

Tetraspanin CD63 Promotes Targetion and Lysosomal Proteolysis of Membrane-Type 1 Matrix Metalloproteinase

メタデータ	言語: eng 出版者: 公開日: 2017-10-05 キーワード (Ja): キーワード (En): 作成者: メールアドレス: 所属:
URL	https://doi.org/10.24517/00027355

This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 International License.



Tetraspanin CD63 Promotes Targeting and Lysosomal Proteolysis of Membrane-Type 1 Matrix Metalloproteinase

Takahisa Takino*, Hisashi Miyamori*, Noriko Kawaguchi*, Takamasa Uekita†

Motoharu Seiki† and Hiroshi Sato*‡

*Department of Molecular Virology and Oncology, Cancer Research Institute,

Kanazawa University, 13-1 Takara-machi, Kanazawa 920-0934, Japan

†Division of Cancer Cell Research, Institute of Medical Science, University of Tokyo, 4-6-1

Shirokanedai, Minato-ku, Tokyo 108-8639, Japan

‡To whom correspondence should be addressed:

Department of Molecular Virology and Oncology, Cancer Research Institute,

Kanazawa University, 13-1 Takara-machi, Kanazawa 920-0934, Japan

Tel: +81-76-265-2749

Fax: +81-76-234-4505

E-mail: yhsato@kenroku.kanazawa-u.ac.jp

17 text pages, 21 references, 4 figures

Number of words: 2,440

SUMMARY

Membrane-type 1 matrix metalloproteinase (MT1-MMP) is known to be internalized from cell surface, however, the fate of internalized MT1-MMP is still unknown. Here we demonstrate that at least a part of internalized MT1-MMP is targeted for lysosomal proteolysis. Treatment with an inhibitor of lysosomal proteinases chloroquine suppressed degradation of internalized MT1-MMP, and induced accumulation of MT1-MMP in CD63-positive lysosomes. Ectopic expression of CD63 accelerated degradation of MT1-MMP, which was blocked by chloroquine. MT1-MMP and CD63 were shown to form a complex through hemopexin-like domain of MT1-MMP and N-terminal region of CD63, and thus accelerated degradation of MT1-MMP was not observed with mutants lacking these domains. CD63 mutant lacking lysosomal targeting motif was unable to promote MT1-MMP degradation. These results suggest that CD63 regulates MT1-MMP by targeting to lysosomes.

Keywords: CD63; internalization; lysosomes; MMP; MT1-MMP; proteolysis; tetraspanins

Matrix metalloproteinases (MMP) are a family of Zn^{2+} -dependent enzymes that are essential for extracellular matrix (ECM) turnover in normal and pathological conditions. MMP anchored to plasma membrane are subgrouped into membrane-type MMP (MT-MMP) [1-5]. Of all the MT-MMPs, MT1-MMP is believed to be a crucial for the invasion of malignant tumors [1, 5, 6]. MT1-MMP activity is regulated by several distinct mechanisms. To express on cell surface, proMT1-MMP requires for being processed by proprotein convertases [7]. The enzymatic activity on the cell surface can be inhibited by the tissue inhibitors of metalloproteinases (TIMP)-2, -3, and -4 but not by TIMP-1. After MMP-2 activation, a part of MT1-MMP is down-regulated through processing by activated MMP-2 and autocatalytically [8]. Recent studies have revealed that the cell-surface MT1-MMP is regulated by internalization [9, 10]. However, it is largely unknown whether MT1-MMP is degraded at endosomes/lysosomes, or whether internalized MT1-MMP is recycled back to the plasma membrane.

Tetraspanins of integral membrane proteins with four predicted transmembrane domains comprises a large group of ubiquitously expressed proteins that function in diverse contexts such as T- and B-cell activation, and cell migration and proliferation [11]. CD63 is a member of tetraspanins and a well-established component of the late endosomal and lysosomal membranes [12]. CD63 is highly expressed in primary melanoma but weaker or absent in metastatic melanomas [13, 14]. Ectopic expression of CD63 in melanoma cells suppresses their metastatic

potential in animal model system [15] . In the present study, we analyzed the fate of internalized MT1-MMP and the effect of CD63 on the stability of MT1-MMP and demonstrated that CD63 plays at least in part a role in promoting lysosomal proteolysis of MT1-MMP.

MATERIALS AND METHODS

Cell Culture and Materials. MDCK cells and HT1080 cells were maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 5% fetal bovine serum (FBS). HeLa cells and 293-EBNA cells (Invitrogen) were cultured in DMEM supplemented with 10% FBS. MMP inhibitor, BB94 was prepared as described previously [16]. Antibodies used were: anti-actin and anti-FLAG M2 mouse monoclonal antibodies (Sigma-Aldrich); an anti-GFP rabbit polyclonal antibody (Santa Cruz Biotechnology); anti-CD63 mouse monoclonal and anti-MT1-MMP rabbit polyclonal antibodies (Chemicon); horseradish peroxidase (HRP) conjugated secondary antibodies and streptavidin (Amersham Pharmacia Biothech).

