

Pathologic significance of immunoglobulin G4-positive plasma cells in extrahepatic cholangiocarcinoma

メタデータ	言語: eng 出版者: 公開日: 2017-10-03 キーワード (Ja): キーワード (En): 作成者: メールアドレス: 所属:
URL	http://hdl.handle.net/2297/33485

Pathologic significance of immunoglobulin G4-positive plasma cells in extrahepatic cholangiocarcinoma

Yasushi Kimura, Kenichi Harada, and Yasuni Nakanuma

Department of Human Pathology, Kanazawa University Graduate School of medicine,
Kanazawa, Japan

Key words: IgG4, regulatory T cell, cholangiocarcinoma, cytotoxic T cell

Running title: IgG4 reaction in cholangiocarcinoma

Abbreviations: AIP, autoimmune pancreatitis; CTLs, cytotoxic T cells; EDTA, ethylenediaminetetraacetate; Foxp3, Forkhead box P3; H&E, Hematoxylin and eosin; HPFs, high-power fields; IgG4, Immunoglobulin G4; IELs, intraepithelial lymphocytes; Th2, T helper 2; Treg, regulatory T cells.

Address correspondence to:

Kenichi Harada, M.D.

Department of Human Pathology
Kanazawa University Graduate School of Medicine
Kanazawa 920-8640, Japan
FAX : (0)76-234-4229 (Japan)
TEL : (0)76-265-2199 (Japan)
E-mail : kenichih@med.kanazawa-u.ac.jp

SUMMARY

IgG4-related sclerosing cholangitis is histologically characterized by the infiltration of IgG4-positive plasma cells and sclerosing change. Moreover, several cases of carcinoma accompanied by IgG4-positive cells in tissue and increased serum IgG4 levels have been reported, but the association between cancer-associated immunity and an IgG4 reaction is still unclear. In this study, we examined the infiltration of IgG4-positive cells in extrahepatic cholangiocarcinoma and the pathological significance of the IgG4 reaction found in cancer tissues in terms of the evasion of immune surveillance by Treg cells. Immunohistochemistry for IgG4, Foxp3, CD4, and CD8 was performed using 68 surgical specimens from patients with extrahepatic cholangiocarcinoma and positive cells were investigated, particularly within and around cancerous tissues. Consequently, although IgG4⁺ cells were few (average, <10 cells/ high power field) in the majority of cases, ≥ 10 and ≥ 50 cells were found in 37% and 6% of cases, respectively. IgG4⁺ cells were predominantly found in the invasive front of carcinoma tissue. In the cases with ≥ 10 IgG4⁺ cells, Foxp3⁺ Treg cells were also distinguishable and a positive correlation was found between the Foxp3⁺/CD4⁺ ratio and IgG4⁺ cell count, but few CD8⁺ cells invaded cancer cells (<10 cells). In conclusion, extrahepatic cholangiocarcinomas are often accompanied by the significant infiltration of IgG4⁺ cells and the IgG4 reaction showed a positive and negative correlation with Foxp3⁺ and CD8⁺ cells, respectively, suggesting the evasion of immune surveillance associated with CD8⁺ cytotoxic T cells via the regulatory function of Foxp3⁺ Treg cells.

1. INTRODUCTION

Immunoglobulin G4 (IgG4) is a minor immunoglobulin subtype composing 3-6% of all the IgG circulating in adults [1], but important for the formation of IgG4-related diseases that feature elevated serum IgG4 levels and abundant infiltration with IgG4-positive plasma cells in affected organs [1-3]. IgG4-related diseases incorporate various IgG4-associated inflammatory disorders including autoimmune pancreatitis (AIP), sclerosing cholangitis, sialoadenitis, retroperitoneal fibrosis, inflammatory abdominal aortic aneurism, intestinal pneumonia, interstitial nephritis, lymphadenopathy, and inflammatory pseudotumor [3-11]. Recently, the number of cases of IgG4-related diseases has increased with the growing recognition of this disease entity, and clinicopathological characteristics for a differential diagnosis have been clarified. However, the pathological significance of increased serum IgG4 levels and marked infiltration of IgG4-positive plasma cells in target organs is still unknown.

IgG4-related diseases have varied clinical symptoms that may include features similar to malignant tumors. Because most IgG4-related diseases involving AIP are resolved by corticosteroid treatment, the diagnosis, particularly differentiation from malignant tumors, is very important [12-14]. Although using an upper normal limit for serum IgG4 of 135 mg/dL, Hamano et al. [1] reported a diagnostic 95% sensitivity and 97% specificity (vs pancreatic cancer) for AIP, pathological examination is necessary to differentiate IgG4-related diseases from tumors in any organs. Moreover, several investigators have recently reported on patients with pancreatic cancer accompanying elevated serum IgG4 levels and some cases are speculated to arise from AIP [15-17]. Raina et al. [18] reported that as many as 7% of patients with pancreatic cancer have serum IgG4 levels above 135mg/dL and concluded that in patients with pancreatic mass

lesions and suspicion of cancer, an IgG4 level measuring between 135 and 200mg/dL should be interpreted cautiously and not accepted as diagnostic of AIP without further evaluation. Some kind of association between tumor immunity and IgG4 reactions has been assumed, but detailed information is not available.

The participation of CD4+CD25+ Forkhead box P3 (Foxp3)+ regulatory T cells (Treg) and T helper 2 (Th2)-type helper T cells in the pathogenesis of the IgG4 reaction in IgG4-related diseases has been proposed [19]. Treg cells play a role in the progression of various malignant tumors, particularly in controlling the immune response against pancreatic ductal carcinoma from the premalignant stage to established cancer [20]. A high prevalence of Treg cells, moreover, seems to indicate a poor prognosis [20].

In this study, we retrospectively evaluated IgG4-positive plasma cells in extrahepatic cholangiocarcinomas including common bile duct cancers, gallbladder cancers, and cancers of the papilla of Vater and investigated the significance of the IgG4 reaction in cholangiocarcinoma from the point of view of tumor immune escape mediated by Treg cells.

