

Innate immunity in the pathogenesis of cholangiopathy: A recent update

メタデータ	言語: eng 出版者: 公開日: 2017-10-03 キーワード (Ja): キーワード (En): 作成者: メールアドレス: 所属:
URL	http://hdl.handle.net/2297/33484

Review Article

Innate immunity in the pathogenesis of cholangiopathy, A recent update

Kenichi Harada and Yasuni Nakanuma

Department of Human Pathology, Kanazawa University Graduate School of Medicine,
Kanazawa, Japan

Address correspondence to:

Kenichi Harada, M.D.

Department of Human Pathology
Kanazawa University Graduate School of Medicine
Kanazawa 920-8640, Japan
FAX: (0)76-234-4229 (Japan)
TEL: (0)76-265-2199 (Japan)
E-mail: kenichih@med.kanazawa-u.ac.jp

ABSTRACT

Biliary innate immunity is involved in the pathogenesis of cholangiopathies in patients with various biliary diseases. Biliary epithelial cells possess an innate immune system consisting of the Toll-like receptor (TLR) family and recognize pathogen-associated molecular patterns (PAMPs). Recently, regulatory mechanisms by intracellular negative regulators including peroxisome proliferator-activated receptor- γ and micro-RNA have been clarified. In primary biliary cirrhosis (PBC) and primary sclerosing cholangitis, dysregulated biliary innate immunity, namely hyper-responsiveness to PAMPs, is associated with the histopathogenesis of cholangiopathy. Moreover, biliary epithelial cells produce monocyte chemoattractant protein-1 (MCP-1/CCL2) as a result of the innate immune response and bile ductules play a role in hepatic fibrosis caused by hepatic stellate cells (HSCs). Also, biliary innate immune responses induce the production of two chemokines, fractalkine and macrophage inflammatory protein-3 α (MIP-3 α), causing the migration of inflammatory cells and a population of antigen-presenting cell found in epithelium, Langerhans cell, and involve chronic cholangitis associated with biliary epithelium-specific innate and acquired immunity in PBC.

KEY WORDS : biliary epithelial cell, cholangiopathy, innate immunity, Toll-like receptor, chemokine, primary biliary cirrhosis

Introduction

Infectious agents are involved in various cholangiopathies including cholangitis, bile duct loss, and lithiasis as either etiopathological or aggravating factors. Several enterobacteria and viruses are speculated to be primary or secondary factors for primary biliary cirrhosis (PBC), primary sclerosing cholangitis (PSC), biliary atresia, and biliary lithiasis. Such cholangiopathy is closely associated with biliary innate immunity and the periductal cytokine network [1-7]. Epithelial cells as well as immunocompetent cells such as dendritic cells (DCs) possess pathogen-recognition receptors such as Toll-like receptors (TLRs) reflecting the proper micro-environment and function of each epithelial cell type. Biliary epithelial cells (BECs, cholangiocytes) also possess TLR-related innate immune systems [6, 8-10]. Because no microorganisms showing BEC-specific tropism have been identified, an innate immune response specific to BEC rather than pathogen-associated molecular patterns (PAMPs) is important in the pathogenesis of cholangiopathy. We have previously reviewed the basic molecular mechanism of biliary innate immunity in this journal [11]. In this review, we attempted to summarize recent progress in the study of the pathophysiological role of biliary innate immunity in the mechanisms of cholangiopathy, mainly based on our own data ([Table 1](#)).

Regulatory system of biliary innate immunity: update

Human BECs possess functional PAMP-recognizing receptors including at least TLR1-5 ([Table 1](#)). These TLRs are distributed throughout the intrahepatic biliary tree in normal and diseased human livers, irrespective of anatomical level, implying that BECs participate directly in innate immunity and show a prompt response to pathogens

without any help from immunocompetent cells [6, 9, 10, 13, 18, 28, 29].

Although the luminal surface of the bile duct is continually exposed to PAMPs in bile, mainly originating from the intestines, BECs physiologically do not elicit an inflammatory response. Tight control of TLR signaling provides tolerance to physiological amounts of intestinal endotoxin in bile to avoid constant innate immune activation in BECs. This failure to respond to PAMPs, especially LPS, could be due to "endotoxin tolerance." In human BECs, IRAK-M and peroxisome proliferator-activated receptor γ (PPAR γ) are closely associated with the maintenance of biliary homeostasis as a regulator of tolerance by attenuating inflammatory signals to commensal PAMPs in bile, as summarized in our previous review of this journal [11].

Recently, novel regulatory mechanisms of biliary innate immunity associated with the TLR system have been clarified. Micro-RNAs play important roles in a wide range of biological events through post-transcriptional suppression of target mRNAs. Recent studies indicate Micro-RNA-mediated post-transcriptional pathways to be critical to host-cell regulatory responses to microbial infections. Cultured human BECs express let-7 family members which post-transcriptionally down-regulate TLR4 expression and infections of *C. parvum* decrease the expression of let-7 resulting in the up-regulation of TLR4 [16]. Moreover, microRNA-98 and let-7, suppressing cytokine-inducible Src homolog 2-containing protein (CIS, a suppressor of cytokine signaling family) at the translational level, are expressed in BECs, and LPS and *C. parvum* infections downregulate these micro-RNAs, suggesting the regulation of the TLR-mediated biliary innate immune response [17].