Expression Plasmids. pSG5-MMP-2, FLAG epitope tagged-MT1-MMP (MT1F), the catalytic domain-deleted mutant (Δ Cat), the hemopexin-like domain-deleted mutant (Δ Pex) and the cytoplasmic tail-deleted mutant (Δ CP) of MT1F were constructed as described previously [10]. pRK-GFP was provided from Dr. K.M. Yamada (National Institutes of Health, Bethesda, MD). The puromycin resistance plasmid pHA262pur was kindly provided by Dr. Hein te Riele (Netherlands

Cancer Institute, Amsterdam, The Netherlands). A cDNA encoding CD63 (GenBank accession number BC002349) was obtained by RT-PCR with human placenta cDNA and was cloned into the pRK-GFP expression vector. pRK-GFP-CD63 was used as a template for PCR-based modification to generate the deletion mutants for the small extracellular loop (Met¹-Ser⁵¹)-deleted mutant (Δ SEL), the large extracellular loop (Val¹⁰⁶-Lys¹⁸⁶)-deleted mutant (Δ LEL), the cytoplasmic domain-deleted mutant (Δ C12; Leu²²⁷-Met²³⁸) of GFP-CD63.

Transfection. Transient transfections were performed by either calcium phosphate methods or using Trans IT-LT1® (PanVera) according to the manufacture's instructions. To obtain MDCK cells stably expressing GFP-CD63, MDCK cells were co-transfected with pRK-GFP-CD63 and pHA262pur plasmids and selected in medium containing puromycin.

Immunoprecipitations and Immunoblottings. The cells were washed with ice-cold phosphate buffered saline (PBS), homogenized in a buffer containing 50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 1 mM EGTA, 1 mM PMSF, 1 mM Na₃VO₄, 1 mM NaF, 1% TritonX-100. The cell lysates were used for immunoprecipitation with anti-FLAG-M2 antibody. The immunoprecipitates were subjected to immunoblotting using indicated antibodies.

Gelatin Zymography. Gelatin zymography was performed with SDS-polyacrylamide gel containing 0.1% gelatin as described previously [3]. Gelatinolytic activity was visualized as a negative staining with Coomassie Brilliant Blue.

Cell-Surface Biotinylation. Transfected HeLa cells were washed with ice-cold PBS, incubated with 0.5 mg/ml sulfo-NHS-biotin (Pierce) in PBS at 4°C for 30 min, and then further incubated with 25 mM lysine. The cells were cultured for 3 h in complete medium with or without chloroquine (116 µM), homogenized and immunoprecipitated with anti-FLAG M2 antibody. The immunoprecipitates were analyzed by immunoblotting using streptavidine-HRP or anti-FLAG antibody.

Immunofluorescence Staining. For cell-surface staining, cells were incubated for 30 min at 4°C with anti-FLAG M2 antibody. After washing, cells were incubated in complete media with BB94 (1 µM) at 37°C for indicated period. The cells were fixed with 4% paraformaldehyde for 15 min and permeabilized with (permeabilized) or without (non-permeabilized) 4% paraformaldehyde/0.5% Triton X-100 for 5min, and then visualized with Cy3- or AlexaTM488-conjugated goat anti-mouse IgG (Molecular Probes) using confocal laser microscopy (Carl Zeiss). For CD63 staining, cells were fixed with 4% paraformaldehyde for 20 min, treated for 30 min with 0.27% NH₄Cl/0.38% glycine in PBS, and permeabilized for 30 min with 0.05% saponin in PBS.

RESULTS

Internalized MT1-MMP is localized at CD63-positive lysosomes. Internalization of

FLAG-tagged MT1-MMP (MT1F) in HeLa cells was examined by fluorescence immunostaining. MT1F was localized at the periphery of permeabilized and non-permeabilized cells at 0 time (Fig. 1A). While the signal became weaker at 30 min and undetectable at 3 h after labeling in non-permeabilized cells, MT1F was detected in cytoplasm as dot-like spots at 3 h after labeling in permeabilized cells. Next, to examine whether MT1F is subjected to lysosomal proteolysis, the effect of a lysosomal proteases inhibitor chloroquine was studied (Fig. 1B). MT1F was localized mainly in the cytoplasm in un-treated cells, however, chloroquine treatment induced accumulation of MT1F and co-localization with CD63 as dot-like spots. Suppression of degradation of surface-labeled MT1-MMP by chloroquine was confirmed by western blotting (Fig. 1C). These results indicate that a part of MT1-MMP internalized from cell surface is transported and degraded at CD63-positive lysosomes.

