (Table 5). The results are summarized in Table 5. The stereochemistry of the major product **30** in Entry 4 was assigned to be (Z)-configuration by comparison with the authentic sample prepared by the coupling reaction of (Z)-2-(*t*-butoxycarbonylamino)-2-butenic acid and methyl glycinate.

Moreover, it was found that treatment of isolable (E)-isomer **22'** with a catalytic amount of iodine in dichloromethane at room temperature resulted in the isomerization to the corresponding (Z)-isomer in an 83% yield (Entry 1 in Table 6). In a similar way, **29'**, **30'**, and **31'** were also isomerized to their (Z)-isomers in good yields (Table 6).

Table 6. Isomerization of (*E*)- to (*Z*)-isomer by Iodine

**22', 29', 30', 31'**

Entry	Substrate	R	AA	Yield/%	<i>E/Z</i>
1	<b>22'</b>	Me	Leu	83	5/95
2	<b>29'</b>	Et	Leu	90	20/80
3	<b>30'</b>	Me	Gly	83	5/95
4	<b>31'</b>	Et	Gly	80	2/98



Scheme 2. Synthesis of protected  $\Delta$ Phe<sup>4</sup>-leucine enkephalin analog. a) H-Leu-O<sup>t</sup>Bu (1.0 mol. amt.), DCC (1 mol. amt.), HOBT (1 mol. amt.) in THF, r.t., 23 h. **32** 77%. b) PhCH<sub>2</sub>NO<sub>2</sub> (2 mol. amt.), KO<sup>t</sup>Bu (2 mol. amt.) in THF, -40 °C → r.t., **33** 70%. c) 1) CH<sub>3</sub>COCl (17 mol. amt.), MeOH (17 mol. amt.) in AcOEt, r.t., 8 h. HCl salt **34** quant. 2) Boc-Gly-OH (1.1 mol. amt.), IBCF (1.0 mol. amt.), NMM (1.1 mol. amt.) in THF, -15 °C, 8 min; **34** (1 mol. amt.), NMM (1.1 mol. amt.) in THF/DMF (2/1), -15 °C to r.t., 5 h. **35** 76%. d) DABCO (7 mol. amt.) in CH<sub>2</sub>Cl<sub>2</sub>, r.t., 8 h. **36** 81%. e) 1) CH<sub>3</sub>COCl (17 mol. amt.), MeOH (17 mol. amt.) in AcOEt, r.t., 8 h. HCl salt **37** quant. 2) Boc-Tyr(Boc)-Gly-OH (**38**) (1 mol. amt.), IBCF (1.1 mol. amt.), NMM (1.1 mol. amt.) in THF, -15 °C, 15 min; **37** (1 mol. amt.), NMM (1.1 mol. amt.) in THF/DMF (2/1), -15 °C to r.t., overnight. **39** 72%. DCC = dicyclohexylcarbodiimide, HOBT = 1-hydroxybenzotriazole, IBCF = isobutyl chloroformate, NMM = 4-methylmorpholine, DABCO = 1,4-diazabicyclo[2.2.2]octane.

As nitro compound-adducts (**3–6** and **9–16**) could be easily converted to the corresponding dipeptide containing an  $\alpha,\beta$ -dehydroamino acid residue under basic conditions in moderate to good yields (Table 4), the present method was applied to the synthesis of protected analog **39** of the opioid peptide, leucine-enkephalin, according to Scheme 2.<sup>7</sup> Initially, *N*-Boc- $\alpha$ -tosylglycine (**1b**) was coupled with *t*-butyl L-leucinate using the DCC–HOBT method to afford the dipeptide **32** in 77% yield, which was treated with (nitromethyl)benzene in the presence of 2 molar amounts of KO<sup>t</sup>Bu to give *t*-butyl *N*-Boc-( $\beta$ -nitro)-phenylalanyl-L-leucinate **33** in 70% yield. Next, selective deprotection of the *N*-Boc protecting group in compound **33** was examined under various acidic conditions. Eventually, it was found that *N*-deprotection of **33** with hydrogen chloride in MeOH<sup>40</sup> gave the hydrogen chloride salt **34** in quantitative yield. Compound **34** was mediated with *N*-Boc-glycine by

means of the mixed anhydride method using isobutyl chloroformate to afford the tripeptide **35** in 76% yield, which was treated with 2 molar amounts of DBU in dichloromethane at room temperature for 2 days under nitrogen to afford the desired Boc-Gly- $\Delta$ Phe-Leu-O<sup>t</sup>Bu (**36**) with (*Z*)-configuration in only 34% yield. The conditions used for the preparation of compound **23** (Entry 8 in Table 4) were then applied. Namely, the treatment of **35** with 7 molar amounts of DABCO in dichloromethane at room temperature for 8 h afforded **36** in 81% yield. The stereochemistry of the  $\alpha,\beta$ -dehydrophenylalanine residue of **36** were determined by NOE measurement. Finally, after *N*-deprotection of **36** with hydrogen chloride under the same reaction conditions as described for compound **33** the resulting amine hydrogen chloride salt **37** was condensed with *N,O*-diBoc-L-tyrosylglycine (**38**) using the mixed anhydride method to give the desired protected leucine-enkephalin analog

(**39**) in 72% yield.

As mentioned above, the  $\alpha$ -tosylglycine residue proved to be a very useful precursor for the  $\alpha,\beta$ -didehydroamino acid residue. The utility of the present method was demonstrated by the synthesis of the protected leucine-enkephalin analog.

### Experimental

All of the melting points were determined with a micro melting apparatus (Yanagimoto Seisakusho) and were uncorrected. The  $^1\text{H}$ NMR, IR, and MS spectra were recorded on JEOL JNM-LA 400FT (400 MHz) and LA 300FT (300 MHz) NMR spectrometers, a JASCO FT/IR-230 infrared spectrometer, and a JEOL SX-102A mass spectrometer, respectively. The chemical shifts of the NMR spectra are reported in  $\delta$  relative to TMS as an internal standard. All of the solvents were distilled and stored over a drying agent. Thin-layer chromatography (TLC) and flash-column chromatography were performed using Merck's silica-gel 60 PF<sub>254</sub> (Art. 7749) and Cica-Merck's silica-gel 60 (No. 9385-5B), respectively.

***N*-Z- $\alpha$ -Tosylglycine (1a).** A solution of benzyl carbamate (1.51 g, 17 mmol), glyoxylic acid monohydrate (1.84 g, 20 mmol), and sodium *p*-toluenesulfonate tetrahydrate (7.50 g, 30 mmol) in 99% formic acid (9 mL) was allowed to stir for 48 h at room temperature under air. The solution was then poured into a large amount of water. The crystalline product was collected, washed with water, and dried to afford the desired product **1a** in 84% yield (3.17 g). Mp 81.0–82.0 °C (ethyl acetate–hexane); IR (KBr) 3289, 2372, 2338, 1687, 1655, 1552, 1436, 1300, 1258, 1110, 1062, 976, 757, 698 cm<sup>-1</sup>;  $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.43 (s, 3H), 5.01 (s, 2H), 5.58 (d, *J* = 9.7 Hz, 1H), 6.26 (d, *J* = 9.9 Hz, 1H), 7.25–7.36 (m, 7H), 7.80 (d, *J* = 8.3 Hz, 2H). One proton of the carboxylic acid was not assigned. **1a** was converted to the corresponding methyl ester by means of diazomethane for elementary analysis. Mp 127.0–127.5 °C (ethyl acetate–hexane). Found: C, 57.34; H, 5.05; N, 3.60%. Calcd for C<sub>18</sub>H<sub>19</sub>NO<sub>6</sub>S: C, 57.28; H, 5.07; N, 3.71%.

***N*-Boc- $\alpha$ -Tosylglycine (1b).** A solution of *t*-butyl carbamate (3.98 g, 34 mmol), glyoxylic acid monohydrate (6.26 g, 68 mmol), and *p*-toluenesulfonate tetrahydrate (25.50 g, 102 mmol) in 50% aqueous formic acid (15 mL) was allowed to stir for 36 h at room temperature under air. The solution was then poured into 800 mL of water. The crystalline product was collected, washed with water, and dried to afford the desired product **1b** in 69% yield (7.73 g). Mp 142.0–144.0 °C (ethyl acetate–hexane); IR (KBr) 3354, 2960, 1727, 1698, 1595, 1517, 1467, 1429, 1366, 1325, 1227, 1162, 1146, 1082, 1056, 923, 854, 815, 778, 659 cm<sup>-1</sup>;  $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.31 (s, 9H), 2.44 (s, 3H), 3.64 (br, 1H), 5.66 (d, *J* = 9.8 Hz, 1H), 5.81 (d, *J* = 9.8 Hz, 1H), 7.36 (d, *J* = 8.1 Hz, 2H), 7.83 (d, *J* = 8.1 Hz, 2H). One proton of the carboxylic acid was not assigned. Found: C, 51.09; H, 5.86; N, 4.09%. Calcd for C<sub>14</sub>H<sub>19</sub>NO<sub>6</sub>S: C, 51.05; H, 5.81; N, 4.25%.

**A General Procedure for the Preparation of Dipeptides Containing a Tosylglycine Residue (2a–i).** To a suspension of methyl leucinate hydrochloride (927 mg, 5.10 mmol) in dry THF (15 mL) was added a solution of triethylamine (526 mg, 5.20 mmol) in dry THF (2 mL) at 0 °C under nitrogen. After stirring for 2 h at room temperature, **1a** (151 g, 4.00 mmol), DCC (825 mg, 4.00 mmol), and *N*-hydroxysuccinimide (460 mg, 4.00 mmol) were added. The mixture was stirred for 2 d at room temperature under air. After filtration, the filtrate was concentrated in vacuo to give a residue, which was redissolved into ethyl acetate.

The insoluble substances were filtered off, and the filtrate was successively washed with 10% citric acid, 10% NaHCO<sub>3</sub>, and brine, dried over anhydrous MgSO<sub>4</sub>, and evaporated under reduced pressure to afford methyl *N*-Z- $\alpha$ -tosylglycyl-L-leucinate (**2a**) as an inseparable mixture of diastereomers in 71% yield (1.39 g). Mp 105.0–106.0 °C (ethyl acetate–hexane); IR (KBr) 3284, 3033, 2957, 2871, 1737, 1717, 1669, 1628, 1597, 1523, 1456, 1328, 1232, 1181, 1147, 1083, 1058, 816, 771, 741, 698, 670 cm<sup>-1</sup>;  $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>) (major product = M)  $\delta$  0.97 (d, *J* = 6.1 Hz, 6H), 1.64–1.73 (m, 2H), 1.83–1.94 (m, 1H), 2.40 (s, 3H), 3.74 (s, 3H), 4.69 (dd, *J* = 7.6, 7.6 Hz, 1H), 4.86 (d, *J* = 12.2 Hz, 1H), 4.94–4.97 (m, 1H), 5.65 (d, *J* = 9.3 Hz, 1H), 6.20 (d, *J* = 8.8 Hz, 1H), 7.12 (d, *J* = 7.8 Hz, 1H), 7.18–7.38 (m, 7H), 7.74 (d, *J* = 8.8 Hz, 2H). (Minor product = m)  $\delta$  0.90–0.94 (m, 6H), 1.64–1.73 (m, 2H), 1.83–1.94 (m, 1H), 2.40 (s, 3H), 3.79 (s, 3H), 4.60–4.68 (m, 1H), 4.96 (d, *J* = 12.4 Hz, 1H), 5.04 (d, *J* = 12.2 Hz, 1H), 5.68 (d, *J* = 10.5 Hz, 1H), 6.15 (d, *J* = 8.8 Hz, 1H), 7.18–7.38 (m, 7H), 7.41 (d, *J* = 7.3 Hz, 1H), 7.76 (d, *J* = 8.5 Hz, 2H). Found: C, 58.97; H, 6.22; N, 5.85%. Calcd for C<sub>24</sub>H<sub>30</sub>N<sub>2</sub>O<sub>7</sub>S: C, 58.76; H, 6.16; N, 5.71%.