In PBC, compared with Th2, a Th1-dominant cytokine milieu is associated with the pathogenesis including bile duct injury [30]. BECs possess the receptor for IFN- γ

(Th1 cytokine) and IFN- γ up-regulates the expression of TLRs and susceptibility to PAMPs in BECs, as mentioned above, impairing the regulation of biliary innate immunity. The upregulation of TLR4 and TLR9 in BECs and of TLR3 and type I IFN signaling pathways in portal tracts and parenchyma are found in PBC [28, 29, 31]. Moreover, IL-4 (Th2 cytokine) and IFN- γ up- and down-regulate the expression of PPAR γ in cultured human BECs [15, 32]. In PBC liver, PPAR γ expression is significantly down-regulated in the affected bile ducts as a Th1-dominant periductal cytokine milieu [15]. These findings indicate an increased susceptibility to PAMPs, suggesting an association with the pathogenesis of cholangiopathy in PBC.

Moreover, in PSC, inappropriate biliary innate immune responses to intestinal endotoxin and subsequent endotoxin intolerance have been reported to contribute to the development and perpetuation of chronic cholangitis [33]. BECs from PSC livers show reversibly increased TLR expression and activation of TLR-related intracellular signaling. Consecutively, PSC BECs exhibit inappropriate innate immune responses to endotoxin and do not develop tolerance after repeated exposure to endotoxin. This hyper-responsiveness is probably because of the stimulatory effect of abundantly expressed IFN- γ and TNF- α in PSC livers, which stimulate TLR4-mediated endotoxin signaling in BECs, leading to increased TLR4-mediated endotoxin incorporation and impaired inactivation of the TLR4 signaling cascade.

Hepatic fibrosis and biliary innate immunity

A major feature of chronic liver diseases, hepatic fibrosis, spreads within portal tracts and also periportal areas in patients with hepatitic and cholestatic liver diseases, irrespective of etiology. In particular, a fibrogenic cytokine-rich environment caused by

inflammation and infection is closely associated with hepatic fibrosis.

Periportal fibrosis is thought to be associated with the accumulation and activation of hepatic stellate cells (HSCs). In damaged liver, HSCs are activated, proliferate, and migrate into the injured area in response to the chemoattractive effects of chemokines and are considered the most important effector cells associated with fibrogenesis in hepatic parenchyma including the interface between portal tracts and periportal hepatocytes, and the fibrous enlargement of portal tracts and fibrous extension from portal areas are closely associated with activated HSCs and their transformed version, myofibroblasts. Then, HSCs, particularly activated HSCs, migrate into damaged areas in response to a chemokine, secreting monocyte chemoattractant protein-1 (MCP-1/CCL2) which attracts monocyte/macrophages [34].

BECs directly promote fibrogenesis by a number of mechanisms including the synthesis of matrix constituents and the release of fibrogenic cytokines such as MCP-1, platelet-derived growth factor, TGF- β , connective tissue growth factor (CTGF), and endothelin-1 [35]. Moreover, only the expression of MCP-1 is positively regulated by TLR ligands (PAMPs) occurs in an NF- κ B-dependent manner, suggesting that the biliary innate immune response is a critical trigger of MCP-1 production.

Proliferating bile ductules are thought to be part of a non-specific reaction in various hepatobiliary diseases, and bile ductules located between interlobular bile ducts in portal tracts and bile canaliculi in hepatocytes are frequently increased in number under a variety of pathologic conditions of the liver and take part in hepatic fibrosis and the progression of hepatobiliary diseases [26, 36, 37]. MCP-1 is expressed in bile ductules in areas of interface hepatitis (Fig.1), whereas normal livers lacked these findings. Moreover, α -smooth muscle actin (α -SMA)-positive activated HSCs

(myofibroblasts) are accumulated around MCP-1-expressing bile ductules (Fig.1), suggesting that MCP-1 derived from BECs consisting of bile ductules plays a role in the chemoattraction of HSCs. Because the interface areas are rich in several cytokines caused by immune-mediated (neco)inflammatory reactions against virus-infected hepatocytes and bile-derived PAMPs, BECs in bile ductules could produce MCP-1, particularly, via biliary innate immunity, suggesting that biliary innate immunity directly plays an important role in the recruitment of HSCs to interface areas and the activation of HSCs resulting in the progression of periportal fibrosis.

Fractalkine and innate immunity in PBC

PBC mainly affects middle-aged females, and histologically, the interlobular bile ducts are primarily damaged and show characteristic findings such as chronic nonsuppurative destructive cholangitis (CNSDC) followed by progressive bile duct loss [38]. There is considerable evidence that the damage of bile ducts is mediated by autoreactive or cytotoxic T cells [39], and molecular mechanisms of the migration of pathogenic T cells around or within bile ducts have gradually been clarified.

There have been several reports that bacterial components such as lipopolysaccharide (LPS) and DNA fragments are detectable in pathologic bile of patients with PBC [5, 6], and also endotoxin and lipoteichoic acids abnormally accumulate in or around the intrahepatic bile ducts of PBC [5-7, 40, 41]. Unusual immune responses to these infectious agents or their components are now suspected to underlie the etiopathogenesis of PBC [42-45].