CD63 down-regulates MT1-MMP. The effect of CD63 on MT1-MMP expression was examined. Co-expression of MT1F with MMP-2 induced processing of MMP-2 from 68-kDa latent form to 64-kDa intermediate and 62-kDa active forms as shown by gelatin zymography (Fig. 2A). Transfection of CD63 plasmid down-regulated MMP-2 processing by dose dependent manner. Expression level of MT1-MMP was drastically reduced by CD63 expression as demonstrated by western blotting (Fig. 2B), which was clearly restored by the treatment of cells with chloroquine. MMP inhibitor BB94 had no effect. These results suggest that CD63

expression reduces the level of cell-surface MT1-MMP and promotes its lysosomal proteolysis. To analyze the mechanism of CD63-dependent proteolysis of MT1-MMP, cell-surface MT1-MMP on CD63-expressing cells was pulse-labeled with the antibody, and the internalization of MT1-MMP was monitored. Internalized MT1-MMP accumulated and co-localized with CD63 in lysosomes (Fig. 2C). These results may suggest that reduction of cell surface MT1-MMP by CD63 expression is due to promoted internalization and degradation at lysosome.

Interaction between CD63 and MT1-MMP. To analyze the interaction between MT1-MMP and CD63, immunoprecipitation was performed. As shown in Fig. 3B, CD63 was co-precipitated with MT1F. CD63 mutants lacking the C-terminal cytoplasmic tail or the C-terminal large extracellular loop (LEL) (Δ LEL) was co-precipitated with MT1-MMP, while mutants lacking the N-terminal small extracellular loop (SEL) (Δ SEL) of CD63 failed. In addition, although WT-CD63 reduced the level of MT1-MMP, the Δ C12, Δ SEL, and Δ LEL mutants of CD63 did not. The MT1-MMP mutant lacking hemopexin-like domain (Δ Pex) was not co-precipitated with CD63, whereas other mutants of MT1-MMP did (Fig. 3D). Consequently, co-expression of CD63 reduced the level of MT1F but not that of MT1F Δ Pex (Fig. 3E). These results suggest that N-terminus of CD63 and hemopexin-like domain of MT1-MMP are necessary for the association, but not sufficient to promote lysosomal proteolysis of MT1-MMP.

Role of cytoplasmic tails of MT1-MMP and CD63. The C-terminal cytoplasmic tail of CD63 contains the lysosomal targeting motif GYEYM, which interacts with the μ 2 subunit of adaptor protein (AP)-2 adaptor and the μ 3 subunit of AP-3 [17, 18]. In fact, the deficiency of AP-3 results in increased cell-surface expression of CD63 due to lack of its lysosomal trafficking [17]. The role of CD63 C-terminal cytoplasmic tail in promotion of MT1-MMP lysosomal proteolysis was studied. MT1F was localized at cell periphery in the cells expressing MT1F alone, and cell-surface staining of MT1F was significantly reduced in cells co-expressing GFP-CD63 (Fig. 4A). CD63 accumulated at lysosomes, however, CD63 Δ C12 was mainly distributed in cytoplasm and cell periphery. Cell-surface localization of MT1F was not significantly affected by CD63 Δ C12 expression. Figure 4B showed the relative levels of cell-surface MT1-MMP in cells co-expressing CD63 or their mutants. CD63 expression markedly reduced cell-surface expression of MT1-MMP, but MT1F Δ Pex was not significantly reduced by CD63 expression. Cell-surface expression of MT1F Δ CP was faintly down-regulated by CD63 expression. In addition, CD63 Δ C12 expression did not attenuate cell-surface expression of MT1F. Hence, CD63 expression apparently down-regulated MMP-2 activation mediated by MT1F, but that by MT1F Δ CP was affected only faintly by CD63 expression (Fig. 4C).

DISCUSSION

In the present study we demonstrated for the first time that a part of MT1-MMP internalized from cell surface is transported to CD63-positive lysosomes and degraded there. Ectopic expression of CD63 promoted lysosomal proteolysis of MT1-MMP (Fig. 2B). The direct interaction between MT1-MMP and CD63 was shown to be mediated through the hemopexin-like domain of MT1-MMP and N-terminal region of CD63 (Fig. 3). Thus, MT1-MMP level was not altered by the expression of CD63 Δ SEL, and MT1F Δ Pex by CD63 (Fig. 4). Targeting of the complex to lysosome is mediated by lysosomal targeting motif of CD63 (GYEYM) which is located at the C-terminal tail, and thus CD63 Δ C12 was unable to stimulate lysosomal proteolysis of MT1-MMP (Fig. 3B).