2. MATERIALS and METHODS

2.1. Patients and tissue preparations

Formalin-fixed and paraffin-embedded sections of 68 surgically resected specimens from 39 patients with gallbladder cancer, 21 patients with common bile duct cancer, and 8 patients with cancer of the papilla of Vater (Average age 74 y.o, male/female=38/30) treated from 1998 to 2009 were obtained from the registry of liver diseases in the Department of Pathology, Kanazawa University School of Medicine. In

30 cases, follow-up data was also obtained in the present study. Each cholangiocarcinoma was classified histologically as well- (including papillary), moderately, or poorly differentiated, based on the predominant histologic grade. Special histological types such as adenosquamous carcinoma and mucinous carcinoma were not included in the present study. Four-micrometer-thick serial sections were prepared from each formalin-fixed, paraffin-embedded block. One was stained with Hematoxylin and eosin (H&E), and the others were used for immunohistochemistry.

2.2. Immunohistochemistry

The deparaffinized and rehydrated sections were microwaved in ethylenediaminetetraacetate (EDTA) buffer for IgG4 and Foxp3, in buffer at pH 9 for CD4, and in citrate buffer for CD8 and vimentin for 20 min in a microwave oven. Following the blocking of endogenous peroxidase, these sections were incubated at 4°C overnight with antibodies against IgG4 (mouse monoclonal; diluted 1:200; Southern Biotech, Birmingham, AL, USA), Foxp3 (mouse monoclonal; 10 µg/ml; Abcam, Cambridge, UK), CD4 (mouse monoclonal; neat, Nichirei, Tokyo, Japan), CD8 (mouse monoclonal; diluted 1:20, Dako, Tokyo, Japan), neutrophil elastase (mouse monoclonal, diluted 1:100, Dako), vimentin (mouse monoclonal, diluted 1:600, Dako), and CD34 (mouse monoclonal, diluted 1:200, Beckman Coulter, Tokyo, Japan) and then at room temperature for 1h with anti-mouse immunoglobulins conjugated to a peroxidase-labeled dextran polymer (Simple Staining Kit; Nichirei). After a benzidine reaction, sections were counterstained lightly with hematoxylin. No positive staining was obtained when the primary monoclonal antibody was replaced with an isotype-matched, non-immunized immunoglobulin as a negative control of the staining

procedures.

2.3. Histological examination

In addition to histological observation by H&E staining, the distribution of the immuno-positive cells was examined. In a primary survey, we examined all tumorous area in each specimen and, for counting IgG4+, Foxp3+, CD4+, or CD8+ mononuclear cells, selected three representative areas containing IgG4+ plasma cells, and expressed results as the mean number of each immuno-positive cell in high-power fields (HPFs). For semi-quantitative evaluation of the IgG4 staining, the cases with ≥ 10 and < 10 IgG4+ cells/HPF on average were evaluated as IgG4-positive and -negative cases, respectively. The ratio of Foxp3+ to CD4+ cells was calculated for the three selected area in each case and the average ratio (Foxp3/CD4) was compared between IgG4-rich and -poor cases, because the absolute number of Foxp3+ cells was prominently affected by the number of infiltrating mononuclear cells. Moreover, the average number of CD8+ cytotoxic T cells (CTLs) within carcinoma cells was evaluated to estimate the extent of the host immune response to the cancer. Finally, to confirm whether the IgG4 reaction is due to cancer-associated immunity, the area with IgG4-positive cells was evaluated in terms of neutrophilic infiltration, fibrosis and granulation. For neutrophilic infiltration, neutrophil elastase-positive cells were counted in the same area as IgG4-positive cells. For fibrosis and granulation, the immunoreactivity of vimentin and CD34, respectively, was used and the degree of change was semiquantitatively graded as follows: 0, absent; 1+, mild; 2+, intermediate; 3+, severe.

2.4. Statistical Analysis

Data were analyzed using Spearman's correlation coefficient test. Survival curves to evaluate the association between prognosis and IgG4 reactions were calculated by the Kaplan-Meier method, and analyses were conducted with the log-rank test. A p-value of less than .05 was considered to be statistically significant.

3. RESULTS

3.1 Detection and distribution of IgG4+ cells in extrahepatic cholangiocarcinoma

Immunohistochemistry revealed IgG4+ plasma cells to be scattered within and around cancerous nests in most cases (Fig.1). Particularly, around nests and in the invasive area facing a non-cancerous biliary wall and surrounding fibroadipose tissue, these positive cells were prominent with some intermingling of other inflammatory cells (Fig.2). Moreover, one characteristic feature of IgG4-related diseases, the perineural infiltration of IgG4+ cells, was commonly seen in extrahepatic cholangiocarcinomas. In contrast, desmoplastic change and vascular invasion by cancer cells are usually seen in extrahepatic cholangiocarcinomas, but other features of IgG4-related diseases, obliterative phlebitis caused by IgG4+ cells and storiform-type fibrosis, are rare.

A quantitative evaluation of IgG4+ cells revealed that 25 (37%), 19 (28%), and 4 (6%) of 68 cholangiocarcinoma patients had ≥ 10 , ≥ 20 , and ≥ 50 IgG4+ cells/HPF, respectively. There was no correlation between the density of IgG4+ cells and any clinicopathological factor including age, gender, anatomical location (common bile ducts, gallbladder, and the papilla of Vater), or the histological differentiation (well-, moderately, and poorly) of extrahepatic cholangiocarcinomas.

3.2. Association between IgG4 reactions and Foxp3+ Treg cells

The association between IgG4⁺ and Foxp3⁺ cells was evaluated in each case. Foxp3⁺ Treg cells were scattered in most cases with a marked IgG4 reaction (Fig.3). The relation between the IgG4 reaction and Foxp3⁺ Treg cells is shown in Fig.4. In IgG4-rich cases (≥ 10 IgG4⁺ cells/HPF), the ratio of Foxp3⁺/CD4⁺ cells correlated closely with the IgG4⁺ cell count, though in IgG4-poor cases (< 10 IgG4⁺ cells/HPF), the correlation varied.

3.3. Association between IgG4 reactions and CD8⁺ CTLs

CD8⁺ CTLs were scattered to various degree in each case irrespective of whether they were within or around cancer nests. As a marker of immune activity against cancers, CTLs invaded cancerous nests resembling intraepithelial lymphocytes (IELs), which are found in non-neoplastic biliary epithelial layers of biliary diseases such as primary biliary cirrhosis (Fig.5)[21] Consequently, patients with many CD8⁺ CTLs showed scant IgG4 reactions (IgG4-poor cases) and all IgG4-rich cases had few CD8⁺ CTLs (< 10 cells/HPF) (Fig.6)

3.4. Histological conditions for IgG4 reactions

As shown in Fig.7, there was no correlation between the numbers of neutrophil elastase-positive neutrophils and IgG4-positive cells. Moreover, the degree of fibrosis and granulation did not correlate with the IgG4-positive cell count either.