The chemokine-adhesion molecule, fractalkine (CX3CL1), plays an important role in the migration of leukocytes to target sites under physiological as well as

pathological conditions. Unlike other chemokines, fractalkine is expressed as a membrane-bound form on cells and also can be shed by ADAM10 or ADAM17 as a soluble chemotactic form. Vascular endothelial cells express fractalkine and its soluble form is a potent chemoattractant for CD16⁺NK cells, CD8⁺ cytotoxic T cells, CD4⁺T cells, $\gamma\delta$ T cells, monocytes, mature macrophages, and mucosal DC cells, expressing its receptor (CX3CR1). The level of soluble fractalkine is significantly elevated in the sera of PBC patients and also *in situ* expression of fractalkine in small bile ducts, particularly damaged bile ducts, is upregulated in PBC [24] (Fig.2), suggesting that fractalkine is an important mediator associated with the continuous portal, particularly periductal, inflammation of PBC.

Upregulation of fractalkine expression in BECs is induced by PAMPs including LPS, suggesting that the dysregulated biliary innate immunity against commensal or pathological PAMPs is closely associated with the production of fractalkine in BECs and that secreted soluble fractalkine may be involved in the chemoattraction of infiltrating cells expressing CX3CR1 around these pathological bile ducts and in the recruitment of biliary intraepithelial leukocytes. Moreover, the up-regulation of fractalkine expression in BECs is promoted by TLR3 ligand (poly(I:C)), via activated monocytes through direct contact and the secretion of TNF- α by monocytes [25]. Intraepithelial lymphocytes in damaged bile ducts of PBC patients contain a significantly higher percentage of CD8⁺ T cells and show strongly increased cytolytic activity against epithelial-derived target cells [46]. Then, CX3CR1-positive leukocytes invade into bile ducts by engaging fractalkine, thereby causing cholangitis including CNSDC [25].

In addition to PBC, fractalkine is associated with the chronic inflammation in

other diseases including rheumatoid arthritis and inflammatory bowel disease. Recently, an anti-fractalkine monoclonal antibody has been reported to almost completely suppress the clinicopathological activities in inflammatory bowel disease models [47]. Therefore, fractalkine-CX3CR1 signaling might be a molecular target for the treatment of PBC.

Antigen-presenting cells and biliary innate immunity in PBC

The presence and response to mitochondrial autoantigen(s) of autoreactive T cells [39] is believed to require interaction with professional antigen-presenting cells (APCs) such as DCs. DCs are generally classified into myeloid DCs (mDCs) and plasmacytoid DCs (pDCs) and reside in tissues of most organs under normal conditions. In the periphery, immature DCs such as Langerhans cells (LCs, a population of DCs only found in epithelia) capture antigens and, under the influence of inflammatory stimuli, subsequently migrate. The presence of APCs in biliary trees is likely important to the bile duct-specific autoimmunity in PBC. In the inflamed bile ducts, several DCs including immature DCs [48], S-100⁺ DCs,[49] and LCs[50, 51] are found, indicating a role in the initiation of inflammatory responses in cases of PBC. Moreover, periductal DCs are mostly mDCs and the major representative type of mDCs, Langerin⁺ LCs, are attached to or embedded in the damaged bile ducts of PBC patients (Fig.3) [23], suggesting that LCs existing around and within bile ducts play an important role as APCs presenting bile duct-derived autoantigens.

Because LCs possess chemokine receptors (CCR4 and CCR6), ligands of these receptors including CCL17, CCL22, and MIP-3 α (CCL20) could cause LCs to migrate. Among them, MIP-3 α plays a central role in recruiting LC precursors into the

epithelium during inflammation and also an important role in the pathogenesis of ulcerative colitis [52, 53]. In PBC, MIP-3 α -expressing BECs are frequently found in the damaged bile ducts accompanying Langerin⁺ LCs, suggesting that the BECs producing MIP-3 α directly play an important role in the migration of LCs in biliary layers. Moreover, the expression of MIP-3 α is selectively restricted at sites of inflammation including cholangitis in PBC (Fig.3) and bile duct-related acquired immunity is speculated to be closely associated with the periductal cytokine milieu and biliary innate immunity [6, 9, 15, 24, 30]. Because human BECs possess receptors for some cytokines (IL-1 β , IL-6, IL-4, IFN- γ , TNF- α , and IL-17) as well as TLRs [9, 21], they could show a response to these cytokines as well as PAMPs and induce various immunological reactions. IL-17 and two inflammatory cytokines (IL-1 β and TNF- α) significantly upregulate the expression of MIP-3 α in human BECs. IL-17-positive cells (namely Th17 cells) are accumulated around the damaged bile ducts in PBC and the expression of IL-1 β and TNF- α was increased in damaged bile ducts including CNSDC of PBC patients [20, 21]. Therefore, the production of MIP-3 α in PBC is closely associated with periductal cytokines including IL-1 β , TNF- α , and IL-17. In addition to these cytokine milieus, the damaged bile ducts and/or portal tracts show an increased susceptibility to PAMPs and enhanced production and secretion of innate inflammatory mediators in PBC [15, 29]. PAMPs directly induce the production of MIP-3 α in human BECs [23]. Therefore, LCs existing around or within biliary epithelial layers are induced to migrate via BEC-derived MIP-3 α expression evoked via a PBC-specific periductal cytokine milieu and biliary innate immunity and are important as biliary epithelium-specific APCs in the development of acquired immunity and the aggravating and continuous cholangitis in PBC.