The physical and spectral data of compounds **2b–2i** are shown in the following. The products, except for **2c** and **2g**, were obtained as an inseparable mixture of diastereomers.

**Methyl *N*-Z- $\alpha$ -Tosylglycyl-L-alanine (2b):** Mp 150.5–151.0 °C (ethyl acetate); IR (KBr) 3343, 3278, 3033, 2953, 1738, 1663, 1597, 1524, 1454, 1380, 1356, 1324, 1305, 1265, 1227, 1180, 1146, 1083, 1056, 982, 912, 833, 818, 772, 741, 698, 669 cm<sup>-1</sup>;  $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  1.47 (d, *J* = 7.3 Hz, 3H), 2.41 (s, 3H), 3.81 (s, 3H), 4.54–4.68 (m, 1H), 4.93–5.03 (m, 2H), 5.62 (d, *J* = 8.5 Hz, 1H), 7.22–7.35 (m, 8H), 7.76 (d, *J* = 8.3 Hz, 2H). (m)  $\delta$  1.51 (d, *J* = 7.3 Hz, 3H), 2.41 (s, 3H), 3.76 (s, 3H), 4.54–4.68 (m, 1H), 4.89–5.00 (m, 2H), 5.62 (d, *J* = 8.5 Hz, 1H), 6.16 (d, *J* = 8.9 Hz, 1H), 7.22–7.35 (m, 7H), 7.52 (d, *J* = 6.1 Hz, 1H), 7.73 (d, *J* = 8.5 Hz, 2H). Found: C, 54.07; H, 6.82; N, 6.31%. Calcd for C<sub>20</sub>H<sub>30</sub>N<sub>2</sub>O<sub>7</sub>S: C, 54.28; H, 6.83; N, 6.33%.

**Methyl *N*-Z- $\alpha$ -Tosylglycyl-L-alanine (2c):** Mp 153.0–154.0 °C (ethyl acetate); IR (KBr) 3312, 3283, 3033, 2947, 2853, 1739, 1704, 1674, 1658, 1597, 1523, 1439, 1386, 1323, 1268, 1233, 1181, 1135, 1107, 1083, 1051, 983, 873, 742, 699 cm<sup>-1</sup>;  $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.41 (s, 3H), 3.79 (s, 3H), 4.09 (dd, *J* = 5.7, 18.1 Hz, 1H), 4.20 (dd, *J* = 5.7, 18.1 Hz, 1H), 4.93 (d, *J* = 12.2 Hz, 1H), 4.99 (d, *J* = 12.4 Hz, 1H), 5.65 (d, *J* = 8.8 Hz, 1H), 6.14 (d, *J* = 8.8 Hz, 1H), 7.23–7.35 (m, 7H), 7.78 (d, *J* = 8.3 Hz, 2H). Found: C, 55.25; H, 5.12; N, 6.46%. Calcd for C<sub>20</sub>H<sub>22</sub>N<sub>2</sub>O<sub>7</sub>S: C, 55.29; H, 5.10; N, 6.45%.

**Methyl *N*-Z- $\alpha$ -Tosylglycyl-L-valinate (2d):** Mp 117.0–118.0 °C (ethyl acetate–diethyl ether–hexane); IR (KBr) 3338, 3270, 3065, 2962, 1745, 1725, 1672, 1598, 1534, 1455, 1438, 1375, 1327, 1265, 1234, 1182, 1147, 1084, 1020, 840, 815, 737, 714, 697, 669 cm<sup>-1</sup>;  $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  0.96 (dd, *J* = 2.1, 7.0 Hz, 6H), 2.18–2.34 (m, 1H), 2.41 (s, 3H), 3.80 (s, 3H), 4.65 (dd, *J* = 4.6, 8.8 Hz, 1H), 4.95 (d, *J* = 11.2 Hz, 1H), 5.03 (d, *J* = 12.2 Hz, 1H), 5.65 (d, *J* = 9.0 Hz, 1H), 6.17 (d, *J* = 8.8 Hz, 1H), 7.13 (d, *J* = 8.8 Hz, 1H), 7.16–7.38 (m, 7H), 7.75 (d, *J* = 8.3 Hz, 2H). (m)  $\delta$  1.04 (dd, *J* = 2.1, 7.0 Hz, 6H), 2.18–2.34 (m, 1H), 2.41 (s, 3H), 3.76 (s, 3H), 4.53 (dd, *J* = 4.9, 8.1 Hz, 1H), 4.85 (d, *J* = 12.4 Hz, 1H), 4.93 (d, *J* = 12.2 Hz, 1H), 5.63 (d, *J* = 9.0 Hz, 1H), 6.17 (d, *J* = 9.0 Hz, 1H), 7.16–7.38 (m, 7H), 7.40 (d, *J* = 8.1 Hz, 1H), 7.75 (d, *J* = 8.3 Hz, 2H). Found: C, 57.75; H, 5.90; N, 5.85%. Calcd for C<sub>23</sub>H<sub>28</sub>N<sub>2</sub>O<sub>7</sub>S: C, 57.97; H, 5.92; N, 5.88%.



**Methyl *N*-Boc- $\alpha$ -Tosylglycyl-L-leucinate (2e):** Mp 84.0–86.0 °C (diethyl ether–hexane); IR (KBr) 3309, 2957, 2871, 1729, 1703, 1681, 1655, 1597, 1556, 1521, 1453, 1393, 1367, 1287, 1256, 1147, 1084, 1056, 1019, 977, 867, 817, 732, 708, 670 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  0.99 (d, *J* = 6.3 Hz, 6H), 1.21 (s, 9H), 1.64–1.78 (m, 2H), 1.82–1.92 (m, 1H), 2.42 (s, 3H), 3.76 (s, 3H), 4.71 (dt, *J* = 7.7, 7.7 Hz, 1H), 5.49 (d, *J* = 8.8 Hz, 1H), 5.92 (d, *J* = 8.5 Hz, 1H), 6.97 (d, *J* = 8.1 Hz, 1H), 7.34 (d, *J* = 7.3 Hz, 2H), 7.80 (d, *J* = 7.8 Hz, 2H). (m)  $\delta$  0.95 (d, *J* = 5.9 Hz, 3H), 0.96 (d, *J* = 5.9 Hz, 3H), 1.26 (s, 9H), 1.64–1.78 (m, 2H), 1.82–1.92 (m, 1H), 2.42 (s, 3H), 3.80 (s, 3H), 4.62–4.68 (m, 1H), 5.49 (d, *J* = 8.8 Hz, 1H), 5.87 (d, *J* = 9.0 Hz, 1H), 7.24 (d, *J* = 7.8 Hz, 1H), 7.34 (d, *J* = 7.8 Hz, 2H), 7.82 (d, *J* = 7.8 Hz, 2H). Found: C, 55.02; H, 7.26; N, 6.19%. Calcd for C<sub>21</sub>H<sub>32</sub>N<sub>2</sub>O<sub>7</sub>S: C, 55.25; H, 7.06; N, 6.14%.

**Methyl *N*-Boc- $\alpha$ -Tosylglycyl-L-alaninate (2f):** Mp 144.5 °C (ethyl acetate); IR (KBr) 3420, 3331, 3278, 2979, 2953, 1742, 1684, 1650, 1597, 1518, 1455, 1366, 1345, 1322, 1308, 1291, 1254, 1224, 1152, 1085, 1050, 1021, 975, 866, 831, 768, 715, 667 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  1.23 (s, 9H), 1.53 (d, *J* = 7.1 Hz, 3H), 2.43 (s, 3H), 3.78 (s, 3H), 4.67 (dq, *J* = 7.1, 7.3 Hz, 1H), 5.52 (d, *J* = 8.8 Hz, 1H), 5.94 (d, *J* = 8.8 Hz, 1H), 7.16 (d, *J* = 7.3 Hz, 1H), 7.35 (d, *J* = 7.6 Hz, 2H), 7.80 (d, *J* = 7.6 Hz, 2H). (m)  $\delta$  1.26 (s, 9H), 1.50 (d, *J* = 7.1 Hz, 3H), 2.43 (s, 3H), 3.81 (s, 3H), 4.64 (dq, *J* = 7.1, 7.3 Hz), 5.50 (d, *J* = 9.0 Hz, 1H), 5.92 (d, *J* = 9.0 Hz, 1H), 7.35 (d, *J* = 8.3 Hz, 2H), 7.46 (d, *J* = 6.6 Hz, 1H), 7.82 (d, *J* = 8.3 Hz, 2H). Found: C, 51.91; H, 6.31; N, 6.63%. Calcd for C<sub>18</sub>H<sub>26</sub>N<sub>2</sub>O<sub>7</sub>S: C, 52.16; H, 6.32; N, 6.76%.

**Methyl *N*-Boc- $\alpha$ -Tosylglycylglycinate (2g):** Mp 158.0–159.5 °C (ethyl acetate); IR (KBr) 3356, 3306, 2977, 1743, 1720, 1683, 1599, 1545, 1517, 1442, 1404, 1375, 1338, 1315, 1300, 1256, 1227, 1209, 1162, 1133, 1081, 1040, 974, 934, 869, 841, 813, 767, 706, 692 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.24 (s, 9H), 2.43 (s, 3H), 3.80 (s, 3H), 4.10 (dd, *J* = 5.9, 18.3 Hz, 1H), 4.26 (dd, *J* = 4.9, 18.3 Hz, 1H), 5.56 (d, *J* = 9.0 Hz, 1H), 5.92 (d, *J* = 9.0 Hz, 1H), 7.26–7.30 (br, 1H), 7.35 (d, *J* = 8.3 Hz, 2H), 7.81 (d, *J* = 8.3 Hz, 2H). Found: C, 50.75; H, 6.01; N, 6.95%. Calcd for C<sub>17</sub>H<sub>24</sub>N<sub>2</sub>O<sub>7</sub>S: C, 50.99; H, 6.04; N, 7.00%.