The importance of cytoplasmic tail of MT1-MMP for the internalization from cell surface was demonstrated by Uekita et al. [10] and in this study (Fig. 4). The cytoplasmic tails of MT1-MMP and CD63 interact with the μ 2 subunit of AP-2 adaptor. Thus, the interaction between MT1-MMP and CD63 may promote their internalization. The facts that expression of CD63 is ubiquitous and internalized MT1-MMP is co-localized with endogenous CD63 at lysosomes suggest that CD63 is one of regulators of MT1-MMP. However, the mechanism of CD63-mediated internalization of MT1-MMP still remained examined. Further study is required to clarify whether the association of CD63/MT1-MMP complex with integrin $\alpha_3\beta_1$, CD81, phosphatidylinositol 4 kinase [19], and

other proteins such as CD44 [20] and claudins [21] can regulate MT1-MMP enzymatic activity and dynamic turnover of MT1-MMP at cell periphery.

In conclusion, we have shown that CD63 is involved in the internalization of MT1-MMP from cell-surface, lysosomal targeting and proteolysis of MT1-MMP. Understanding the mechanism by which MT1-MMP is internalized from cell surface, transported to lysosomes, and presumably recycled back to plasma membrane might provide a step to regulate MT1-MMP activity which is closely associated with the invasiveness and metastasis of malignant tumor cells.

ACKNOWLEDGMENTS

This work was supported by a Grant-in-Aid for Science Research on Priority Areas from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (to T.T and H.S).

REFERENCE

- [1] H. Sato, T. Takino, Y. Okada, J. Cao, A. Shinagawa, E. Yamamoto, and M. Seiki, A matrix metalloproteinase expressed on the surface of invasive tumour cells, *Nature* 370 (1994) 61-65.
- [2] T. Takino, H. Sato, E. Yamamoto, and M. Seiki, Cloning of a human gene potentially encoding a novel matrix metalloproteinase having a C-terminal transmembrane domain, *Gene* 155 (1995) 293-298.

- [3] T. Takino, H. Sato, A. Shinagawa, and M. Seiki, Identification of the second membrane-type matrix metalloproteinase (MT-MMP-2) gene from a human placenta cDNA library. MT-MMPs form a unique membrane-type subclass in the MMP family, *J Biol Chem* 270 (1995) 23013-23020.
- [4] D. Pei, Identification and characterization of the fifth membrane-type matrix metalloproteinase MT5-MMP, *J Biol Chem* 274 (1999) 8925-8932.
- [5] M. Seiki, Membrane-type matrix metalloproteinases, *Apmis* 107 (1999) 137-143.
- [6] Y. Tsunozuka, H. Kinoh, T. Takino, Y. Watanabe, Y. Okada, A. Shinagawa, H. Sato, and M. Seiki, Expression of membrane-type matrix metalloproteinase 1 (MT1-MMP) in tumor cells enhances pulmonary metastasis in an experimental metastasis assay, *Cancer Res* 56 (1996) 5678-5683.
- [7] I. Yana, and S. J. Weiss, Regulation of membrane type-1 matrix metalloproteinase activation by proprotein convertases, *Mol Biol Cell* 11 (2000) 2387-2401.
- [8] K. Lehti, J. Lohi, H. Valtanen, and J. Keski-Oja, Proteolytic processing of membrane-type-1 matrix metalloproteinase is associated with gelatinase A activation at the cell surface, *Biochem J* 334 (Pt 2) (1998) 345-353.
- [9] A. Jiang, K. Lehti, X. Wang, S. J. Weiss, J. Keski-Oja, and D. Pei, Regulation of membrane-type matrix metalloproteinase 1 activity by dynamin-mediated endocytosis, *Proc Natl Acad Sci U S A* 98 (2001) 13693-13698.