Conclusion and perspectives

Biliary innate immunity consisting of an bile duct-specific system is important for the mucosal immunity in intrahepatic and extrahepatic bile ducts and also associated with the pathogenesis of several cholangiopathies in biliary diseases as well as defense against microbial infections. We speculate that biliary innate immunity is solely associated with the etiology of biliary diseases as the initial event and that the presence of causative microorganisms is not necessary in the pathogenesis of cholangiopathy caused by a subsequent acquired immunity. It is mandatory to understand the molecular basis underlying the immunophysiology and immunopathology of cholangiopathy in terms of innate as well as acquired immunity.

DISCLOSURE

This manuscript is an updated version of my previously published manuscript.

REFERENCE

- [1] Xu L, Shen Z, Guo L, Fodera B, Keogh A, Joplin R, O'Donnell B, Aitken J, Carman W, Neuberger J, Mason A. Does a betaretrovirus infection trigger primary biliary cirrhosis? *Proc Natl Acad Sci U S A* 2003; 100(14):8454-9.
- [2] Tyler KL, Sokol RJ, Oberhaus SM, Le M, Karrer FM, Narkewicz MR, Tyson RW, Murphy JR, Low R, Brown WR. Detection of reovirus RNA in hepatobiliary tissues from patients with extrahepatic biliary atresia and choledochal cysts. *Hepatology* 1998; 27(6):1475-82.
- [3] Nilsson I, Kornilovs'ka I, Lindgren S, Ljungh A, Wadstrom T. Increased prevalence of seropositivity for non-gastric *Helicobacter* species in patients with autoimmune liver disease. *J Med Microbiol* 2003; 52(Pt 11):949-53.
- [4] Fox JG, Dewhirst FE, Shen Z, Feng Y, Taylor NS, Paster BJ, Ericson RL, Lau CN, Correa P, Araya JC, Roa I. Hepatic *Helicobacter* species identified in bile and gallbladder tissue from Chileans with chronic cholecystitis. *Gastroenterology* 1998; 114(4):755-63.
- [5] Hiramatsu K, Harada K, Tsuneyama K, Sasaki M, Fujita S, Hashimoto T, Kaneko S, Kobayashi K, Nakanuma Y. Amplification and sequence analysis of partial bacterial 16S ribosomal RNA gene in gallbladder bile from patients with primary biliary cirrhosis. *J Hepatol* 2000; 33(1):9-18.
- [6] Harada K, Ohira S, Isse K, Ozaki S, Zen Y, Sato Y, Nakanuma Y. Lipopolysaccharide activates nuclear factor-kappaB through toll-like receptors and related molecules in cultured biliary epithelial cells. *Lab Invest* 2003; 83(11):1657-67.
- [7] Tsuneyama K, Harada K, Kono N, Hiramatsu K, Zen Y, Sudo Y, Gershwin ME, Ikemoto M, Arai H, Nakanuma Y. Scavenger cells with gram-positive bacterial lipoteichoic acid infiltrate around the damaged interlobular bile ducts of primary biliary cirrhosis. *J Hepatol* 2001; 35(2):156-63.
- [8] Harada K, Isse K, Sato Y, Ozaki S, Nakanuma Y. Endotoxin tolerance in human intrahepatic biliary epithelial cells is induced by upregulation of IRAK-M. *Liver Int* 2006; 26(8):935-42.
- [9] Harada K, Isse K, Nakanuma Y. Interferon gamma accelerates NF-kappaB activation of biliary epithelial cells induced by Toll-like receptor and ligand interaction. *J Clin Pathol* 2006; 59(2):184-90.
- [10] Harada K, Sato Y, Itatsu K, Isse K, Ikeda H, Yasoshima M, Zen Y, Matsui A,