**Methyl *N*-Boc- $\alpha$ -Tosylglycyl-L-valinate (2h):** Mp 135.0–136.0 °C (ethyl acetate–diethyl ether–hexane); IR (KBr) 3341, 3069, 2965, 2882, 1737, 1720, 1679, 1597, 1551, 1513, 1440, 1366, 1334, 1320, 1307, 1255, 1223, 1166, 1140, 1081, 1058, 1021, 983, 908, 864, 845, 819, 774, 729, 708 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  1.03 (d, *J* = 7.0 Hz, 3H), 1.08 (d, *J* = 7.0 Hz, 3H), 1.20 (s, 9H), 2.20–2.36 (m, 1H), 3.78 (s, 3H), 4.67 (dd, *J* = 4.8, 8.8 Hz, 1H), 5.50 (d, *J* = 8.8 Hz, 1H), 5.93 (d, *J* = 8.8 Hz, 1H), 7.05 (d, *J* = 8.8 Hz, 1H), 7.34 (d, *J* = 8.6 Hz, 2H), 7.81 (d, *J* = 8.6 Hz, 2H). (m)  $\delta$  0.98 (d, *J* = 7.0 Hz, 6H), 1.26 (s, 9H), 2.20–2.36 (m, 1H), 3.81 (s, 3H), 4.56 (dd, *J* = 5.0, 8.1 Hz, 1H), 5.50 (d, *J* = 8.8 Hz, 1H), 5.87 (d, *J* = 9.0 Hz, 1H), 7.29–7.31 (br, 1H), 7.34 (d, *J* = 8.6 Hz, 2H), 7.81 (d, *J* = 8.6 Hz, 2H). Found: C, 54.07; H, 6.82; N, 6.31%. Calcd for C<sub>20</sub>H<sub>30</sub>N<sub>2</sub>O<sub>7</sub>S: C, 54.28; H, 6.83; N, 6.33%.

**Methyl *N*-Boc- $\alpha$ -Tosylglycyl-L-isoleucinate (2i):** A solid; IR (KBr) 3343, 2971, 2879, 1739, 1680, 1597, 1495, 1458, 1439, 1368, 1327, 1305, 1253, 1211, 1164, 1146, 1083, 1057, 1020, 980, 900, 864, 814, 768, 733, 707, 660 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  0.96 (t, *J* = 7.6 Hz, 3H), 1.05 (d, *J* = 6.8 Hz, 3H), 1.20 (s, 9H), 1.30–1.37 (m, 2H), 1.46–1.56 (m, 1H), 2.42 (s, 3H), 3.77 (s, 3H), 4.71 (dd, *J* = 4.6, 8.8 Hz, 1H), 5.52 (d, *J* = 8.8 Hz, 1H), 5.94 (d, *J* = 8.8 Hz, 1H), 7.11 (d, *J* = 8.8

Hz, 1H), 7.34 (d, *J* = 8.3 Hz, 2H), 7.81 (d, *J* = 8.3 Hz, 2H). (m)  $\delta$  0.95 (t, *J* = 7.3 Hz, 3H), 1.05 (d, *J* = 6.8 Hz, 3H), 1.20 (s, 9H), 1.30–1.37 (m, 2H), 1.46–1.56 (m, 1H), 2.42 (s, 3H), 3.80 (s, 3H), 4.61 (dd, *J* = 4.6, 7.8 Hz, 1H), 5.51 (d, *J* = 8.8 Hz, 1H), 5.91 (d, *J* = 9.3 Hz, 1H), 7.35 (d, *J* = 8.3 Hz, 1H), 7.37–7.40 (br, 1H), 7.81 (d, *J* = 8.3 Hz, 2H). EI-MS *m/z* 456 (M<sup>+</sup>, 0.12%).

#### A General Procedure for the Preparation of Dipeptides Containing a $\beta$ -Nitro- $\alpha$ -amino Acid Residue 3–16. Method A:

To a solution of KO<sup>t</sup>Bu (270 mg, 2.4 mmol) in dry THF (1 mL) was added a solution of 2-nitropropane (428 mg, 4.8 mmol) in dry THF (2 mL) at –40 °C under nitrogen. After 5 min, a solution of **2a** (392 mg, 0.80 mmol) in dry THF (2 mL) was added over 60 min. The reaction mixture was gradually warmed to room temperature, and then quenched with a saturated aqueous solution of NH<sub>4</sub>Cl. The THF was removed in vacuo to give a residue, which was partitioned between ethyl acetate and water. The aqueous layer was extracted with ethyl acetate. The combined extracts were washed with brine and dried over anhydrous MgSO<sub>4</sub>. Removal of the solvent gave methyl *N*-Z- $\beta$ -nitrovalyl-L-leucinate **3** as an inseparable mixture of diastereomers in quantitative yield (335 mg). Mp 92.0–93.0 °C (ethyl acetate–hexane); IR (KBr) 3291, 3068, 2960, 1713, 1692, 1652, 1551, 1455, 1390, 1360, 1288, 1248, 1153, 1052, 748, 697 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  0.89 (d, *J* = 6.1 Hz, 3H), 0.91 (d, *J* = 6.1 Hz, 3H), 1.50–1.67 (m, 3H), 1.57 (s, 3H), 1.71 (s, 3H), 3.70 (s, 3H), 4.49–4.56 (m, 1H), 4.96 (d, *J* = 8.3 Hz, 1H), 5.11–5.17 (m, 2H), 5.97–6.01 (m, 1H), 6.66 (d, *J* = 7.8 Hz, 1H), 7.32–7.36 (m, 5H). (m)  $\delta$  0.89–0.92 (m, 6H), 1.50–1.67 (m, 3H), 1.60 (s, 3H), 1.71 (s, 3H), 3.72 (s, 3H), 4.49–4.56 (m, 1H), 4.98 (d, *J* = 8.8 Hz, 1H), 5.11–5.17 (m, 2H), 5.97–6.01 (m, 1H), 6.68–6.74 (br, 1H), 7.32–7.36 (m, 5H). Found: C, 56.57; H, 6.90; N, 9.92%. Calcd for C<sub>20</sub>H<sub>29</sub>N<sub>3</sub>O<sub>7</sub>: C, 56.73; H, 6.90; N, 9.92%.

**Method B:** To a suspension of KO<sup>t</sup>Bu (185 mg, 1.65 mmol) in dry THF (6 mL) was added a solution of 2-nitropropane (1.453 g, 1.63 mmol) in THF (2 mL) at –40 °C under nitrogen. After 15 min, a solution of **2a** (402 mg, 0.82 mmol) in THF (4 mL) was added dropwise over 30 min at –40 °C. The mixture was stirred for 1 h and then quenched with a saturated aqueous solution of NH<sub>4</sub>Cl. The THF was removed in vacuo to give a residue, which was partitioned between ethyl acetate and water. The aqueous layer was extracted with ethyl acetate. The combined extracts were washed with brine and dried over anhydrous MgSO<sub>4</sub>. Removal of the solvent and recrystallization from AcOEt–hexane gave **3** as an inseparable mixture of diastereomers in 61% yield (211 mg).

**Methyl 2-(Benzyloxycarbonylamino)-3-nitrobutanoyl-L-leucinate (4):** Mp 85.0–86.0 °C (ethyl acetate–hexane); IR (KBr) 3287, 3076, 2959, 1748, 1713, 1693, 1658, 1555, 1455, 1389, 1361, 1285, 1244, 1153, 1056, 1030, 873, 739, 698 cm<sup>-1</sup>. Compound **4** was found to be a mixture of four diastereomers based on the NMR spectrum. Found: C, 55.94; H, 6.62; N, 10.16%. Calcd for C<sub>19</sub>H<sub>27</sub>N<sub>3</sub>O<sub>7</sub>: C, 55.74; H, 6.65; N, 10.26%.

**Methyl *N*-Z- $\beta$ -Nitrovalyl-L-alaninate (5):** Mp 110.0–112.0 °C (ethyl acetate–hexane); IR (KBr) 3328, 3269, 3070, 3002, 2953, 1754, 1736, 1715, 1665, 1540, 1457, 1400, 1383, 1345, 1324, 1289, 1262, 1213, 1152, 1061, 1016, 912, 859, 797, 757, 741, 728, 698, 677 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  1.37 (d, *J* = 7.1 Hz, 3H), 1.61 (s, 9H), 1.73 (s, 3H), 3.73 (s, 3H), 4.42–4.58 (m, 1H), 4.93 (d, *J* = 9.8 Hz, 1H), 5.10–5.19 (m, 2H), 5.97 (d, *J* = 7.8 Hz, 1H), 6.76–6.90 (br, 1H), 7.30–7.40 (m, 5H). (m)  $\delta$  1.37 (d, *J* = 7.1 Hz, 3H), 1.57 (s, 3H), 1.72 (s, 3H), 4.42–4.58 (m, 1H), 4.97 (d, *J* = 9.5 Hz, 1H), 5.10–5.19

(m, 2H), 5.99 (d,  $J = 8.8$  Hz, 1H), 6.76–6.90 (br, 1H), 7.30–7.40 (m, 5H). Found: C, 53.43; H, 6.13; N, 11.09%. Calcd for  $C_{17}H_{23}N_3O_7$ : C, 53.54; H, 6.08; N, 11.02%.

**Methyl *N*-*Z*- $\beta$ -Nitrovalylglycinate (6):** Mp 140.5–141.5 °C (ethyl acetate); IR (KBr) 3303, 3067, 2954, 2851, 1745, 1717, 1665, 1540, 1499, 1455, 1439, 1393, 1376, 1345, 1323, 1259, 1223, 1137, 1105, 1057, 1012, 981, 902, 856, 794, 778, 742, 697  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ )  $\delta$  1.63 (s, 3H), 1.73 (s, 3H), 3.73 (s, 3H), 3.89 (dd,  $J = 5.6, 18.3$  Hz, 1H), 4.07 (dd,  $J = 5.6, 18.3$  Hz, 1H), 5.01 (d,  $J = 9.8$  Hz, 1H), 5.10 (d,  $J = 12.2$  Hz, 1H), 5.15 (d,  $J = 12.0$  Hz, 1H), 6.03 (d,  $J = 9.8$  Hz, 1H), 6.94 (br, 1H), 7.31–7.38 (m, 5H). Found: C, 52.54; H, 5.83; N, 11.51%. Calcd for  $C_{16}H_{21}N_3O_7$ : C, 52.31; H, 5.76; N, 11.44%.

**Methyl *N*-*Z*- $\beta$ -Nitrovalyl-L-alaninate (7):** Mp 110.0–112.0 °C (ethyl acetate–hexane); IR (KBr) 3328, 3269, 3070, 2953, 1754, 1736, 1715, 1665, 1540, 1457, 1400, 1383, 1345, 1324, 1289, 1262, 1213, 1152, 1061, 1016, 912, 859, 797, 757, 741, 728, 698, 677  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ ) (M)  $\delta$  1.37 (d,  $J = 7.1$  Hz, 3H), 1.61 (s, 9H), 1.73 (s, 3H), 3.73 (s, 3H), 4.42–4.58 (m, 1H), 4.93 (d,  $J = 9.8$  Hz, 1H), 5.10–5.19 (m, 2H), 5.97 (d,  $J = 8.8$  Hz, 1H), 7.30–7.40 (m, 5H). (m)  $\delta$  1.37 (d,  $J = 7.1$  Hz, 3H), 1.57 (s, 3H), 1.72 (s, 3H), 3.73 (s, 3H), 4.42–4.58 (m, 1H), 4.97 (d,  $J = 9.5$  Hz, 1H), 5.10–5.19 (m, 2H), 5.99 (d,  $J = 8.8$  Hz, 1H), 6.76–6.90 (br, 1H), 7.30–7.40 (m, 5H). Found: C, 53.43; H, 6.13; N, 11.09%. Calcd for  $C_{17}H_{23}N_3O_7$ : C, 53.54; H, 6.08; N, 11.02%.