3.5. Association between IgG4 reactions and patients' survival

After the surgical resection of extrahepatic cholangiocarcinomas, of the 30 patients with available outcome data, 21 died from recurrence of the cancer. The overall

survival curve for the 30 patients, obtained using the Kaplan-Meier estimator is shown in Fig.7. The patients with <20 IgG4+ cells/HPF had a better prognosis than those with ≥ 20 cells ($p < 0.05$).

4. DISCUSSION

Elevated serum IgG4 levels and the infiltration of organs by numerous IgG4+ plasma cells are clinicopathological hallmarks of IgG4-related diseases. Moreover, obliterative phlebitis, storiform-type sclerosing fibrosis, and sometimes mass forming-type sclerosing fibrosis, are also characteristic of this disease category. It is clinically and pathologically important to distinguish IgG4-related diseases from tumors of affected organs. In particular, because desmoplastic change is a common feature of biliary and pancreatic cancers, IgG4-related diseases and cancers in these organs show similar radiological behaviors. Moreover, patients with pancreatic adenocarcinoma accompanying an IgG4 reaction and/or elevated serum IgG4 levels [16,18,22,23] and with pancreatic and biliary cancers arising from IgG4-related disease [16,24,25] have been reported, though a cause-and-effect relationship between the IgG4 reaction and cancers has not been demonstrated. Therefore, the presence of IgG4+ cells is not a histological hallmark of IgG4-related diseases and the IgG4 reaction is speculated to occur non-specifically in carcinoma tissues [26]. In this study, we retrospectively examined IgG4 reactions in cases of extrahepatic cholangiocarcinoma including common bile duct cancers, gallbladder cancer, and cancers of the papilla of Vater. Consequently, 10 or more IgG4+ cells/HPF were observed in 37% of cases and the cases with marked infiltration (≥ 50 IgG4-positive cells/HPF) and resembling IgG4-related diseases made up 6% of the total. As the pathological diagnostic criteria of

IgG4-related disease, the essential number of IgG4+ cells varied from 5 to 50/HPF depending on the affected organs, but in IgG4-related sclerosing cholangitis, ≥ 10 IgG4+ cells/HPF is proposed according to the HISORT criteria published for AIP [27,28]. In our cases, therefore, several cancers accompanying remarkable infiltration of IgG4-positive cells (≥ 50 cells/HPF) were speculated to originate from the preceding IgG4-related disease, but follow-up data prior to discovery of the cancers is needed to demonstrate this. Because cases of pancreatic and biliary cancers arising from IgG4-related disease were reported [16,24,25], IgG4-related sclerosing cholangitis is thought to be an important preceding disease in the carcinogenesis of cholangiocarcinoma. Irrespective of whether the patients have cholangiocarcinoma and a marked IgG4 reaction or cancers arising from IgG4-related sclerosing cholangitis, the presence of adenocarcinoma should be taken into account in the pathological diagnosis of IgG4-related cholangitis, particularly using small specimens such as biopsy materials.

The present study demonstrated that a IgG4 reaction is often found to some degree in extrahepatic cholangiocarcinoma as well as IgG4-related diseases. Moreover, the perineural infiltration of IgG4+ cells which is a feature of IgG4-related cholangitis and AIP, was prominent in IgG4-rich cholangiocarcinoma cases. In contrast, patients with IgG4-related cholangitis are generally older men (85%) [27], but this male domination was not found in the IgG4 reaction of cholangiocarcinoma. Obliterative phlebitis and storiform fibrosis are also characteristic of IgG4-related diseases, but rare in cholangiocarcinoma. These histological features of IgG4-related diseases differing from cholangiocarcinoma are not located at the superficial biliary mucosa, unfortunately suggesting that these characteristic findings are not useful for a differential diagnosis

using biopsy specimens. The IgG4 reaction is not a specific immunereaction of IgG4-related diseases, but the immunopathogenesis of IgG4 reactions should be different in IgG4-related diseases and cancers. As shown in Fig.7, no correlation between the number of IgG4-positive cells and degree of neutrophilic infiltration, fibrosis or granulation was found, suggesting that the IgG4 reaction in cholangiocarcinoma is due to cancer-associated immunity via the different mechanisms of IgG4-related diseases characterized by fibrosis as well as IgG4 reactions. Further study is necessary to clarify the histogenesis of IgG4 reactions in cholangiocarcinoma.

IgG4 does not have the ability to activate complement and its physiological and pathological significance are still unknown in healthy and IgG4-related diseased patients. As the pathogenesis of the IgG4 reaction in IgG4-related sclerosing cholangitis and pancreatitis, the participation of the Th2-type cytokine milieu and IL-10 produced by Treg cells is assumed to involve IgG4 class switching and/or the progressive proliferation/differentiation of IgG4+ plasma cells [10,19,29-31]. In the carcinogenesis of pancreatic cancer, the prevalence of Treg cells increases and that of cytotoxic CD8+ cells conversely diminishes in cancer tissues [20]. Moreover, the prevalence of Treg cells is negatively correlated with the prognosis of patients with pancreatic cancers [20]. These findings suggest that Treg cells play a role in controlling the immune response against pancreatic cancer, especially the evasion of tumor-associated immune surveillance. The present study using extrahepatic cholangiocarcinomas also demonstrated that in the IgG4-rich cases (≥ 10 IgG4+ cells/HPF), the number of IgG4+ cells in cancer tissue positively correlated with that of Foxp3+ Treg cells and conversely the number of cytotoxic CD8+ CTLs was constantly small. Therefore, extrahepatic cholangiocarcinomas as well as pancreatic cancers could cause the evasion of immune

surveillance via the regulatory function of Treg cells, involving the concomitant IgG4 reaction. However, high Foxp3⁺/CD4⁺ cells ratio were also seen in many IgG4-poor cases, suggesting that the presence of Foxp3⁺ cells is not sufficient for the induction of IgG4 reaction. A functional analysis of infiltrating Foxp3⁺ Treg cells and possible other mechanisms of IgG4 reactions in cholangiocarcinoma also should be considered.