- Nakanuma Y. Innate immune response to double-stranded RNA in biliary epithelial cells is associated with the pathogenesis of biliary atresia. *Hepatology* 2007; 46(4):1146-1154.
- [11] Harada K, Nakanuma Y. Biliary innate immunity in the pathogenesis of biliary diseases. *Inflamm Allergy Drug Targets* 2010; 9(2):83-90.
- [12] Chen XM, O'Hara SP, Nelson JB, Splinter PL, Small AJ, Tietz PS, Limper AH, LaRusso NF. Multiple TLRs are expressed in human cholangiocytes and mediate host epithelial defense responses to *Cryptosporidium parvum* via activation of NF-kappaB. *J Immunol* 2005; 175(11):7447-56.
- [13] Yokoyama T, Komori A, Nakamura M, Takii Y, Kamihira T, Shimoda S, Mori T, Fujiwara S, Koyabu M, Taniguchi K, Fujioka H, Migita K, Yatsushashi H, Ishibashi H. Human intrahepatic biliary epithelial cells function in innate immunity by producing IL-6 and IL-8 via the TLR4-NF-kappaB and -MAPK signaling pathways. *Liver Int* 2006; 26(4):467-76.
- [14] Harada K, Sato Y, Isse K, Ikeda H, Nakanuma Y. Induction of innate immune response and absence of subsequent tolerance to dsRNA in biliary epithelial cells relate to the pathogenesis of biliary atresia. *Liver Int* 2008; 28(5):614-21.
- [15] Harada K, Isse K, Kamihira T, Shimoda S, Nakanuma Y. Th1 cytokine-induced downregulation of PPARgamma in human biliary cells relates to cholangitis in primary biliary cirrhosis. *Hepatology* 2005; 41(6):1329-38.
- [16] Chen XM, Splinter PL, O'Hara SP, LaRusso NF. A cellular micro-RNA, let-7i, regulates Toll-like receptor 4 expression and contributes to cholangiocyte immune responses against *Cryptosporidium parvum* infection. *J Biol Chem* 2007; 282(39):28929-38.
- [17] Hu G, Zhou R, Liu J, Gong AY, Eischeid AN, Dittman JW, Chen XM. MicroRNA-98 and let-7 confer cholangiocyte expression of cytokine-inducible Src homology 2-containing protein in response to microbial challenge. *J Immunol* 2009; 183(3):1617-24.
- [18] Harada K, Ohba K, Ozaki S, Isse K, Hirayama T, Wada A, Nakanuma Y. Peptide antibiotic human beta-defensin-1 and -2 contribute to antimicrobial defense of the intrahepatic biliary tree. *Hepatology* 2004; 40(4):925-932.
- [19] Zen Y, Harada K, Tsuneyama K, Katayanagi K, Yamamoto Y, Nakanuma Y. Lipopolysaccharide induces overexpression of MUC2 and MUC5AC in cultured biliary epithelial cells: possible key phenomenon of hepatolithiasis. *Am J Pathol* 2002; 161(4):1475-84.
- [20] Yasoshima M, Kono N, Sugawara H, Katayanagi K, Harada K, Nakanuma Y.

- Increased expression of interleukin-6 and tumor necrosis factor-alpha in pathologic biliary epithelial cells: in situ and culture study. *Lab Invest* 1998; 78(1):89-100.
- [21] Harada K, Shimoda S, Sato Y, Isse K, Ikeda H, Nakanuma Y. Periductal interleukin-17 production in association with biliary innate immunity contributes to the pathogenesis of cholangiopathy in primary biliary cirrhosis. *Clin Exp Immunol* 2009; 157:261-270.
- [22] Harada K, Chiba M, Okamura A, Hsu M, Sato Y, Igarashi S, Ren XS, Ikeda H, Ohta H, Kasashima S, Kawashima A, Nakanuma Y. Monocyte chemoattractant protein-1 derived from biliary innate immunity contributes to hepatic fibrogenesis. *J Clin Pathol* 2011; 64(8):660-5.
- [23] Harada K, Shimoda S, Ikeda H, Chiba M, Hsu M, Sato Y, Kobayashi M, Ren XS, Ohta H, Kasashima S, Kawashima A, Nakanuma Y. Significance of periductal Langerhans cells and biliary epithelial cell-derived macrophage inflammatory protein-3alpha in the pathogenesis of primary biliary cirrhosis. *Liver Int* 2011; 31(2):245-53.
- [24] Isse K, Harada K, Zen Y, Kamihira T, Shimoda S, Harada M, Nakanuma Y. Fractalkine and CX3CR1 are involved in the recruitment of intraepithelial lymphocytes of intrahepatic bile ducts. *Hepatology* 2005; 41(3):506-16.
- [25] Shimoda S, Harada K, Niino H, Taketomi A, Maehara Y, Tsuneyama K, Kikuchi K, Nakanuma Y, Mackay IR, Gershwin ME, Akashi K. CX3CL1 (fractalkine): A signpost for biliary inflammation in primary biliary cirrhosis. *Hepatology* 2009.
- [26] Isse K, Harada K, Nakanuma Y. IL-8 expression by biliary epithelial cells is associated with neutrophilic infiltration and reactive bile ductules. *Liver Int* 2007; 27(5):672-80.
- [27] Sawada S, Harada K, Isse K, Sato Y, Sasaki M, Kaizaki Y, Nakanuma Y. Involvement of *Escherichia coli* in pathogenesis of xanthogranulomatous cholecystitis with scavenger receptor class A and CXCL16-CXCR6 interaction. *Pathol Int* 2007; 57(10):652-63.
- [28] Wang AP, Migita K, Ito M, Takii Y, Daikoku M, Yokoyama T, Komori A, Nakamura M, Yatsunami H, Ishibashi H. Hepatic expression of toll-like receptor 4 in primary biliary cirrhosis. *J Autoimmun* 2005; 25(1):85-91.
- [29] Takii Y, Nakamura M, Ito M, Yokoyama T, Komori A, Shimizu-Yoshida Y, Nakao R, Kusumoto K, Nagaoka S, Yano K, Abiru S, Ueki T, Matsumoto T, Daikoku M, Taniguchi K, Fujioka H, Migita K, Yatsunami H, Nakashima M, Harada M, Ishibashi H. Enhanced expression of type I interferon and toll-like receptor-3 in primary biliary cirrhosis. *Lab Invest* 2005; 85(7):908-20.