- [10] T. Uekita, Y. Itoh, I. Yana, H. Ohno, and M. Seiki, Cytoplasmic tail-dependent internalization of membrane-type 1 matrix metalloproteinase is important for its invasion-promoting activity, *J Cell Biol* 155 (2001) 1345-1356.
- [11] F. Berditchevski, Complexes of tetraspanins with integrins: more than meets the eye, *J Cell Sci* 114 (2001) 4143-4151.
- [12] M. Fukuda, Lysosomal membrane glycoproteins. Structure, biosynthesis, and intracellular trafficking, *J Biol Chem* 266 (1991) 21327-21330.
- [13] H. Hotta, A. H. Ross, K. Huebner, M. Isobe, S. Wendeborn, M. V. Chao, R. P. Ricciardi, Y. Tsujimoto, C. M. Croce, and H. Koprowski, Molecular cloning and characterization of an antigen associated with early stages of melanoma tumor progression, *Cancer Res* 48 (1988) 2955-2962.
- [14] M. Kondoh, M. Ueda, M. Ichihashi, and Y. Mishima, Decreased expression of human melanoma-associated antigen ME491 along the progression of melanoma pre-canceroses to invasive and metastatic melanomas, *Melanoma Res* 3 (1993) 241-245.
- [15] K. J. Radford, J. Mallesch, and P. Hersey, Suppression of human melanoma cell growth and metastasis by the melanoma-associated antigen CD63 (ME491), *Int J Cancer* 62 (1995) 631-635.
- [16] T. Kinoshita, H. Sato, A. Okada, E. Ohuchi, K. Imai, Y. Okada, and M. Seiki, TIMP-2 promotes activation of progelatinase A by membrane-type 1 matrix metalloproteinase immobilized on agarose beads, *J Biol Chem* 273 (1998) 16098-16103.

[17] E. C. Dell'Angelica, V. Shotelersuk, R. C. Aguilar, W. A. Gahl, and J. S. Bonifacino, Altered trafficking of lysosomal proteins in Hermansky-Pudlak syndrome due to mutations in the beta 3A subunit of the AP-3 adaptor, *Mol Cell* 3 (1999) 11-21.

[18] T. Kobayashi, U. M. Vischer, C. Rosnoblet, C. Lebrand, M. Lindsay, R. G. Parton, E. K. Kruithof, and J. Gruenberg, The tetraspanin CD63/lamp3 cycles between endocytic and secretory compartments in human endothelial cells, *Mol Biol Cell* 11 (2000) 1829-1843.

[19] F. Berditchevski, K. F. Tolia, K. Wong, C. L. Carpenter, and M. E. Hemler, A novel link between integrins, transmembrane-4 superfamily proteins (CD63 and CD81), and phosphatidylinositol 4-kinase, *J Biol Chem* 272 (1997) 2595-2598.

[20] H. Mori, T. Tomari, N. Koshikawa, M. Kajita, Y. Itoh, H. Sato, H. Tojo, I. Yana, and M. Seiki, CD44 directs membrane-type 1 matrix metalloproteinase to lamellipodia by associating with its hemopexin-like domain, *Embo J* 21 (2002) 3949-3959.

[21] H. Miyamori, T. Takino, Y. Kobayashi, H. Tokai, Y. Itoh, M. Seiki, and H. Sato, Claudin promotes activation of pro-matrix metalloproteinase-2 mediated by membrane-type matrix metalloproteinases, *J Biol Chem* 276 (2001) 28204-28211.

FIGURE LEGENDS

FIG. 1. Localization of internalized MT1-MMP. (A) HeLa cells were transfected with MT1F, and

incubated with anti-FLAG antibody for 30 min at 4 °C. After washing, the cells were cultured in complete media for 0, 30 min, or 3 h, and then performed immunofluorescence staining under either permeabilized (+TX100) or non-permeabilized (-TX100) conditions as described in Materials and Methods. (B) HeLa cells were transfected with MT1F, and cultured in the presence (+) or absence (-) of chloroquine (CHQ; 58 μ M) for 6h. The cells were fixed under permeabilized condition, and then stained with anti-CD63 and anti-MT1-MMP antibodies. Bar, 20 μ m. (C) Cell-surface biotinylation and immunoprecipitation were carried out as described in Materials and Methods. Cell-surface MT1F was detected by streptavidine-HRP (Blot: SA-HRP), and total MT1F was by anti-FLAG antibody (Blot: FLAG). The whole-cell lysates (WCL) were immunoblotted with anti-actin (Blot: Actin) antibody.