Finally, we examined the prognosis. Clinical follow-up data was available for only 30 of the 68 patients, because of the retirement or transfer of primary doctors, a cessation of digestive surgery at the affiliated hospitals, etc. Moreover, as more than 10 years had passed since the operation in several cases, sometimes the clinical records themselves had been destroyed. In addition to the analysis using cytotoxic CD8⁺ CTLs as mentioned above, survival curves obtained using the follow-up data for the 30 patients also indicated the patients with <20 IgG4-positive cells/HPF to have a better prognosis than those with ≥ 20 cells. Although the role of IgG4⁺ plasma cells in cancer tissue is unclear, the degree of IgG4⁺ cell infiltration might be a pathological marker of extrahepatic cholangiocarcinoma.

In conclusion, this study revealed that extrahepatic cholangiocarcinoma often accompanies significant infiltration by IgG4⁺ cells, indicating a need to distinguish it from IgG4-related diseases, especially when using small specimens such as biopsy materials. Moreover, the IgG4 reaction in cholangiocarcinoma might be associated with the evasion of immune surveillance by CD8⁺ CTLs and tumor progression through Treg cells.

5. ACKNOWLEDGEMENTS

The authors thank Dr. Itoh (Nagasaki Medical Center), Dr. Watanabe

(Watanabe's Consultancy for Pathological Diagnosis), Dr. Shinagawa (Wajima Municipal hospital), Dr. Takigawa (Ushitsu general hospital), Dr. Takabatake (Kanazawa Arimatsu hospital), Dr. Kawanishi (Yahata Medical Center), and Dr. Nakamura (Nakamura Hospital) for generously providing clinical information. This work was supported by grant No.23590393 from the Ministry of Education, Culture, Sports, Science and Technology of Japan (K.H.) and by Health and Labour Sciences Research Grants for Research on Measures for Intractable Diseases.

REFERENCE

1. Hamano H, Kawa S, Horiuchi A, Unno H, Furuya N, Akamatsu T, Fukushima M, et al. High serum IgG4 concentrations in patients with sclerosing pancreatitis. *N Engl J Med* 2001; 344: 732-738.
2. Hamano H, Kawa S, Ochi Y, Unno H, Shiba N, Wajiki M, Nakazawa K, et al. Hydronephrosis associated with retroperitoneal fibrosis and sclerosing pancreatitis. *Lancet* 2002; 359: 1403-1404.
3. Zen Y, Nakanuma Y. IgG4-related disease: a cross-sectional study of 114 cases. *Am J Surg Pathol* 2010; 34: 1812-1819.
4. Kamisawa T, Nakajima H, Egawa N, Funata N, Tsuruta K, Okamoto A. IgG4-related sclerosing disease incorporating sclerosing pancreatitis, cholangitis, sialadenitis and retroperitoneal fibrosis with lymphadenopathy. *Pancreatology* 2006; 6: 132-137.
5. Kitagawa S, Zen Y, Harada K, Sasaki M, Sato Y, Minato H, Watanabe K, et al. Abundant IgG4-positive plasma cell infiltration characterizes chronic sclerosing sialadenitis (Kuttner's tumor). *Am J Surg Pathol* 2005; 29: 783-791.
6. Kasashima S, Zen Y, Kawashima A, Konishi K, Sasaki H, Endo M, Matsumoto Y, et al. Inflammatory abdominal aortic aneurysm: close relationship to IgG4-related periaortitis. *Am J Surg Pathol* 2008; 32: 197-204.
7. Saeki T, Saito A, Yamazaki H, Emura I, Imai N, Ueno M, Nishi S, et al. Tubulointerstitial nephritis associated with IgG4-related systemic disease. *Clin Exp Nephrol* 2007; 11: 168-173.
8. Zen Y, Inoue D, Kitao A, Onodera M, Abo H, Miyayama S, Gabata T, et al. IgG4-related lung and pleural disease: a clinicopathologic study of 21 cases. *Am J Surg Pathol* 2009; 33: 1886-1893.
9. Zen Y, Onodera M, Inoue D, Kitao A, Matsui O, Nohara T, Namiki M, et al. Retroperitoneal fibrosis: a clinicopathologic study with respect to immunoglobulin G4. *Am J Surg Pathol* 2009; 33: 1833-1839.
10. Okazaki K, Uchida K, Fukui T. Recent advances in autoimmune pancreatitis: concept, diagnosis, and pathogenesis. *J Gastroenterol* 2008; 43: 409-418.
11. Nakanuma Y, Zen Y. Pathology and immunopathology of immunoglobulin G4-related sclerosing cholangitis: The latest addition to the sclerosing cholangitis family. *Hepatol Res* 2007; 37 Suppl 3: S478-486.
12. Okazaki K, Uchida K, Matsushita M, Takaoka M. Autoimmune pancreatitis. *Intern Med* 2005; 44: 1215-1223.
13. Kamisawa T, Okamoto A. Autoimmune pancreatitis: proposal of IgG4-related sclerosing disease. *J Gastroenterol* 2006; 41: 613-625.
14. Okazaki K, Chiba T. Autoimmune related pancreatitis. *Gut* 2002; 51: 1-4.
15. Inoue H, Miyatani H, Sawada Y, Yoshida Y. A case of pancreas cancer with autoimmune pancreatitis. *Pancreas* 2006; 33: 208-209.
16. Kamisawa T, Chen PY, Tu Y, Nakajima H, Egawa N, Tsuruta K, Okamoto A, et al. Pancreatic cancer