- [30] Harada K, Van de Water J, Leung PS, Coppel RL, Ansari A, Nakanuma Y, Gershwin ME. In situ nucleic acid hybridization of cytokines in primary biliary cirrhosis: predominance of the Th1 subset. *Hepatology* 1997; 25(4):791-6.
- [31] Karrar A, Broome U, Sodergren T, Jaksch M, Bergquist A, Bjornstedt M, Sumitran-Holgersson S. Biliary epithelial cell antibodies link adaptive and innate immune responses in primary sclerosing cholangitis. *Gastroenterology* 2007; 132(4):1504-14.
- [32] Nakajima T, Kamijo Y, Tanaka N, Sugiyama E, Tanaka E, Kiyosawa K, Fukushima Y, Peters JM, Gonzalez FJ, Aoyama T. Peroxisome proliferator-activated receptor alpha protects against alcohol-induced liver damage. *Hepatology* 2004; 40(4):972-80.
- [33] Mueller T, Beutler C, Pico AH, Shibolet O, Pratt DS, Pascher A, Neuhaus P, Wiedenmann B, Berg T, Podolsky DK. Enhanced innate immune responsiveness and intolerance to intestinal endotoxins in human biliary epithelial cells contributes to chronic cholangitis. *Liver Int* 2011; 31(10):1574-88.
- [34] Marra F, Romanelli RG, Giannini C, Failli P, Pastacaldi S, Arrighi MC, Pinzani M, Laffi G, Montalto P, Gentilini P. Monocyte chemotactic protein-1 as a chemoattractant for human hepatic stellate cells. *Hepatology* 1999; 29(1):140-8.
- [35] Kinnman N, Housset C. Peribiliary myofibroblasts in biliary type liver fibrosis. *Front Biosci* 2002; 7:d496-503.
- [36] Nakanuma Y, Hosono M, Sanzen T, Sasaki M. Microstructure and development of the normal and pathologic biliary tract in humans, including blood supply. *Microsc Res Tech* 1997; 38(6):552-70.
- [37] Harada K, Kono N, Tsuneyama K, Nakanuma Y. Cell-kinetic study of proliferating bile ductules in various hepatobiliary diseases. *Liver* 1998; 18(4):277-84.
- [38] Nakanuma Y, Ohta G. Histometric and serial section observations of the intrahepatic bile ducts in primary biliary cirrhosis. *Gastroenterology* 1979; 76(6):1326-32.
- [39] Shimoda S, Nakamura M, Ishibashi H, Hayashida K, Niho Y. HLA DRB4 0101-restricted immunodominant T cell autoepitope of pyruvate dehydrogenase complex in primary biliary cirrhosis: evidence of molecular mimicry in human autoimmune diseases. *J Exp Med* 1995; 181(5):1835-45.
- [40] Sasatomi K, Noguchi K, Sakisaka S, Sata M, Tanikawa K. Abnormal accumulation of endotoxin in biliary epithelial cells in primary biliary cirrhosis and primary sclerosing cholangitis. *J Hepatol* 1998; 29(3):409-16.
- [41] Haruta I, Hashimoto E, Kato Y, Kikuchi K, Kato H, Yagi J, Uchiyama T, Kobayashi M, Shiratori K. Lipoteichoic acid may affect the pathogenesis of bile duct damage in

- primary biliary cirrhosis. *Autoimmunity* 2006; 39(2):129-35.
- [42] Schiffenbauer J, Soos J, Johnson H. The possible role of bacterial superantigens in the pathogenesis of autoimmune disorders. *Immunol Today* 1998; 19(3):117-20.
- [43] Shimoda S, Nakamura M, Ishibashi H, Kawano A, Kamihira T, Sakamoto N, Matsushita S, Tanaka A, Worman HJ, Gershwin ME, Harada M. Molecular mimicry of mitochondrial and nuclear autoantigens in primary biliary cirrhosis. *Gastroenterology* 2003; 124(7):1915-25.
- [44] Harada K, Tsuneyama K, Sudo Y, Masuda S, Nakanuma Y. Molecular identification of bacterial 16S ribosomal RNA gene in liver tissue of primary biliary cirrhosis: is *Propionibacterium acnes* involved in granuloma formation? *Hepatology* 2001; 33(3):530-6.
- [45] Selmi C, Balkwill DL, Invernizzi P, Ansari AA, Coppel RL, Podda M, Leung PS, Kenny TP, Van De Water J, Nantz MH, Kurth MJ, Gershwin ME. Patients with primary biliary cirrhosis react against a ubiquitous xenobiotic-metabolizing bacterium. *Hepatology* 2003; 38(5):1250-7.
- [46] Isse K, Harada K, Sato Y, Nakanuma Y. Characterization of biliary intra-epithelial lymphocytes at different anatomical levels of intrahepatic bile ducts under normal and pathological conditions: numbers of CD4+CD28- intra-epithelial lymphocytes are increased in primary biliary cirrhosis. *Pathol Int* 2006; 56(1):17-24.
- [47] Nishimura M, Kuboi Y, Muramoto K, Kawano T, Imai T. Chemokines as novel therapeutic targets for inflammatory bowel disease. *Ann N Y Acad Sci* 2009; 1173:350-6.
- [48] Tanimoto K, Akbar SM, Michitaka K, Onji M. Immunohistochemical localization of antigen presenting cells in liver from patients with primary biliary cirrhosis: highly restricted distribution of CD83-positive activated dendritic cells. *Pathol Res Pract* 1999; 195(3):157-62.
- [49] Demetris AJ, Sever C, Kakizoe S, Oguma S, Starzl TE, Jaffe R. S100 protein positive dendritic cells in primary biliary cirrhosis and other chronic inflammatory liver diseases. Relevance to pathogenesis? *Am J Pathol* 1989; 134(4):741-7.
- [50] Tobe K, Endo H, Nagashima H, Kobayashi T. Langerhans cells among bile duct epithelial cells in chronic liver disease. *Acta Pathol Jpn* 1982; 32(1):93-9.
- [51] Ballardini G, Fallani M, Bianchi FB, Pisi E. Antigen presenting cells in liver biopsies from patients with primary biliary cirrhosis. *Autoimmunity* 1989; 3(2):135-44.
- [52] Watanabe S, Yamakawa M, Hiroaki T, Kawata S, Kimura O. Correlation of dendritic cell infiltration with active crypt inflammation in ulcerative colitis. *Clin*