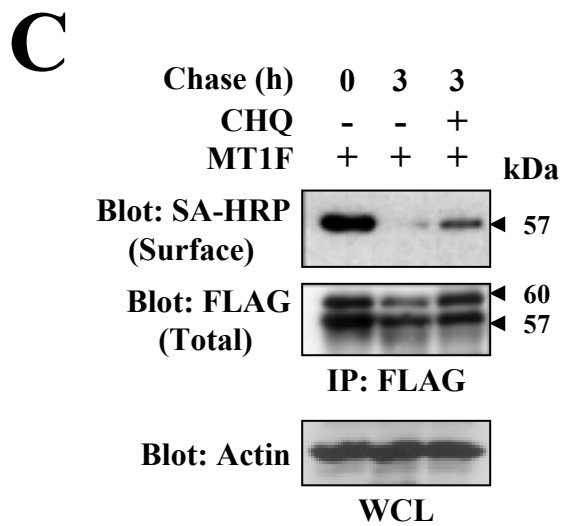
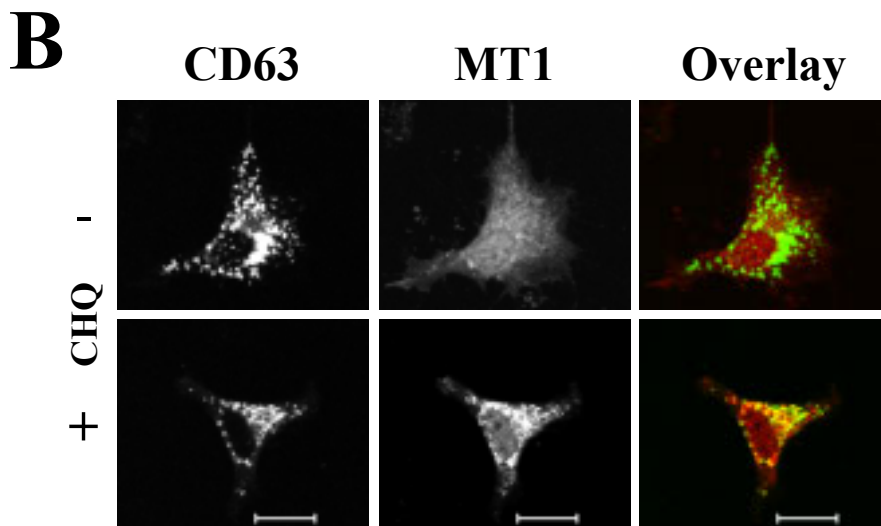
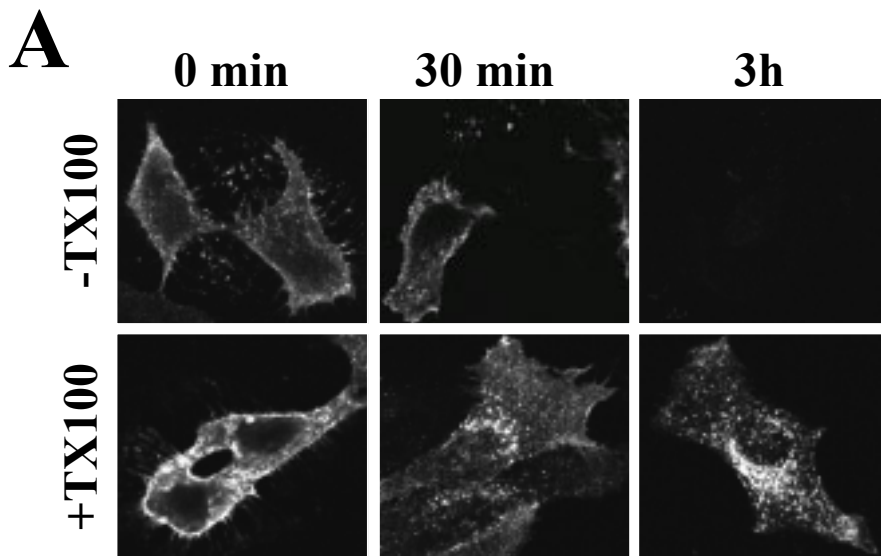
FIG. 2. CD63 down-regulates MT1-MMP. (A) 293-EBNA cells were co-transfected with MMP-2, MT1F and 50 ng, 200 ng, or 1 μ g of GFP-CD63 plasmids. The conditioned media were analyzed by gelatin zymography. (B) 293-EBNA cells were co-transfected with MT1F and either GFP or GFP-CD63 plasmids, and treated with DMSO, BB94, chloroquine (CHQ) overnight. The cells were homogenized, and immunoblotted with anti-FLAG (Blot: FLAG) or anti-GFP (Blot: GFP) antibody. (C) MDCK cells stably expressing GFP-CD63 were transiently transfected with MT1F, incubated with anti-FLAG antibody conjugated with AlexaTM546 for 30 min, and cultured further