- with a high serum IgG4 concentration. *World J Gastroenterol* 2006; 12: 6225-6228.
17. Sakashita F, Tanahashi T, Yamaguchi K. Case of pancreatic tail cancer associated with auto-immune pancreatitis. *Jpn J Gastroenterol* 2006; 39: 78-83.
 18. Raina A, Krasinskas AM, Greer JB, Lamb J, Fink E, Moser AJ, Zeh HJ, 3rd, et al. Serum immunoglobulin G fraction 4 levels in pancreatic cancer: elevations not associated with autoimmune pancreatitis. *Arch Pathol Lab Med* 2008; 132: 48-53.
 19. Zen Y, Fujii T, Harada K, Kawano M, Yamada K, Takahira M, Nakanuma Y. Th2 and regulatory immune reactions are increased in immunoglobulin G4-related sclerosing pancreatitis and cholangitis. *Hepatology* 2007; 45: 1538-1546.
 20. Hiraoka N, Onozato K, Kosuge T, Hirohashi S. Prevalence of FOXP3+ regulatory T cells increases during the progression of pancreatic ductal adenocarcinoma and its premalignant lesions. *Clin Cancer Res* 2006; 12: 5423-5434.
 21. Isse K, Harada K, Sato Y, Nakanuma Y. Characterization of biliary intra-epithelial lymphocytes at different anatomical levels of intrahepatic bile ducts under normal and pathological conditions: numbers of CD4+CD28- intra-epithelial lymphocytes are increased in primary biliary cirrhosis. *Pathol Int* 2006; 56: 17-24.
 22. Ghazale A, Chari ST, Smyrk TC, Levy MJ, Topazian MD, Takahashi N, Clain JE, et al. Value of serum IgG4 in the diagnosis of autoimmune pancreatitis and in distinguishing it from pancreatic cancer. *Am J Gastroenterol* 2007; 102: 1646-1653.
 23. Dhall D, Suriawinata AA, Tang LH, Shia J, Klimstra DS. Use of immunohistochemistry for IgG4 in the distinction of autoimmune pancreatitis from peritumoral pancreatitis. *Hum Pathol* 2010; 41: 643-652.
 24. Motosugi U, Ichikawa T, Yamaguchi H, Nakazawa T, Katoh R, Itakura J, Fujii H, et al. Small invasive ductal adenocarcinoma of the pancreas associated with lymphoplasmacytic sclerosing pancreatitis. *Pathol Int* 2009; 59: 744-747.
 25. Oh HC, Kim JG, Kim JW, Lee KS, Kim MK, Chi KC, Kim YS, et al. Early bile duct cancer in a background of sclerosing cholangitis and autoimmune pancreatitis. *Intern Med* 2008; 47: 2025-2028.
 26. Harada K, Shimoda S, Kimura Y, Sato Y, Ikeda H, Igarashi S, Ren XS, et al. Significance of IgG4-positive cells in extrahepatic cholangiocarcinoma: Molecular mechanism of IgG4 reaction in cancer tissue. *Hepatology* 2012 (in press).
 27. Ghazale A, Chari ST, Zhang L, Smyrk TC, Takahashi N, Levy MJ, Topazian MD, et al. Immunoglobulin G4-associated cholangitis: clinical profile and response to therapy. *Gastroenterology* 2008; 134: 706-715.
 28. Chari ST, Smyrk TC, Levy MJ, Topazian MD, Takahashi N, Zhang L, Clain JE, et al. Diagnosis of autoimmune pancreatitis: the Mayo Clinic experience. *Clin Gastroenterol Hepatol* 2006; 4: 1010-1016; quiz 1934.

29. Jeannin P, Lecoanet S, Delneste Y, Gauchat JF, Bonnefoy JY. IgE versus IgG4 production can be differentially regulated by IL-10. *J Immunol* 1998; 160: 3555-3561.
30. Nouri-Aria KT, Wachholz PA, Francis JN, Jacobson MR, Walker SM, Wilcock LK, Staple SQ, et al. Grass pollen immunotherapy induces mucosal and peripheral IL-10 responses and blocking IgG activity. *J Immunol* 2004; 172: 3252-3259.
31. Koyabu M, Uchida K, Miyoshi H, Sakaguchi Y, Fukui T, Ikeda H, Takaoka M, et al. Analysis of regulatory T cells and IgG4-positive plasma cells among patients of IgG4-related sclerosing cholangitis and autoimmune liver diseases. *J Gastroenterol* 2010; 45: 732-741.

FIGURES and LEGENDS

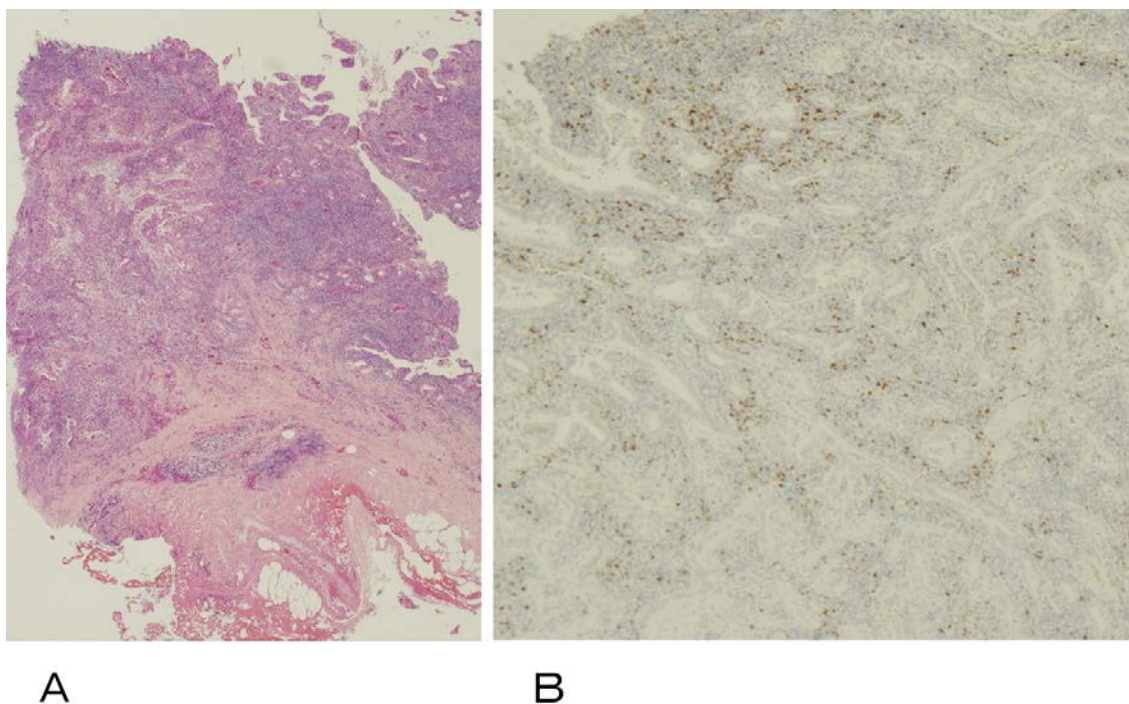


Fig.1

Fig.1 Common bile duct cancer. A: Papillary adenocarcinoma. Inflammatory cells are prominent. H&E staining. Original magnification, x20. B: Immunohistochemistry for IgG4. Numerous IgG4+ cells are present in the inflamed stroma. Original magnification, x100.