**Methyl 2-(Benzyloxycarbonylamino)-3-nitrobutanoyl-L-alaninate (8):** Mp 131.5–134.0 °C (ethyl acetate–hexane); IR (KBr) 3287, 3068, 3035, 2966, 2934, 2878, 2851, 2786, 1957, 1744, 1715, 1664, 1539, 1456, 1437, 1376, 1346, 1258, 1146, 1115, 1102, 1087, 1059, 1014, 911, 857, 787, 754, 739, 699, 686  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ ) (M)  $\delta$  (d,  $J = 6.8$  Hz, 3H), 0.89 (d,  $J = 6.8$  Hz, 3H), 1.60 (s, 3H), 1.72 (s, 3H), 2.10–2.20 (m, 1H), 3.72 (s, 3H), 4.43 (dd,  $J = 4.9, 8.3$  Hz, 1H), 4.96 (d,  $J = 9.8$  Hz, 1H), 5.15 (s, 2H), 6.04 (d,  $J = 9.8$  Hz, 1H), 6.73–6.75 (br, 1H), 7.32–7.38 (m, 5H). (m)  $\delta$  0.86 (d,  $J = 6.8$  Hz, 3H), 0.89 (d,  $J = 6.8$  Hz, 3H), 1.58 (s, 3H), 1.72 (s, 3H), 2.10–2.20 (m, 1H), 3.72 (s, 3H), 4.46 (dd,  $J = 4.7, 8.4$  Hz, 1H), 5.00 (d,  $J = 8.8$  Hz, 1H), 5.15 (s, 2H), 5.98 (d,  $J = 8.8$  Hz, 1H), 6.64–6.66 (br, 1H), 7.32–7.38 (m, 5H). Found: C, 55.59; H, 6.74; N, 10.28%. Calcd for  $C_{19}H_{27}N_3O_7$ : C, 55.74; H, 6.65; N, 10.26%.

**Methyl *N*-Boc- $\beta$ -Nitrovalyl-L-leucinate (9):** Mp 141.0–144.0 °C (ethyl acetate–hexane); IR (KBr) 3280, 3105, 2957, 2874, 1743, 1692, 1655, 1546, 1457, 1435, 1394, 1369, 1345, 1332, 1272, 1163, 1099, 1060, 1020, 984, 879, 854, 803, 754, 715  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ ) (M)  $\delta$  0.93 (d,  $J = 5.3$  Hz, 6H), 1.43–1.68 (m, 3H), 1.47 (s, 9H), 1.59 (s, 3H), 1.74 (s, 3H), 3.72 (s, 3H), 4.40–4.60 (m, 1H), 4.81 (d,  $J = 9.7$  Hz, 1H), 5.66–5.76 (br, 1H), 6.55 (d,  $J = 7.9$  Hz, 1H). (m)  $\delta$  0.91 (d,  $J = 5.5$  Hz, 6H), 1.43–1.68 (m, 3H), 1.46 (d,  $J = 5.5$  Hz, 6H), 1.62 (s, 3H), 1.74 (s, 3H), 3.72 (s, 3H), 4.40–4.60 (m, 1H), 4.80 (d,  $J = 7.9$  Hz, 1H), 5.66–5.76 (br, 1H), 6.55 (d,  $J = 7.9$  Hz, 1H). Found: C, 52.39; H, 8.04; N, 10.58%. Calcd for  $C_{17}H_{31}N_3O_7$ : C, 52.43; H, 8.02; N, 10.79%.

**Methyl 2-(*t*-Butoxycarbonylamino)-3-nitrobutanoyl-L-leucinate (10):** Mp 161.0–164.0 °C (ethyl acetate–hexane); IR (KBr) 3342, 3278, 3088, 2976, 1755, 1687, 1655, 1554, 1520, 1455, 1392, 1369, 1305, 1253, 1204, 1160, 1112, 1055, 874, 755  $cm^{-1}$ . Compound **10** was found to be a mixture of four diastereomers based on the NMR spectrum. Found: C 51.12; H, 7.83; N, 11.18%. Calcd for  $C_{16}H_{29}N_3O_7$ : C, 51.19; H, 7.79; N, 11.19%.

**Methyl *N*-Boc-( $\beta$ -nitro)phenylalanyl-L-leucinate (11):** Mp 169.0–170.0 °C (ethyl acetate–hexane); IR (KBr) 3302, 3089, 2959, 2871, 1726, 1693, 1657, 1557, 1525, 1458, 1438, 1369, 1329, 1289, 1249, 1208, 1168, 1050, 1024, 863, 801, 750, 719  $cm^{-1}$ . Compound **11** was found to be a mixture of four diastereomers based on the NMR spectrum. Found: C 57.53; H, 7.17; N, 9.60%. Calcd for  $C_{21}H_{31}N_3O_7$ : C, 57.65; H, 7.14; N, 9.60%.

**Methyl *N*-Boc- $\beta$ -Nitroisoleucyl-L-leucinate (12):** Mp 143.5–145.0 °C (ethyl acetate); IR (KBr) 3326, 2979, 2883, 1750, 1688, 1655, 1548, 1461, 1380, 1369, 1350, 1323, 1275, 1254, 1212, 1156, 1061, 1012, 985, 881, 857, 769, 669  $cm^{-1}$ . Compound **12** was found to be a mixture of four diastereomers based on the NMR spectrum. Found: C, 53.57; H, 8.23; N, 10.41%. Calcd for  $C_{18}H_{33}N_3O_7$ : C, 53.58; H, 8.24; N, 10.42%.

**Methyl *N*-Boc- $\beta$ -Nitrovalyl-L-alaninate (13):** Mp 152.5–154.0 °C (ethyl acetate); IR (KBr) 3279, 3089, 2982, 2955, 1753, 1696, 1665, 1546, 1456, 1381, 1369, 1347, 1328, 1281, 1254, 1218, 1162, 1052, 1013, 877, 856, 835, 806, 751, 669  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ ) (M)  $\delta$  1.39 (d,  $J = 7.1$  Hz, 3H), 1.47 (s, 9H), 1.62 (s, 3H), 1.74 (s, 3H), 3.74 (s, 3H), 4.46–4.58 (m, 1H), 4.80 (d,  $J = 9.8$  Hz, 1H), 5.69 (d,  $J = 9.3$  Hz, 1H), 6.71 (d,  $J = 5.9$  Hz, 1H). (m)  $\delta$  1.40 (d,  $J = 7.3$  Hz, 3H), 1.47 (s, 9H), 1.59 (s, 3H), 1.75 (s, 3H), 3.75 (s, 3H), 4.46–4.58 (m, 1H), 4.84 (d,  $J = 10.0$  Hz, 1H), 5.69 (d,  $J = 9.3$  Hz, 1H), 6.70–6.82 (m, 1H). Found: C, 48.26; H, 7.31; N, 12.00%. Calcd for  $C_{14}H_{25}N_3O_7$ : C, 48.41; H, 7.25; N, 12.10%.

**Methyl *N*-Boc- $\beta$ -Nitrovalylglycinate (14):** Mp 173.0–180.0 °C (ethyl acetate); IR (KBr) 3281, 3096, 3005, 2984, 1762, 1697, 1668, 1549, 1456, 1438, 1405, 1394, 1381, 1370, 1350, 1322, 1282, 1258, 1209, 1179, 1100, 1059, 1038, 1015, 982, 931, 877, 855, 791, 706, 662  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ )  $\delta$  1.47 (s, 9H), 1.62 (s, 3H), 1.77 (s, 3H), 3.75 (s, 3H), 3.97 (dd,  $J = 5.6, 18.3$  Hz, 1H), 4.05 (dd,  $J = 5.6, 18.3$  Hz, 1H), 4.83 (d,  $J = 9.8$  Hz, 1H), 5.68 (d,  $J = 9.8$  Hz, 1H), 6.62 (br, 1H). Found: C, 46.75; H, 6.98; N, 12.56%. Calcd for  $C_{13}H_{23}N_3O_7$ : C, 46.84; H, 6.95; N, 12.61%.

**Methyl *N*-Boc- $\beta$ -Nitrovalyl-L-valinate (15):** Mp 143.0 °C (benzene–hexane); IR (KBr) 3326, 2971, 2935, 2879, 1749, 1691, 1653, 1549, 1457, 1392, 1379, 1369, 1351, 1327, 1278, 1255, 1216, 1157, 1062, 1012, 981, 953, 881, 858, 792, 769, 752, 662  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ ) (M)  $\delta$  0.89 (d,  $J = 7.1$  Hz, 3H), 0.93 (d,  $J = 7.1$  Hz, 3H), 1.47 (s, 9H), 1.61 (s, 3H), 1.75 (s, 3H), 2.13–2.23 (m, 1H), 4.44 (dd,  $J = 4.6, 8.8$  Hz, 1H), 4.77 (d,  $J = 9.5$  Hz, 1H), 5.78 (d,  $J = 9.5$  Hz, 1H), 6.56–6.58 (br, 1H). (m)  $\delta$  0.87 (d,  $J = 7.1$  Hz, 3H), 0.89 (d,  $J = 7.1$  Hz, 3H), 1.47 (s, 9H), 1.59 (s, 3H), 1.74 (s, 3H), 2.13–2.20 (m, 1H), 3.74 (s, 3H), 4.46 (dd,  $J = 4.6, 10.0$  Hz, 1H), 4.85 (d,  $J = 9.5$  Hz, 1H), 5.69 (d,  $J = 9.5$  Hz, 1H), 6.53–6.56 (br, 1H). Found: C, 51.00; H, 7.81; N, 11.07%. Calcd for  $C_{16}H_{29}N_3O_7$ : C, 51.19; H, 7.79; N, 11.19%.

**Methyl *N*-Boc- $\beta$ -Nitrovalyl-L-isoleucinate (16):** Mp 123.0–123.5 °C (benzene–hexane); IR (KBr) 3326, 2979, 2883, 1750, 1688, 1655, 1548, 1461, 1380, 1369, 1350, 1323, 1275, 1254, 1212, 1156, 1061, 1012, 881, 857, 769  $cm^{-1}$ ;  $^1H$ NMR (400 MHz,  $CDCl_3$ ) (M)  $\delta$  0.89 (t,  $J = 9.3$  Hz, 3H), 0.90 (d,  $J = 11.0$  Hz, 3H), 1.11–1.21 (m, 1H), 1.35–1.42 (m, 1H), 1.47 (s, 9H), 1.61 (s, 3H), 1.74 (s, 3H), 1.82–1.96 (m, 1H), 3.73 (s, 3H), 4.48 (m, 1H), 4.80 (d,  $J = 9.8$  Hz, 1H), 5.77 (d,  $J = 9.3$  Hz, 1H), 6.63–6.66 (m, 1H). (m)  $\delta$  0.90 (d,  $J = 11.0$  Hz, 3H), 0.90 (t,  $J = 9.3$  Hz, 3H), 1.11–1.21 (m, 1H), 1.35–1.42 (m, 1H), 1.47 (s, 9H), 1.59 (s, 3H), 1.74 (s, 3H), 1.82–1.96 (m, 1H), 3.73 (s, 3H), 4.41 (dd,  $J = 4.6, 8.5$  Hz, 1H), 4.85 (d,  $J = 9.3$  Hz, 1H), 5.70 (d,  $J =$

9.3 Hz, 1H), 6.60–6.63 (m, 1H). Found: C, 52.18; H, 8.01; N, 10.59%. Calcd for C<sub>17</sub>H<sub>31</sub>N<sub>3</sub>O<sub>7</sub>: C, 52.43; H, 8.02; N, 10.79%.