Immunol 2007; 122(3):288-97.

- [53] Dieu-Nosjean MC, Massacrier C, Homey B, Vanbervliet B, Pin JJ, Vicari A, Lebecque S, Dezutter-Dambuyant C, Schmitt D, Zlotnik A, Caux C. Macrophage inflammatory protein 3alpha is expressed at inflamed epithelial surfaces and is the most potent chemokine known in attracting Langerhans cell precursors. *J Exp Med* 2000; 192(5):705-18.

Table 1

Summary of biliary innate immunity

Receptors recognizing PAMPs**TLRs**

TLR1 [12, 13]

TLR2 [9, 12, 13]

TLR3 [9, 12-14]

TLR4 [6, 9, 12, 13]

TLR5 [9, 12, 13]

TLR6 [9, 12, 13]

TLR7 [12]

TLR8 [12]

TLR9 [12, 13]

RIG-I [10]

MDA-5 [10]

Negative regulators of TLR signalings

IRAK-M [8]

PPAR- γ [15]

micro-RNA [16, 17]

Antibiotics, cytokine, and chemokines produced via biliary innate immunity

defensin [18]

IFN- β 1 [10]TNF- α [19, 20]

IL-6 [13, 20, 21]

IL-1 β [21]

IL-23 p19 [21]

MCP-1[13, 22]

MIP-3 α [23]

Fractalkine (CX3CL1) [24, 25]

IL-8 [13, 26]

CXCL16 [27]

Parentheses denote reference numbers.