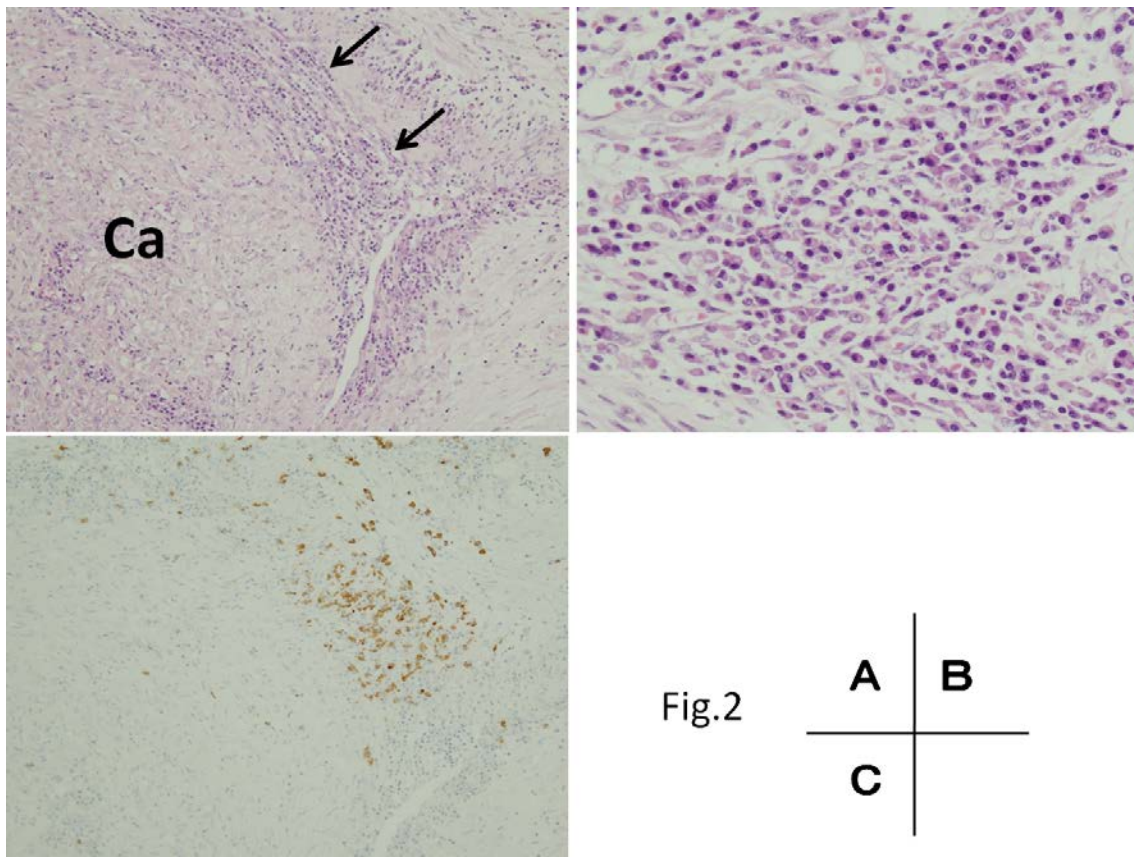


Fig.2 Common bile duct cancer. A: A marginal zone (arrows) of poorly differentiated adenocarcinoma (Ca). Many inflammatory cells are present. H&E staining. Original magnification, x100. B: Higher magnification of the marginal zone. Inflammatory cells are mostly composed of plasma cells. H&E staining. Original magnification, x400. C: Immunohistochemistry for IgG4. Many positive cells are scattered in the marginal zone. Original magnification, x100

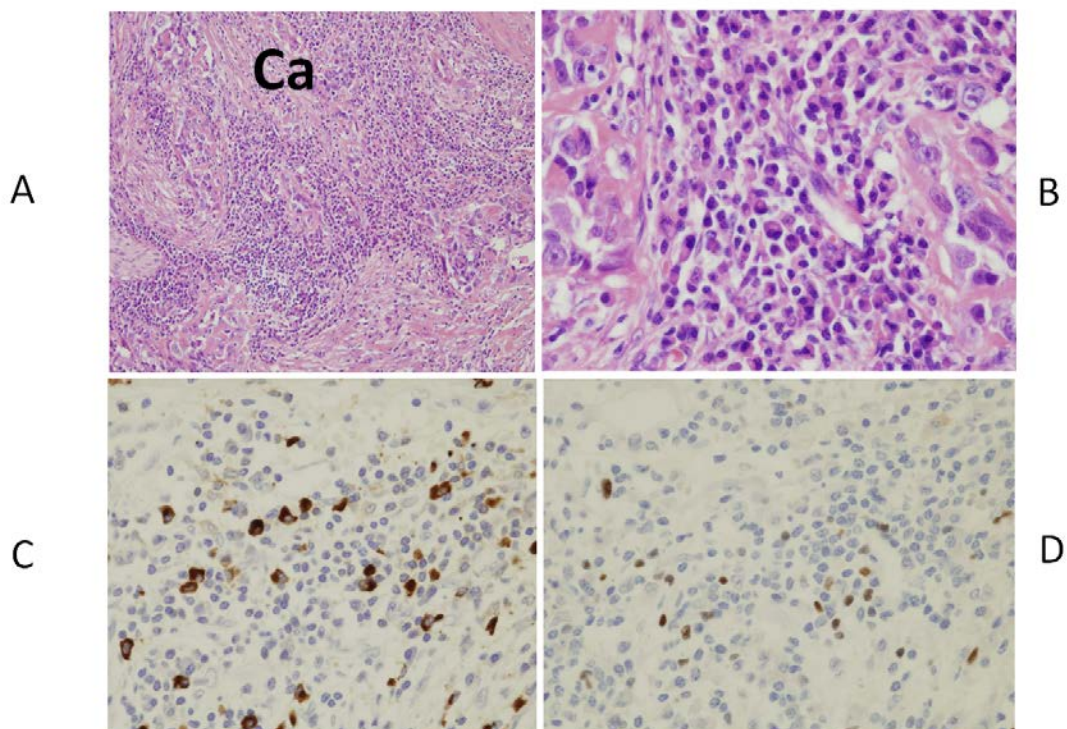


Fig.3

Fig.3 Gallbladder cancer. A: An invasive area of poorly differentiated adenocarcinoma (Ca). Numerous inflammatory cells are found. H&E staining. Original magnification, x100. B: Higher magnification of the invasive area. Many plasma cells are evident. H&E staining. Original magnification, x400. C: Immunohistochemistry for IgG4. Several IgG4-positive cells have accumulated. Original magnification, x400. D: Immunohistochemistry for Foxp3. Positive cells are scattered and slightly accumulated here and there. Original magnification, x400.