**A General Procedure for the Preparation of Dipeptide Containing an  $\alpha,\beta$ -Didehydroamino Acid Residue (17–28).** To a solution of compound **16** (164 mg, 0.42 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (4 mL) was added a solution of DBU (128 mg, 0.42 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2 mL) at 0 °C under nitrogen. After stirring for 24 h at room temperature, the solvent was removed in vacuo to afford a residue, which was partitioned between ethyl acetate and water. The aqueous layer was extracted with ethyl acetate, and the combined extracts were washed with 10% citric acid, 10% NaHCO<sub>3</sub>, and brine, dried over anhydrous MgSO<sub>4</sub>, and evaporated under reduced pressure to afford a residue, which was subjected to preparative TLC (SiO<sub>2</sub>, hexane:ethyl acetate = 2/1, v/v) to afford methyl *N*-Boc- $\alpha,\beta$ -didehydrovalyl-L-isoleucinate (**28**) in 91% yield (131 mg). Mp 78.0–79.0 °C (hexane);  $[\alpha]_D^{25} = -7.1^\circ$  (0.27, MeOH); IR (KBr) 3319, 3060, 2973, 1745, 1691, 1670, 1629, 1549, 1458, 1368, 1348, 1254, 1201, 1168, 1055, 856 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.92 (t, *J* = 7.3 Hz, 3H), 0.93 (d, *J* = 6.8 Hz, 3H), 1.14–1.30 (m, 1H), 1.40–1.50 (m, 1H), 1.46 (s, 9H), 1.79 (s, 3H), 1.87–1.97 (m, 1H), 2.04 (s, 3H), 3.73 (s, 3H), 4.61 (dd, *J* = 4.9, 8.5 Hz, 1H), 5.95 (s, 1H), 6.85 (br, 1H). Found: C, 59.39; H, 8.87; N, 7.93%. Calcd for C<sub>17</sub>H<sub>30</sub>N<sub>2</sub>O<sub>5</sub>: C, 59.63; H, 8.83; N, 8.18%.

The physical and spectral data of compounds **17–27** are shown in the following.

**Methyl *N*-Z- $\alpha,\beta$ -Didehydrovalyl-L-leucinate (17):** Mp 128.0–129.0 °C (ethyl acetate–hexane);  $[\alpha]_D^{25} = -24.4^\circ$  (0.59, MeOH); IR (KBr) 3284, 3048, 2956, 2869, 1756, 1703, 1626, 1559, 1509, 1250, 1229, 1202, 1157, 1060, 738 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.87–0.96 (br, 6H), 1.51–1.72 (m, 3H), 1.78 (s, 3H), 2.06 (s, 3H), 4.60–4.67 (br, 1H), 5.15 (s, 2H), 5.95 (s, 1H), 6.45–6.55 (br, 1H), 7.35–7.52 (m, 5H). Found: C, 63.59; H, 7.53; N, 7.36%. Calcd for C<sub>20</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>: C, 63.81; H, 7.50; N, 7.44%.

**Methyl (Z)-2-Benzyloxycarbonylamino-2-butenoyl-L-leucinate (18):** Mp 77.5–78.0 °C (ethyl acetate);  $[\alpha]_D^{25} = -27.8^\circ$  (0.50, MeOH); IR (KBr) 3244, 3042, 2956, 2870, 1752, 1698, 1671, 1630, 1551, 1507, 1252, 1232, 1202, 1158, 1099, 740, 696 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.90–0.92 (m, 2H), 1.51–1.66 (m, 2H), 1.74 (d, *J* = 7.0 Hz, 3H), 3.71 (s, 3H), 4.62–4.70 (m, 1H), 5.14 (s, 2H), 6.53 (q, *J* = 7.0 Hz, 1H), 6.57 (d, *J* = 8.3 Hz, 1H), 7.31–7.38 (br, 5H). Found: C, 61.25; H, 7.21; N, 7.32%. Calcd for C<sub>19</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub>·1/2H<sub>2</sub>O: C, 61.42; H, 7.33; N, 7.54%.

**Methyl *N*-Z- $\alpha,\beta$ -Didehydrovalyl-L-alaninate (19):** Mp 83.0–84.0 °C (benzene–hexane);  $[\alpha]_D^{25} = -7.8^\circ$  (0.83, MeOH); IR (KBr) 3276, 2987, 2951, 1745, 1687, 1631, 1543, 1507, 1257, 1219, 1161, 1042, 860, 755, 732, 698 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.32–1.50 (br, 3H), 1.77 (s, 3H), 2.05 (s, 3H), 3.72 (s, 3H), 4.50–4.70 (br, 1H), 5.14 (s, 2H), 6.22 (s, 1H), 6.60–6.80 (br, 1H), 7.28–7.40 (br, 5H). Found: C, 61.02; H, 6.64; N, 8.22%. Calcd for C<sub>17</sub>H<sub>22</sub>N<sub>2</sub>O<sub>5</sub>: C, 61.07; H, 6.63; N, 8.38%.

**Methyl *N*-Z- $\alpha,\beta$ -Didehydrovalylglycinate (20):** Mp 107.0–108.0 °C (ethyl acetate–hexane); IR (KBr) 3298, 2947, 1764, 1707, 1663, 1499, 1446, 1388, 1356, 1329, 1208, 1186, 1118, 1011, 961, 928, 915, 787, 765, 752, 718, 698 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.78 (s, 3H), 2.07 (s, 3H), 3.74 (s, 3H), 4.03–4.17 (br, 2H), 5.13 (s, 2H), 6.17 (s, 1H), 6.50–6.78 (br, 1H), 7.35–7.40 (m, 5H). Found: C, 59.93; H, 6.31; N, 8.60%. Calcd for C<sub>16</sub>H<sub>20</sub>N<sub>2</sub>O<sub>5</sub>: C, 59.99; H, 6.29; N, 8.74%.

**Methyl *N*-Boc- $\alpha,\beta$ -Didehydrovalyl-L-leucinate (21):** Mp 133.0 °C (ethyl acetate–hexane);  $[\alpha]_D^{25} = -37.5^\circ$  (0.36, MeOH); IR (KBr) 3279, 2960, 2868, 1757, 1699, 1624, 1541, 1492, 1458, 1370, 1333, 1251, 1232, 1201, 1162, 1053, 1031, 986, 897, 698 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.90–0.92 (m, 6H), 1.46 (s, 9H), 1.54–1.73 (m, 3H), 1.79 (s, 3H), 2.05 (s, 3H), 3.73 (s, 3H), 4.63 (dt, *J* = 4.8, 8.8 Hz, 1H), 5.68 (br, 1H), 6.54–6.65 (br, 1H). Found: C, 59.42; H, 8.91; N, 8.04%. Calcd for C<sub>17</sub>H<sub>30</sub>N<sub>2</sub>O<sub>5</sub>: C, 59.63; H, 8.83; N, 8.18%.

**Methyl (Z)-2-(*t*-Butoxycarbonylamino)-2-butenoyl-L-leucinate (22):** Mp 110.0–112.0 °C (ethyl acetate)  $[\alpha]_D^{25} = -33.7^\circ$  (1.00, MeOH) IR (KBr) 3334, 3220, 3049, 2964, 2868, 1759, 1688, 1670, 1626, 1548, 1488, 1453, 1371, 1344, 1304, 1252, 1233, 1201, 1159, 1096, 1053, 1031, 989, 905, 859, 785, 711, 666 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.94 (d, *J* = 6.3 Hz, 3H), 0.95 (d, *J* = 6.1 Hz, 3H), 1.46 (s, 9H), 1.57–1.71 (m, 3H), 1.76 (d, *J* = 7.1 Hz, 3H), 3.73 (s, 3H), 4.67 (m, 1H), 6.25 (s, 1H), 6.53 (q, *J* = 7.1 Hz, 1H), 6.69 (d, *J* = 8.3 Hz, 1H). HRMS (FAB) (*M*<sup>+</sup> + 1): Found: *m/z* 329.2068. Calcd for C<sub>16</sub>H<sub>29</sub>N<sub>2</sub>O<sub>5</sub>: 329.2076. When the carbamate NH proton was irradiated, 3.3%, 2.5%, and 2.2% of the NOE were observed for the <sup>t</sup>Bu protons of the Boc group, the methyl protons at the double bond, and the amide NH proton, respectively.

**Methyl (Z)-*N*-Boc- $\alpha,\beta$ -Didehydrophenylalanyl-L-leucinate (23):** Mp 104.0–105.0 °C (ethyl acetate–hexane);  $[\alpha]_D^{25} = -43.0^\circ$  (1.00, MeOH) [Lit. mp 99–101.0 °C,  $[\alpha]_D^{23} = -41.8^\circ$  (1.00, MeOH)];<sup>41</sup> IR (KBr) 3230, 3056, 2957, 2871, 1743, 1680, 1658, 1629, 1550, 1504, 1489, 1446, 1391, 1366, 1342, 1304, 1250, 1201, 1164, 1078, 1061, 1029, 962, 938, 882, 841, 783, 754, 689 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.59 (d, *J* = 5.7 Hz, 3H), 0.97 (d, *J* = 5.7 Hz, 3H), 1.40 (s, 9H), 1.58–1.79 (m, 3H), 3.73 (s, 3H), 4.70–4.78 (m, 1H), 6.29 (s, 1H), 6.90 (d, *J* = 8.1 Hz, 1H), 7.21 (s, 1H), 7.27–7.46 (m, 5H). Found: C, 64.38; H, 7.79; N, 7.04%. Calcd for C<sub>21</sub>H<sub>30</sub>N<sub>2</sub>O<sub>5</sub>: C, 64.60; H, 7.74; N, 7.17%. When the carbamate NH proton was irradiated, 2.4% and 8.9% of the NOE were observed for the amide NH proton and the ortho protons of the phenyl group, respectively.