Figures and legends

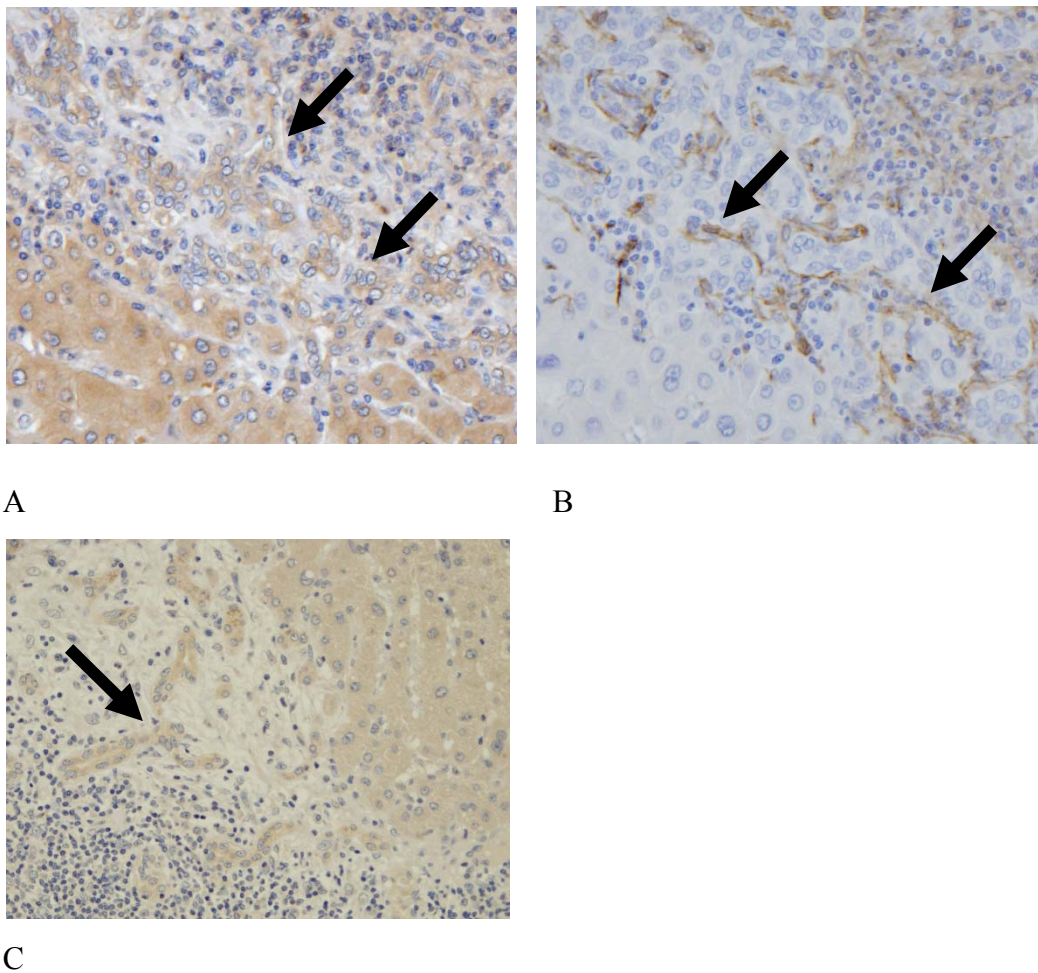
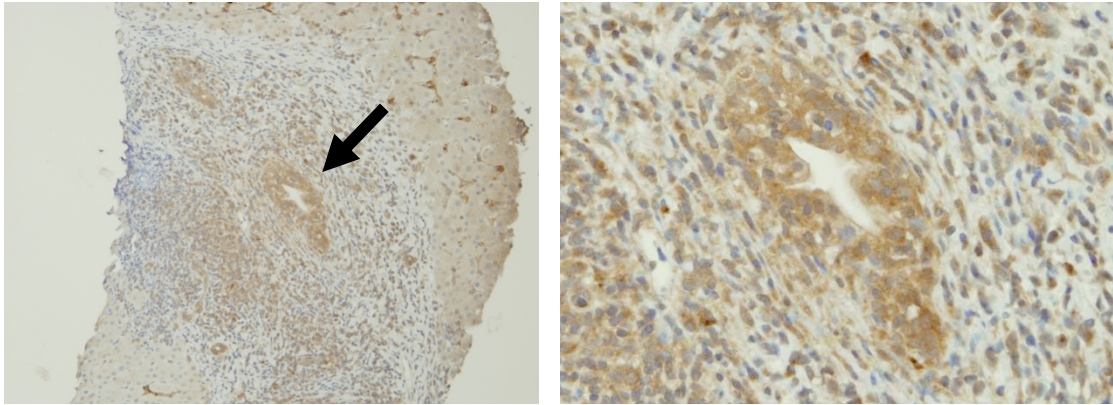


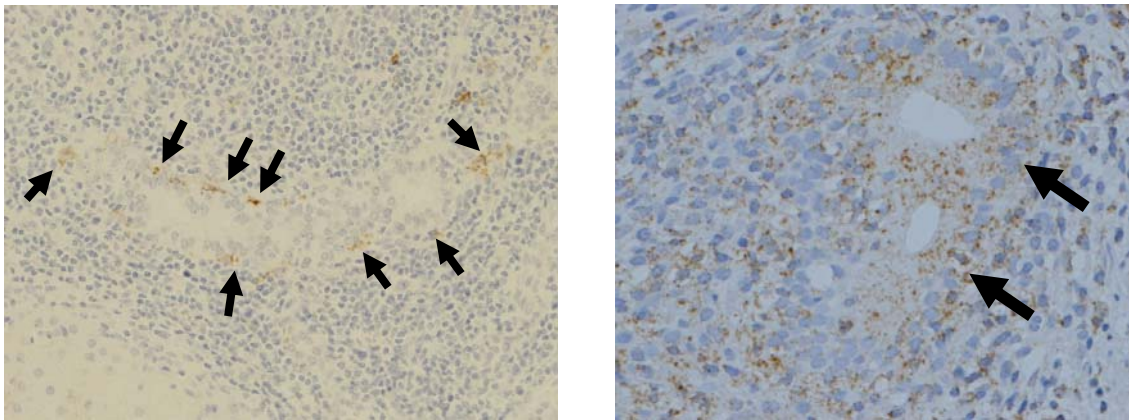
Fig.1 Immunohistochemistry for MCP-1 and α SMA. A and B; HCV-related chronic viral hepatitis. Proliferating bile ductules at the interface of periportal areas express MCP-1(A, arrows) and α SMA-positive cells showing hepatic stellate cell (HSC) morphology are found in the same area (B, arrows). C; primary biliary cirrhosis. MCP-1-positive proliferating bile ductules are found (arrow).



A

B

Fig.2 Immunohistochemistry for fractalkine. Primary biliary cirrhosis. Fractalkine is expressed in damaged bile duct (arrow in A) and periductal inflammatory cells in portal tracts and intrasinusoidal cells, probably Kupffer cells, in parenchyma. B is a higher magnification of A.



A

B

Fig.3. Detection of Langerhans cells (LCs) and macrophage inflammatory protein-3 α (MIP-3 α) in liver sections. (A) Langerin⁺ LCs are found around and within bile ducts showing cholangitis in primary biliary cirrhosis (PBC). (B) MIP-3 α is expressed in infiltrating mononuclear cells and the damaged bile ducts (arrows) in PBC.