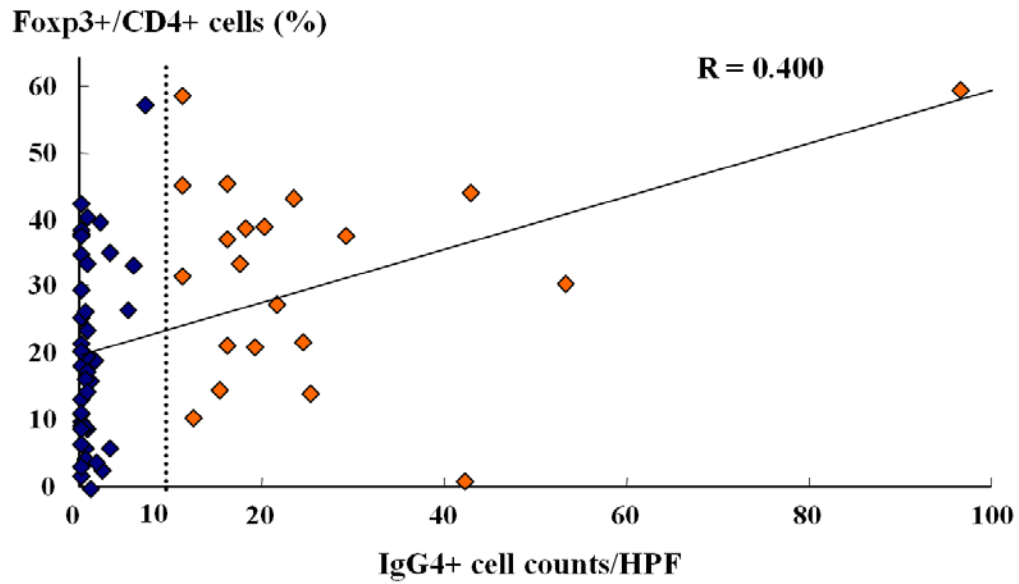


Fig.4

Fig.4 Relationship between IgG4+ cells and Treg cells in extrahepatic cholangiocarcinoma. In cases with ≥ 10 /HPF IgG4+ cells (IgG4-rich cases), there is a good correlation between IgG4+ and the ratio of Fosp3+/CD4+ cells (R = 0.400).

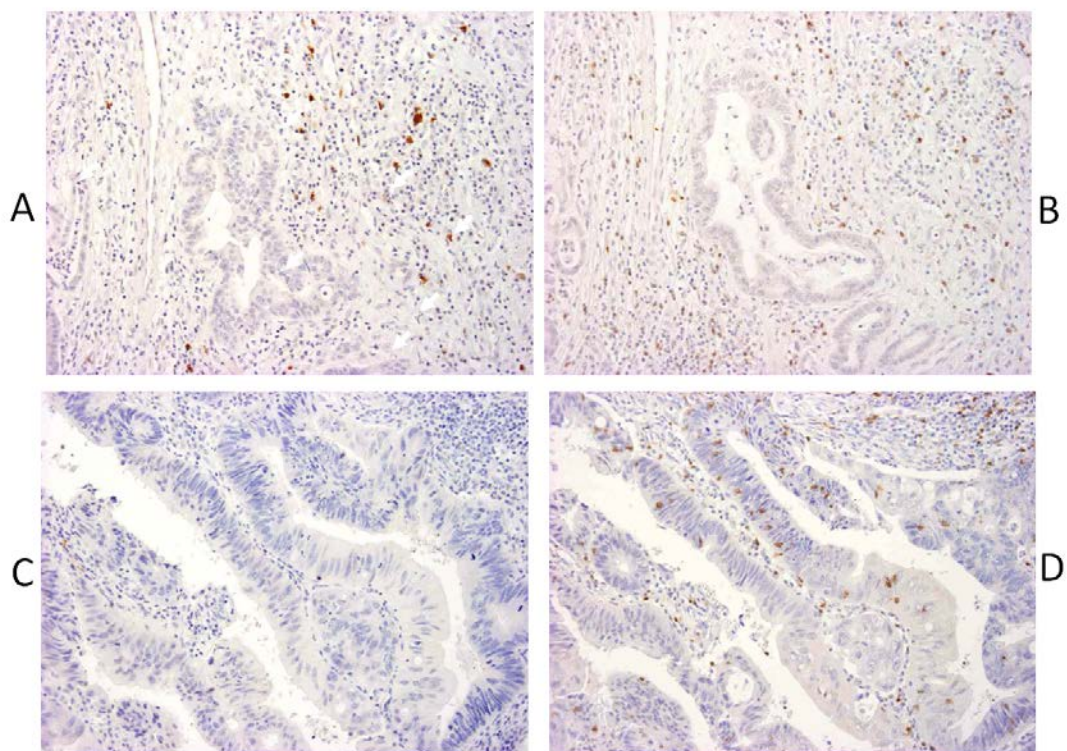


Fig.5

Fig.5 Comparison of CD8+ cytotoxic T cells in IgG4-rich (A and B) and -poor (C and D) cases of extrahepatic cholangiocarcinoma. Immunohistochemistry for IgG4 (A and C) and CD8 (B and D). In IgG4-rich cases, CD8+ as well as IgG4+ cells are found around adenocarcinoma tissue, but no CD8+ cells are present in cancer cells lining the adenocarcinoma. In contrast, many CD8+ cells invade cancer cells lining the adenocarcinoma in IgG4-poor cases. Original magnification, x200.