**Methyl (*E,Z*)-*N*-Boc- $\alpha,\beta$ -Didehydroisoleucyl-L-leucinate (24):** A mixture of (*Z*)- and (*E*)-isomers. Mp 108.0–108.5 °C (ethyl acetate); IR (KBr) 3349, 3058, 2938, 2740, 2344, 1758, 1690, 1637, 1491, 1386, 1332, 1252, 1201, 1093, 1030 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>) (*Z*)-isomer  $\delta$  0.93 (d, *J* = 5.9 Hz, 3H), 0.94 (d, *J* = 5.9 Hz, 3H), 1.03 (t, *J* = 7.6 Hz, 3H), 1.45 (s, 9H), 1.55–1.73 (m, 3H), 2.01 (s, 3H), 2.15 (q, *J* = 7.6 Hz, 2H), 3.72 (s, 3H), 4.62–4.68 (m, 1H), 5.89 (s, 1H), 7.30 (br, 1H). (*E*)-isomer  $\delta$  0.93 (d, *J* = 5.9 Hz, 3H), 0.94 (d, *J* = 5.9 Hz, 3H), 1.08 (t, *J* = 7.6 Hz, 3H), 1.45 (s, 9H), 1.55–1.73 (m, 3H), 1.76 (s, 3H), 2.15 (q, *J* = 7.6 Hz, 2H), 3.72 (s, 3H), 4.62–4.68 (m, 1H), 7.30 (br, 1H). Found: C, 60.45; H, 9.06; N, 7.78%. Calcd for C<sub>18</sub>H<sub>32</sub>N<sub>2</sub>O<sub>5</sub>: C, 60.65; H, 9.05; N, 7.86%.

**Methyl *N*-Boc- $\alpha,\beta$ -Didehydrovalyl-L-alaninate (25):** Mp 118.5–119.0 °C (ethyl acetate–hexane);  $[\alpha]_D^{25} = -14.4^\circ$  (0.73, MeOH); IR (KBr) 3282, 3060, 2982, 1753, 1688, 1665, 1635, 1542, 1457, 1367, 1339, 1278, 1252, 1210, 1164, 1055, 1029, 689 cm<sup>-1</sup>; <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.43 (d, *J* = 7.1 Hz, 3H), 1.46 (s, 9H), 1.79 (s, 3H), 2.05 (s, 3H), 3.75 (s, 3H), 4.63 (dq, *J* = 7.1, 7.3 Hz, 1H), 5.83 (s, 1H), 6.75 (br, 1H). Found: C, 56.04; H, 8.13; N, 9.13%. Calcd for C<sub>14</sub>H<sub>24</sub>N<sub>2</sub>O<sub>5</sub>: C, 55.99; H, 8.05; N, 9.33%.

**Methyl *N*-Boc- $\alpha,\beta$ -Didehydrovalylglycinate (26):** Mp 164.0–165.0 °C (ethyl acetate–hexane); IR (KBr) 3335, 3189, 2985, 1761, 1691, 1659, 1634, 1540, 1456, 1435, 1397, 1365,



1333, 1288, 1258, 1202, 1175, 1135, 1085, 1058, 1034, 1013, 981, 900, 831, 779, 658 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.46 (s, 9H), 1.80 (s, 3H), 2.07 (s, 3H), 3.76 (s, 3H), 4.11 (d, *J* = 5.7 Hz, 2H), 5.74 (br, 1H), 6.70 (br, 1H). Found: C, 54.39; H, 7.76; N, 9.79%. Calcd for C<sub>13</sub>H<sub>22</sub>N<sub>2</sub>O<sub>5</sub>: C, 54.53; H, 7.74; N, 9.78%.

**Methyl *N*-Boc- $\alpha,\beta$ -Didehydrovalyl-L-valinate (27):** Mp 120.5–121.0 °C (hexane); [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -9.7° (0.48, MeOH); IR (KBr) 3299, 2976, 2364, 1750, 1686, 1638, 1543, 1510, 1365, 1273, 1252, 1174, 1057, 900 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 0.92 (d, *J* = 6.8 Hz, 3H), 0.97 (d, *J* = 6.8 Hz, 3H), 1.46 (s, 9H), 1.80 (s, 3H), 2.05 (s, 3H), 2.16–2.24 (m, 1H), 3.73 (dd, *J* = 4.8, 8.7 Hz, 1H), 5.95 (s, 1H), 5.76 (br, 1H), 6.77 (br, 1H). Found: C, 58.39; H, 8.65; N, 8.32%. Calcd for C<sub>16</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>: C, 58.52; H, 8.59; N, 8.53%.

**Reaction of *N*-Boc- $\alpha$ -Tosylglycylamino Acid Methyl Ester with Aldehydes.** To a solution of Bu<sub>3</sub>P (124 mg, 0.61 mmol) and DBU (73 mg, 0.48 mmol) in dry THF (3 mL) was added 0.12 mL of propanal (95 mg, 1.6 mmol) at 0 °C under nitrogen. A solution of **2e** (187 mg, 0.41 mmol) in dry THF (1 mL) was added to the solution over 10 min at 0 °C under nitrogen. The mixture was allowed to stir for 5.5 h at room temperature. After removal of the solvent, a residue was partitioned between ethyl acetate and water. The aqueous layer was extracted with ethyl acetate. The combined extracts were washed with brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated under reduced pressure. The residual oil was subjected to preparative TLC (SiO<sub>2</sub>, hexane:ethyl acetate = 4/1, v/v) to afford (*Z*)-isomer **29** (132 mg, 88%) and (*E*)-isomer **29'** (13 mg, 9%) in 97% yields, respectively. (*Z*)-isomer **29**: Mp 136.5–137.5 °C (ethyl acetate); [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -34.4° (1.00, MeOH); IR (KBr) 3350, 3224, 3053, 2965, 2870, 1759, 1690, 1667, 1631, 1549, 1485, 1368, 1324, 1296, 1249, 1230, 1159, 1096, 1053, 1020, 981, 903, 857, 825, 790, 752, 669 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 0.94 (d, *J* = 5.9 Hz, 3H), 0.95 (d, *J* = 5.6 Hz, 3H), 1.06 (t, *J* = 7.6 Hz, 3H), 1.46 (s, 9H), 1.54–1.74 (m, 2H), 2.17 (dq, *J* = 7.3, 7.6 Hz, 2H), 3.73 (s, 3H), 4.69 (dt, *J* = 4.6, 8.5 Hz, 1H), 5.95 (s, 1H), 6.40–6.43 (m, 1H), 6.57 (d, *J* = 8.1 Hz, 1H). Found: C, 59.34; H, 8.86; N, 8.13%. Calcd for C<sub>17</sub>H<sub>30</sub>N<sub>2</sub>O<sub>5</sub>: C, 59.36; H, 8.83; N, 8.18%. When the carbamate NH proton of the major product **29** was irradiated, 2.2% and 4.4% of the NOE were observed for the amide NH proton and methylene protons of the ethyl group, respectively.

(*E*)-isomer **29'**: A solid; [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -45.5° (0.40, MeOH); IR (KBr) 3255, 3058, 3009, 2964, 2935, 2870, 2365, 1760, 1693, 1655, 1636, 1545, 1503, 1457, 1384, 1369, 1329, 1251, 1232, 1197, 1158, 1050, 1022, 985 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 0.95 (d, *J* = 5.6 Hz, 3H), 0.96 (d, *J* = 5.9 Hz, 3H), 1.10 (t, *J* = 7.6 Hz, 3H), 1.46 (s, 9H), 1.57–1.71 (m, 3H), 2.44 (dq, *J* = 7.3, 7.6 Hz, 2H), 3.75 (s, 3H), 4.70 (dt, *J* = 4.9, 8.3 Hz, 1H), 6.16 (br, 1H), 6.32–6.46 (m, 2H). HRMS (FAB) (M<sup>+</sup> + 1): Found: *m/z* 343.2247. Calcd for C<sub>17</sub>H<sub>31</sub>N<sub>2</sub>O<sub>5</sub>: 343.2233.

The physical and spectral data of compounds **22'**, **30**, **30'**, **31**, and **31'** are shown in the following.

**Methyl (*E*)-2-(*t*-Butoxycarbonylamino)-2-butenoyl-L-leucinate (22')**: A solid; [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -15.8° (0.18, MeOH); IR (KBr) 3336, 3225, 3060, 3014, 2965, 2927, 2869, 2367, 2346, 1761, 1693, 1663, 1631, 1555, 1504, 1455, 1386, 1371, 1252, 1199, 1158, 1130, 1048, 1024, 987 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 0.95 (d, *J* = 6.3 Hz, 3H), 0.97 (d, *J* = 6.1 Hz, 3H), 1.45 (s, 9H), 1.55–1.85 (m, 3H), 2.01 (d, *J* = 7.6 Hz, 3H), 3.75 (s, 3H), 4.71 (dt, *J* = 5.1, 8.5 Hz, 2H), 6.28 (br, 1H), 6.38 (q, *J* = 7.6 Hz, 1H). HRMS (FAB) (M<sup>+</sup> + 1): Found: *m/z* 329.2077. Calcd for C<sub>16</sub>H<sub>29</sub>N<sub>2</sub>O<sub>5</sub>: 329.2076.

**Methyl (*Z*)-2-(*t*-Butoxycarbonylamino)-2-butenoylglycinate (30):** Mp 116.0–118.0 °C (ethyl acetate); IR (KBr) 3324, 3207, 2973, 1762, 1691, 1674, 1631, 1536, 1498, 1456, 1436, 1405, 1367, 1298, 1273, 1254, 1208, 1173, 1097, 1050, 1010, 986 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.46 (s, 9H), 1.77 (d, *J* = 7.1 Hz, 3H), 3.75 (s, 3H), 4.10 (d, *J* = 5.4 Hz, 2H), 5.70–5.85 (br, 1H), 6.52 (q, *J* = 7.1 Hz, 1H), 6.50–6.60 (br, 1H). Found: C, 52.86; H, 7.51; N, 10.21%. Calcd for C<sub>12</sub>H<sub>20</sub>N<sub>2</sub>O<sub>5</sub>: C, 52.93; H, 7.40; N, 10.29%. In order to confirm the stereochemistry of the major product **30**, an authentic sample was prepared as follows: (*Z*)-2-(*t*-butoxycarbonylamino)butenoic acid and methyl glycinate were condensed by the DCC–HOBt method (57%). The NMR spectrum of the product was consistent with that of **30**.

**Methyl (*E*)-2-(*t*-Butoxycarbonylamino)-2-butenoylglycinate (30')**: A solid; IR (KBr) 3326, 3195, 2928, 2850, 1756, 1687, 1662, 1640, 1552, 1519, 1433, 1397, 1364, 1284, 1258, 1205, 1165, 1122, 1050, 1033, 1017, 982 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.46 (s, 9H), 2.02 (d, *J* = 7.5 Hz, 3H), 3.79 (s, 3H), 4.16 (d, *J* = 6.3 Hz, 2H), 6.20–6.30 (br, 1H), 6.30–6.40 (br, 1H), 6.45–6.60 (m, 1H). HRMS (FAB) (M<sup>+</sup> + 1): Found: *m/z* 273.1434. Calcd for C<sub>12</sub>H<sub>21</sub>N<sub>2</sub>O<sub>5</sub>: 273.1450.