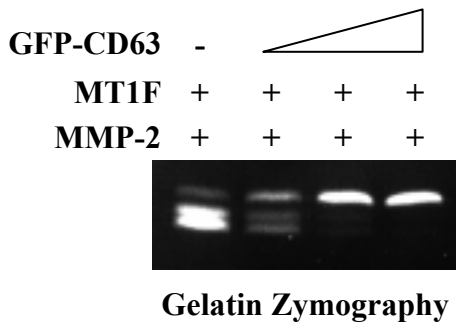
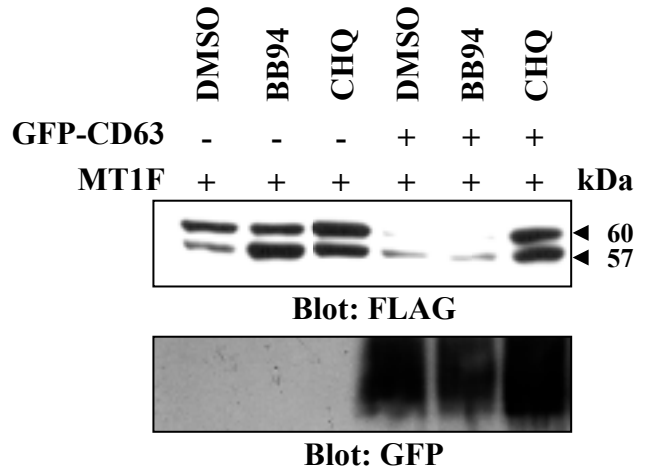
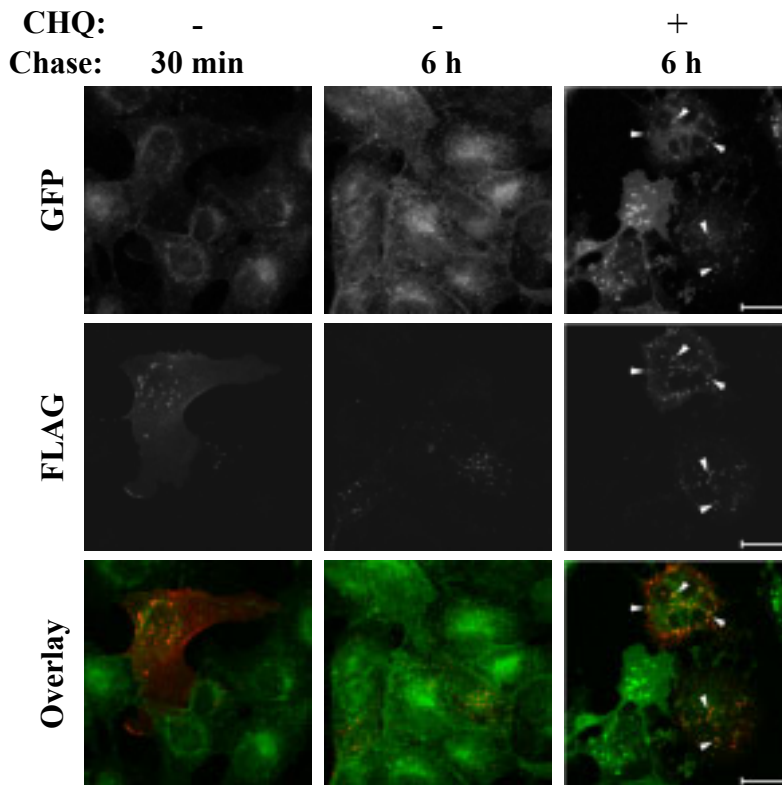
30 min or 6 h in the presence of BB94 with or without chloroquine (CHQ). The cells were fixed with 4% paraformaldehyde. Bar, 20 μ m.

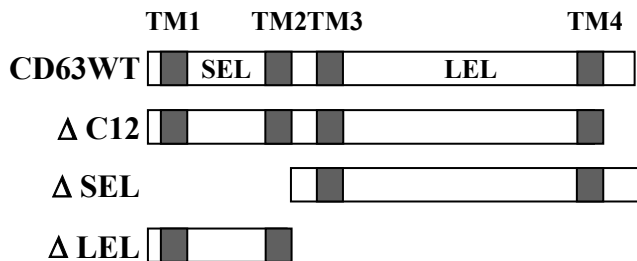
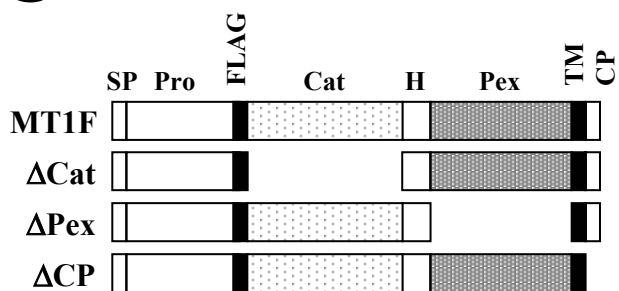
FIG. 3. Interaction between CD63 and MT1-MMP. (A) CD63 composes with four transmembrane domains (TM1 to TM4), and two extracellular loop, SEL and LEL. 293-EBNA cells were co-transfected with MT1F and GFP-CD63 mutants (B) or co-transfected with GFP-CD63 Δ C12 and MT1-MMP mutants (D). The cell lysates were precipitated with anti-FLAG antibody (IP: FLAG), and analyzed by immunoblotting with anti-GFP antibody (Blot: GFP). The WCL were immunoblotted with anti-FLAG (Blot: FLAG) or anti-GFP (Blot: GFP) antibody. (C) a schematic representation of MT1-MMP mutants. SP, signal peptide; Pro, propeptide; Cat, catalytic domain; H, hinge region; Pex, hemopexin-like domain; TM, transmembrane domain; CP, cytoplasmic domain; FLAG, FLAG epitope. (E), HeLa cells were co-transfected with MT1F or MT1F Δ Pex and GFP or GFP-CD63 plasmids, and cultured in the presence of BB94 (1 μ M) for 12 h. The cells were homogenized, and immunoblotted with anti-FLAG (Blot: FLAG) antibody.