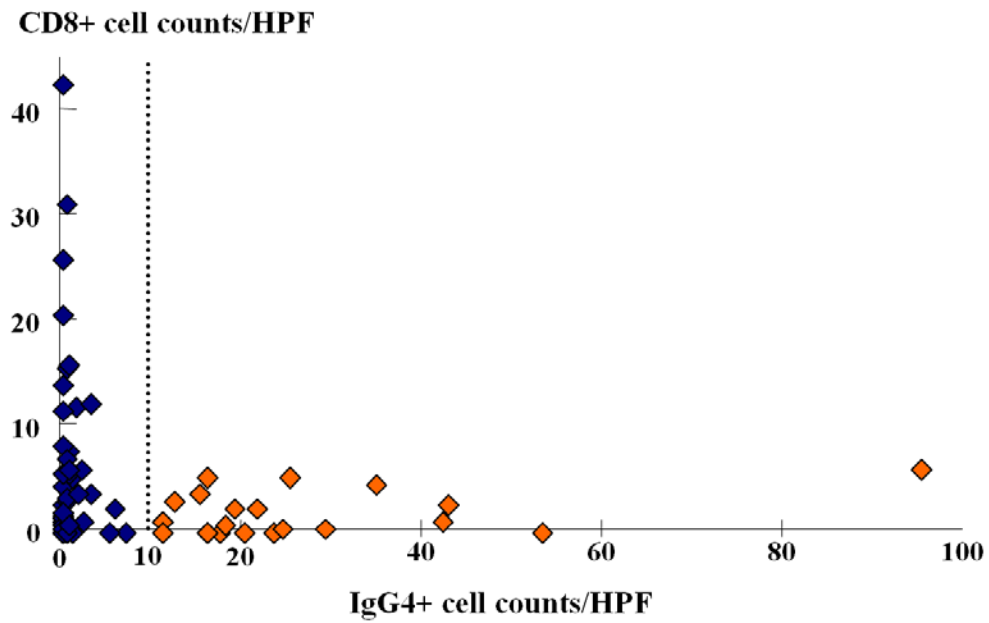


Fig.6

Fig.6 Relationship between IgG4+ and CD8+ cytotoxic T cells in extrahepatic cholangiocarcinoma. In all cases with ≥ 10 /HPF IgG4+ cells (IgG4-rich cases), the number of CD8+ cells is less than 10.

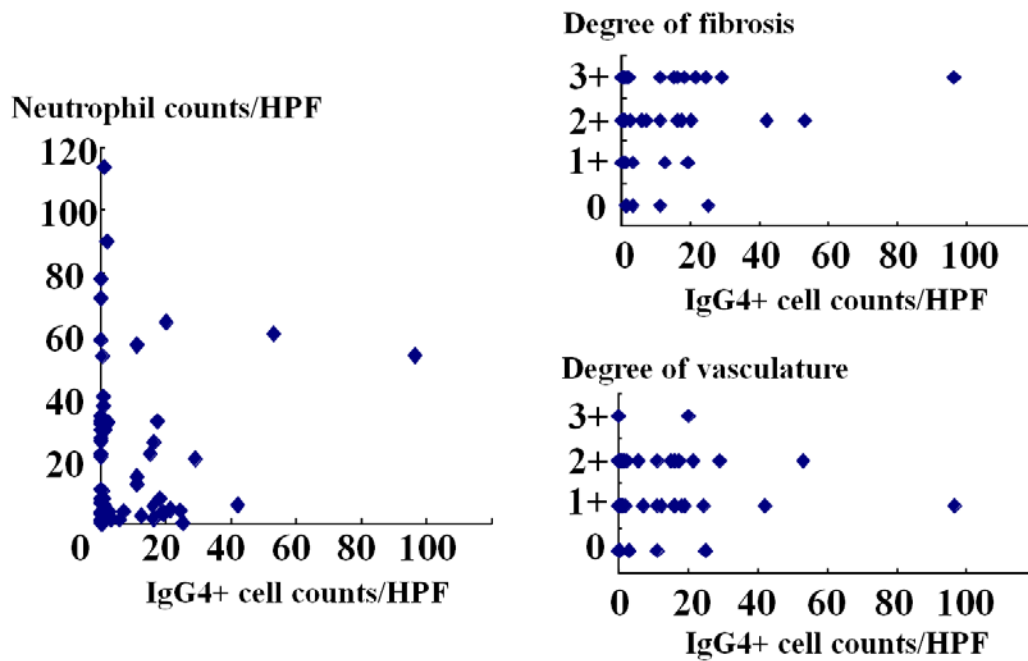


Fig.7

Fig.7 Correlation of IgG4 reactions with neutrophil infiltration, fibrosis, and granulation in cholangiocarcinoma. The degree of neutrophil infiltration was evaluated from the neutrophil elastase-positive cell counts. The degree of fibrosis and granulation was semiquantitatively graded as follows: 0, absent; 1+, mild; 2+, intermediate; 3+, severe, based on the vimentin-positive area and CD34-positive neovasculature, respectively. Consequently, no correlation was found between the degree of neutrophil infiltration, fibrosis, or granulation and IgG4-positive cells.

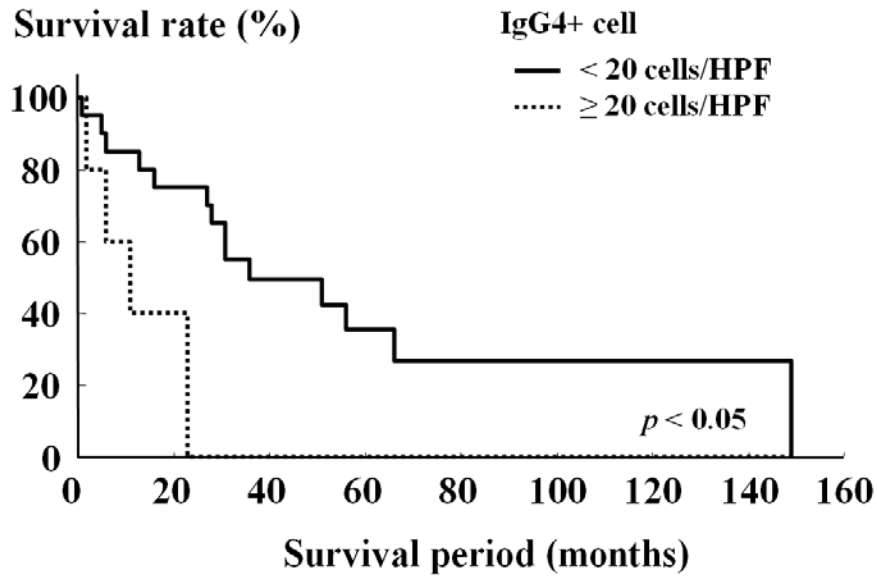


Fig.8

Fig.8 Survival curve of patients with extrahepatic cholangiocarcinoma. Kaplan-Meier plots of the postoperative survival period were made for two groups of patients, those with <20 and ≥ 20 IgG4+positive cells/HPF. Log-rank analysis of the postoperative survival periods indicated that the patients with <20 IgG4+ cells/HPF had a better prognosis ($p < 0.05$).