**Methyl (*Z*)-2-(*t*-Butoxycarbonylamino)-2-pentenoylglycinate (31):** A solid; IR (KBr) 3265, 3060, 3006, 2978, 2937, 2875, 1763, 1748, 1704, 1683, 1666, 1639, 1513, 1460, 1434, 1414, 1392, 1366, 1319, 1279, 1254, 1213, 1173, 1124, 1107, 1059, 1039, 1020, 984 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.07 (t, *J* = 7.6 Hz, 3H), 1.47 (s, 9H), 2.18 (dq, *J* = 7.3, 7.6 Hz, 2H), 3.77 (s, 3H), 4.12 (d, *J* = 5.1 Hz, 2H), 5.80 (br, 1H), 6.41 (br, 1H), 6.62 (br, 1H). HRMS (FAB) (M<sup>+</sup> + 1): Found: *m/z* 287.1614. Calcd for C<sub>13</sub>H<sub>23</sub>N<sub>2</sub>O<sub>5</sub>: 287.1607. When the methylene protons of the ethyl group were irradiated, 9.7%, 6.7%, and 2.7% of the NOE were observed for the methyl protons of the ethyl group, the olefinic proton, and the carbamate NH proton, respectively.

**Methyl (*E*)-2-(*t*-Butoxycarbonylamino)-2-pentenoylglycinate (31')**: A solid; IR (film) 3323, 2929, 2850, 1749, 1732, 1716, 1697, 1684, 1521, 1506, 1489, 1457, 1369, 1211, 1160, 1144, 1069, 1047, 1021, 984 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.10 (t, *J* = 7.6 Hz, 3H), 1.46 (s, 9H), 2.45 (dq, *J* = 7.3, 7.6 Hz, 2H), 3.78 (s, 3H), 4.14 (d, *J* = 5.1 Hz, 2H), 6.12 (br, 1H), 6.37 (br, 1H), 6.52 (br, 1H). HRMS (FAB) (M<sup>+</sup> + 1): Found: *m/z* 287.1600. Calcd for C<sub>13</sub>H<sub>23</sub>N<sub>2</sub>O<sub>5</sub>: 287.1607.

**Isomerization of the (*E*)- to (*Z*)-Isomer by Iodine.** To a solution of **22'** (105 mg, 0.32 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (2 mL) was added a solution of a catalytic amount of iodine in dry CH<sub>2</sub>Cl<sub>2</sub> (2 mL) at room temperature under a nitrogen. The solution was allowed to stand overnight with stirring, and was then concentrated in vacuo to give a residue, which was redissolved into AcOEt. The solution was washed with NaHSO<sub>3</sub>, NaHCO<sub>3</sub>, and brine, dried over MgSO<sub>4</sub>, and concentrated in vacuo to afford a residue, which was subjected to preparative TLC (SiO<sub>2</sub>, hexane:AcOEt = 2:1, v/v); (*Z*)-isomer 83 mg, (*E*)-isomer 5 mg.

Compounds **29'**, **30'**, and **31'** were isomerized to their (*Z*)-isomer in the same way. The results are summarized in Table 6.

**Synthesis of Protected Leucine-Enkephalin Analog (39).** ***t*-Butyl *N*-Boc- $\alpha$ -Tosylglycyl-L-leucinate (32):** To a suspension of *t*-butyl L-leucinate hydrochloride (985 mg, 44 mmol) in dry THF (2 mL) was added dropwise a solution of 4-methylmorpholine (546 mg, 5.4 mmol) in dry THF (2 mL) at 0 °C under air. After stirring for 1 h at room temperature, **16** (1.472 g, 4.5 mmol), DCC (927 mg, 4.5 mmol), and HOBt (608 mg, 4.5 mmol) were added. After the mixture was stirred for 23 h at room temperature,

the insoluble substances were filtered off and washed with AcOEt. The filtrate was concentrated in vacuo to afford a residue, which was dissolved into AcOEt. The AcOEt solution was successively washed with 10% citric acid, 10% NaHCO<sub>3</sub>, and brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated under reduced pressure to give the desired product as a mixture of diastereomers, which was recrystallized from ethyl acetate–hexane (1.726 g, 77%). Mp 143.0–146.0 °C (ethyl acetate–hexane);  $[\alpha]_D^{25} = -29.1^\circ$  (1.00, MeOH); IR (KBr) 3421, 3377, 2967, 2871, 1719, 1698, 1596, 1549, 1499, 1394, 1367, 1317, 1289, 1255, 1227, 1148, 1084, 1054, 950, 871, 848, 818, 737, 708, 666 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) (M)  $\delta$  0.99 (d,  $J = 6.6$  Hz, 6H), 1.21 (s, 9H), 1.48 (s, 9H), 1.64–1.71 (m, 2H), 1.80–1.92 (m, 1H), 2.42 (s, 3H), 4.49–4.63 (m, 1H), 5.46 (d,  $J = 3.5$  Hz, 1H), 5.91 (d,  $J = 8.8$  Hz, 1H), 7.18 (d,  $J = 7.5$  Hz, 1H), 7.35 (d,  $J = 3.5$  Hz, 2H), 7.79 (d,  $J = 5.3$  Hz, 2H). (m)  $\delta$  0.95 (d,  $J = 4.6$  Hz, 6H), 1.26 (s, 9H), 1.52 (s, 9H), 1.64–1.71 (m, 2H), 1.80–1.92 (m, 1H), 2.42 (s, 3H), 4.49–4.62 (m, 1H), 5.43 (d,  $J = 3.5$  Hz, 1H), 5.85 (d,  $J = 9.4$  Hz, 1H), 6.92 (d,  $J = 7.5$  Hz, 1H), 7.32 (d,  $J = 3.7$  Hz, 2H), 7.82 (d,  $J = 5.5$  Hz, 2H). Found: C, 57.68; H, 7.68; N, 5.55%. Calcd for C<sub>24</sub>H<sub>38</sub>N<sub>2</sub>O<sub>7</sub>S: C, 57.81; H, 7.68; N, 5.62%.

***t*-Butyl *N*-Boc-( $\beta$ -Nitro)phenylalanyl-L-leucinate (33):** To a suspension of KO<sup>t</sup>Bu (169 mg, 1.5 mmol) in dry THF (1.5 mL) was added dropwise a solution of (nitromethyl)benzene<sup>5a</sup> (384 mg, 2.8 mmol) in dry THF (2 mL) at -40 °C under nitrogen. After 10 min, a solution of **32** (249 mg, 0.5 mmol) in dry THF (3 mL) was added dropwise over 40 min. The reaction mixture was gradually warmed to room temperature and then quenched with a saturated aqueous solution of NH<sub>4</sub>Cl. The THF was removed in vacuo to afford a residue, which was partitioned between AcOEt and water. The aqueous solution was extracted with AcOEt. The combined extracts were washed with brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated under reduced pressure. The residue was subjected to preparative TLC (SiO<sub>2</sub>, hexane:ethyl acetate = 10/1, v/v) to give **33** as a mixture of diastereomers in 70% yield (168 mg). Mp 131.0–136.0 °C (ethyl acetate–hexane); IR (KBr) 3317, 2979, 2871, 2352, 1726, 1696, 1658, 1556, 1529, 1457, 1393, 1369, 1327, 1283, 1249, 1168, 1049, 1023, 947, 849, 803, 743, 717, 695 cm<sup>-1</sup>. The product was found to be a mixture of four diastereomers based on the NMR spectrum. Found: C, 59.92; H, 7.83; N, 8.71%. Calcd for C<sub>24</sub>H<sub>37</sub>N<sub>3</sub>O<sub>7</sub>: C, 60.11; H, 7.78; N, 8.76%.

***t*-Butyl *N*-Boc-Glycyl-( $\beta$ -nitro)phenylalanyl-L-leucinate (35):** A mixed solution of AcOEt (3 mL), abs. methanol (122 mg = 0.15 mL, 3.8 mmol), and acetyl chloride (299 mg = 0.27 mL, 3.8 mmol) was stirred for 30 min at room temperature under nitrogen. The solid **33** (108 mg, 0.22 mmol) was added to the resulting solution at 0 °C, and the solution was then stirred for 8 h at room temperature to afford the hydrogen chloride salt **34** in quantitative yield, which was used in the subsequent coupling reaction without further purification.

To a solution of *N*-Boc-Gly-OH (40 mg, 0.23 mmol) in dry THF (0.5 mL) was added a solution of isobutyl chloroformate (30 mg, 0.22 mmol) in dry THF (1.5 mL) at -15 °C under nitrogen, followed by the addition of a solution of NMM (24 mg, 0.23 mmol) in dry THF (1.5 mL). After 3 min, a solution of **34** and NMM (24 mg, 0.23 mmol) in dry DMF (1.5 mL) was added to the mixed-anhydride solution prepared above. The mixture was kept at -15 °C for 30 min and then stirred for 2 h at room temperature. After the insoluble substances were filtered off and washed with AcOEt, the filtrate was concentrated under reduced pressure to afford a residue, which was partitioned between AcOEt and wa-

ter. The aqueous layer was extracted with AcOEt and the combined extracts were successively washed with 10% citric acid, 10% NaHCO<sub>3</sub>, and brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated under reduced pressure. The residue was subjected to preparative TLC (SiO<sub>2</sub>, hexane:ethyl acetate = 2/1, v/v) to give **35** as a mixture of diastereomers in 76% yield (82 mg). Mp 114.0–116.0 °C (ethyl acetate–hexane); IR (KBr) 3308, 3068, 2961, 2872, 1732, 1699, 1657, 1556, 1471, 1456, 1368, 1280, 1247, 1152, 1047, 947, 847, 718, 694 cm<sup>-1</sup>. Found: C, 57.53; H, 7.17; N, 9.55%. Calcd for C<sub>24</sub>H<sub>37</sub>N<sub>3</sub>O<sub>7</sub>: C, 57.65; H, 7.14; N, 9.60%.

***t*-Butyl *N*-Boc-Glycyl- $\alpha,\beta$ -didehydrophenylalanyl-L-leucinate (36):** To a solution of **35** (29 mg, 0.06 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (0.5 mL) was added a solid of DABCO (7 mg, 0.06 mmol) at 0 °C under air. After stirring for 5 h at room temperature, an additional DABCO (42 mg, 0.36 mmol) was added. The solution was stirred for 18 h and quenched with a phosphate buffer solution (pH = 7). The solvent was removed in vacuo to afford a residue, which was partitioned between AcOEt and water. The aqueous layer was extracted with AcOEt, and the combined extracts were washed with brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated under reduced pressure. The residue was subjected to preparative TLC (SiO<sub>2</sub>, chloroform:methanol = 15/1, v/v) to afford **36** in 81% yield (22 mg). Mp 114.0–116.0 °C;  $[\alpha]_D^{25} = -8.9^\circ$  (0.42, MeOH); IR (KBr) 3365, 3290, 3056, 2976, 2932, 2871, 1725, 1687, 1657, 1623, 1523, 1457, 1391, 1367, 1274, 1253, 1153, 1050, 1030, 969, 946, 849, 764, 689 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.94 (d,  $J = 6.6$  Hz, 6H), 1.42 (s, 9H), 1.46 (s, 9H), 1.62 (dd,  $J = 7.1, 7.1$  Hz, 2H), 1.75 (tq,  $J = 6.6, 6.6$  Hz, 1H), 3.86 (d,  $J = 5.1$  Hz, 2H), 4.53 (dt,  $J = 7.3, 7.3$  Hz, 1H), 5.48 (t,  $J = 5.1$  Hz, 1H), 7.06 (d,  $J = 6.8$  Hz, 1H), 7.20 (s, 1H), 7.28–7.42 (m, 5H), 8.10 (s, 1H). EI-MS  $m/z$  447 (M<sup>+</sup>: 18.4%). When the amide NH proton of the  $\alpha,\beta$ -didehydrophenylalanine residue was irradiated, 3.2%, 2.1%, 1.4%, and 1.8% of the NOE were observed for the ortho-protons of the phenyl group, the NH and the methylene protons of the glycine residue, and the NH proton of the leucine residue.

***N,O*-(DiBoc)-L-Tyrosylglycine (38):** A mixed solution of Boc-Tyr(Boc)-OH<sup>10</sup> (2.830 g, 7.43 mmol), HONSu (941 mg, 8.17 mmol), and DCC (1.533 g, 7.43 mmol) in dry THF (40 mL) was stirred for 4 h at room temperature under air. After filtration, the filtrate was treated with an aqueous solution (3 mL) of H-Gly-OH (837 mg, 11.5 mmol) and NaHCO<sub>3</sub> (937 mg, 11.15 mmol) at room temperature overnight. The THF was removed in vacuo to give a residue, which was diluted with a small amount of water and extracted with AcOEt. The aqueous layer was acidified with 10% citric acid. After salting out, the liberated oil was extracted with AcOEt. The organic solution was washed with brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated under reduced pressure to afford the desired product, which was recrystallized from ethyl acetate (2.51 g, 77%). Mp 147.0–148.5 °C;  $[\alpha]_D^{25} = -1.2^\circ$  (1.0, MeOH); IR (KBr) 3310, 2935, 2981, 2602, 2345, 1760, 1655, 1532, 1510, 1455, 1394, 1369, 1335, 1280, 1257, 1225, 1152, 1105, 1049, 1019, 897, 846, 834, 782, 671 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.39 (s, 9H), 1.55 (s, 9H), 3.04 (br, 2H), 3.98 (br, 2H), 4.50 (br, 1H), 5.22 (br, 1H), 6.65 (br, 1H), 7.08 (d,  $J = 8.5$  Hz, 2H), 7.21 (d,  $J = 8.5$  Hz, 2H). One proton of the carboxylic acid was not assigned. Found: C, 57.32; H, 6.97; N, 6.34%. Calcd for C<sub>21</sub>H<sub>30</sub>N<sub>2</sub>O<sub>8</sub>: C, 57.52; H, 6.90; N, 6.39%.

***t*-Butyl *N,O*-(DiBoc)-L-Tyrosylglycylglycyl- $\alpha,\beta$ -didehydrophenylalanyl-L-leucinate (39):** To 1.5 mL of AcOEt was added methanol (60 mg = 0.076 mL, 1.87 mmol) followed by the addition of acetyl chloride (147 mg = 0.134 mL, 1.87 mmol) at room

temperature under nitrogen. The solution was stirred for 80 min at room temperature, and a solution of **36** (54 mg, 0.11 mmol) in ethyl acetate (2 mL) was then added at 0 °C. The solution was gradually warmed to room temperature and then stirred for 20 h. The solvent was removed under reduced pressure to afford the hydrogen chloride salt **37** in quantitative yield (47 mg), which was then used in the subsequent reaction. To a solution of **38** (44 mg, 0.10 mmol) in dry THF (2 mL) was added a solution of NMM (11 mg, 0.11 mmol) in dry THF (1 mL) followed by the addition of IBCF (15 mg, 0.11 mmol) in dry THF (1 mL) at -15 °C under nitrogen. After 30 min, a solution of the hydrogen chloride salt prepared above and NMM (12 mg, 0.12 mmol) in dry DMF (1 mL) was added while the solution was kept -15 °C for 15 min, and then gradually warmed to room temperature. After stirring overnight, the THF was removed in vacuo to afford a residue, which was partitioned between ether and water. The aqueous layer was extracted with ether. The combined extracts were successively washed with 10% citric acid, 10% NaHCO<sub>3</sub>, and brine, dried over anhydrous MgSO<sub>4</sub>, and concentrated under reduced pressure. The residual oil was subjected to preparative TLC (SiO<sub>2</sub>, chloroform:ethyl acetate:ethanol = 40:8:1, v/v). Yield, 64 mg (a solid, 72%); [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -1.4° (0.41, MeOH); IR (film) 3314, 3059, 2960, 2933, 2872, 2354, 1759, 1667, 1510, 1392, 1369, 1275, 1256, 1223, 1150, 1048, 1019, 970, 896, 846, 758, 693 cm<sup>-1</sup>; selected <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.36 (s, 9H), 1.45 (s, 9H), 1.55 (s, 9H); HRMS (FAB) (M<sup>+</sup> + 1): Found: *m/z* 810.4253. Calcd for C<sub>42</sub>H<sub>60</sub>N<sub>5</sub>O<sub>11</sub>: 810.4289. Amino acid analysis showed the presence of tyrosine, glycine, and leucine in the ratio of 0.61:2.00:1.10.

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## References

- a) T. Ueno, T. Nakashima, Y. Hayashi, and H. Fukami, *Agric. Biol. Chem.*, **39**, 1115 (1975). b) R. D. Durbin, "Toxins in Plant disease," Physiological Ecology Series, Academic Press (1981), p. 357. c) D. Botes, A. A. Tuinman, P. L. Wessels, C. C. Vilzoen, H. Kruger, D. H. Williams, S. Santikarn, R. J. Smith, and S. J. Hammond, *J. Chem. Soc., Perkin Trans. 1*, **1984**, 2311. d) M. F. Mackay, A. Van Donkelaar, and C. C. J. Culvenor, *J. Chem. Soc., Chem. Commun.*, **1986**, 1219. e) K. Nagaoka, M. Matsumoto, J. Oono, K. Yokoi, S. Ishizeki, and T. Nakashima, *J. Antibiot.*, **39**, 1527 (1986). f) K. L. Rinehart, K. Harada, M. Namikoshi, C. Chen, C. A. Harvis, M. H. G. Munro, J. W. Blunt, P. E. Mulligan, Y. R. Beasley, A. M. Dahlem, and W. W. Carmichael, *J. Am. Chem. Soc.*, **110**, 8557 (1988). g) G. R. Pettit, Y. Kamano, C. L. Herald, C. Dufresne, R. L. Cerny, D. L. Herald, J. M. Schmidt, and H. J. Kizu, *J. Am. Chem. Soc.*, **111**, 5015 (1989). h) E. D. de Silva, D. E. Williams, R. J. Anderson, H. Klux, C. F. B. Holmes, and T. M. Allen, *Tetrahedron Lett.*, **33**, 1561 (1992). i) L. D. Boeck, D. M. Berry, F. P. Mertz, and R. W. Wetzel, *J. Antibiot.*, **45**, 1222 (1992). j) L. M. Nogle, R. T. Williamson, and W. H. Gerwick, *J. Nat. Prod.*, **64**, 716 (2001).
- a) D. E. Palmer, C. Pattaroni, K. Nunami, R. K. Chadha, M. Goodman, T. Wakamiya, K. Fukase, S. Horimoto, M. Kitazawa, H. Fujita, A. Kubo, and T. Shiba, *J. Am. Chem. Soc.*, **114**, 5634 (1992). b) S. Bhatnager, G. S. Rao, and T. P. Singh, *BioSystems*, **34**, 143 (1995). c) G. Pletczynski and B. Rzeszotarska, *Pol. J. Chem.*, **69**, 1595 (1995). d) M. Crisma, F. Formaggio, C. Toniolo, T. Yoshikawa, and T. Wakamiya, *J. Am. Chem. Soc.*, **121**, 3272 (1999).
- M. L. English and C. H. Stammer, *Biochem. Biophys. Res. Commun.*, **85**, 1464 (1978).
- a) I. Photaki, *J. Am. Chem. Soc.*, **85**, 1123 (1963). b) J. W. Hines, E. G. Breitholle, and C. H. Stammer, *J. Org. Chem.*, **41**, 1466 (1976). c) N. Izumiya, S. Lee, T. Kanmera, and H. Aoyagi, *J. Am. Chem. Soc.*, **99**, 8346 (1977). d) A. Srinivasan, R. W. Stephenson, and R. K. Olsen, *J. Org. Chem.*, **42**, 2253 (1977). e) D. H. Rich and J. P. Tam, *J. Org. Chem.*, **42**, 3815 (1977). f) S. Nomoto, A. Sano, and T. Shiba, *Tetrahedron Lett.*, **20**, 521 (1979). g) C. Shin, Y. Yonezawa, and J. Yoshimura, *Tetrahedron Lett.*, **20**, 4085 (1979). h) Y. Yonezawa, C. Shin, Y. Ono, and J. Yoshimura, *Bull. Chem. Soc. Jpn.*, **53**, 2905 (1980). i) C. Shin, Y. Yonezawa, M. Takahashi, and J. Yoshimura, *Bull. Chem. Soc. Jpn.*, **54**, 1132 (1981). j) C. Shin, T. Yamada, and Y. Yonezawa, *Tetrahedron Lett.*, **24**, 2175 (1983). k) C. Shin, N. Takamatsu, and Y. Yonezawa, *Agric. Biol. Chem.*, **50**, 797 (1986). l) C. Shin, Y. Yonezawa, and M. Ikeda, *Bull. Chem. Soc. Jpn.*, **59**, 3573 (1986). m) D. Ranganathan, K. Shah, and N. Vaish, *J. Chem. Soc., Chem. Commun.*, **1992**, 1145. n) U. Schmidt, H. Griesser, V. Leitenberger, A. Lieberknecht, R. Mangold, R. Meyer, and B. Riedl, *Synthesis*, **1992**, 487. o) P. A. Goghlan and C. J. Easton, *Tetrahedron Lett.*, **40**, 4745 (1999). p) F. Yokokawa and T. Shioiri, *Tetrahedron Lett.*, **43**, 8673 (2002). q) F. Yokokawa and T. Shioiri, *Tetrahedron Lett.*, **43**, 8679 (2002). r) H. Saito, T. Yamada, K. Okumura, Y. Yonezawa, and C. Shin, *Chem. Lett.*, **2002**, 1098.
- a) T. Nagano and H. Kinoshita, *Bull. Chem. Soc. Jpn.*, **73**, 1605 (2000). b) R. Kimura, T. Nagano, and H. Kinoshita, *Bull. Chem. Soc. Jpn.*, **75**, 2517 (2002).
- C. J. Easton, P. D. Roselt, and E. R. T. Tiekink, *Tetrahedron*, **51**, 7809 (1995).
- a) Y. Shimohigashi, M. Waki, and N. Izumiya, *Tanpakushitsu, Kakusan Koso*, **28**, 1321 (1983). b) Y. Shimohigashi and C. H. Stammer, *J. Chem. Soc., Perkin Trans. 1*, **1983**, 803.
- E. Schnabel, H. Herzog, P. Hoffmann, E. Klauke, and I. Ugi, *Justus Liebigs Ann. Chem.*, **716**, 175 (1968).