FIG. 4. Role of cytoplasmic tails of MT1-MMP and CD63. (A) HeLa cells were co-transfected with MT1F or MT1F Δ CP and either GFP, GFP-CD63 or GFP-CD63 Δ C12 plasmids. The cell-surface MT1-MMP were labeled with anti-FLAG M2 antibody for 30 min at 4°C, and the

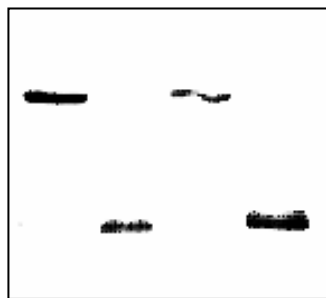
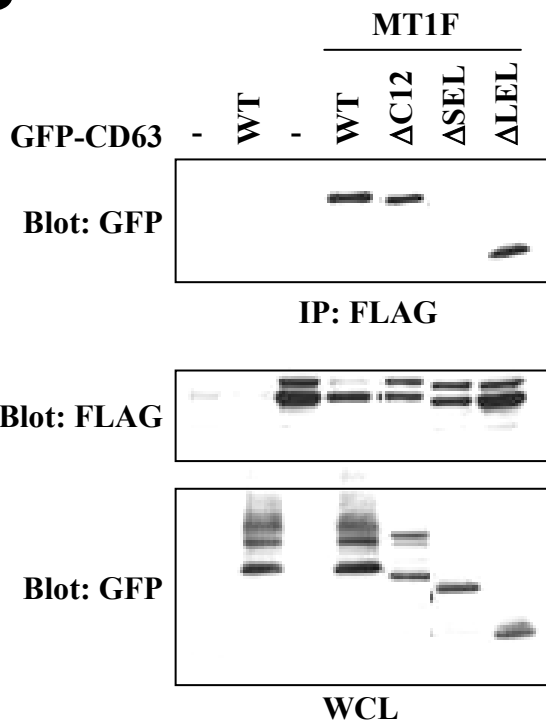
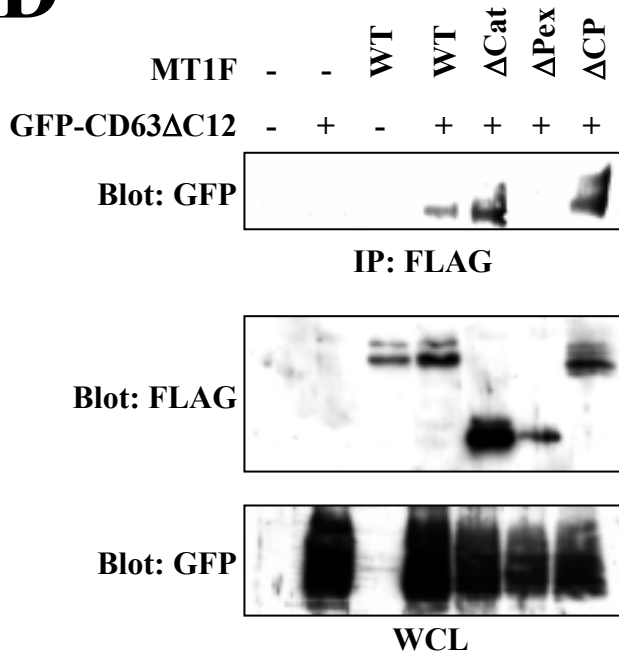
immunofluorescence staining was performed under non-permeabilized condition. Bar, 20 μm . (B) the values of fluorescence intensity per cell were determined by immunofluorescence staining. Data was collected from at least 30 cells from three independent experiments. Asterisk, $P < 0.0001$. Error bars indicate SD. (C) HeLa cells were co-transfected with MT1F or MT1F Δ CP, MMP-2, and GFP or GFP-CD63 plasmids as described above. The conditioned media were analyzed by gelatin zymography.

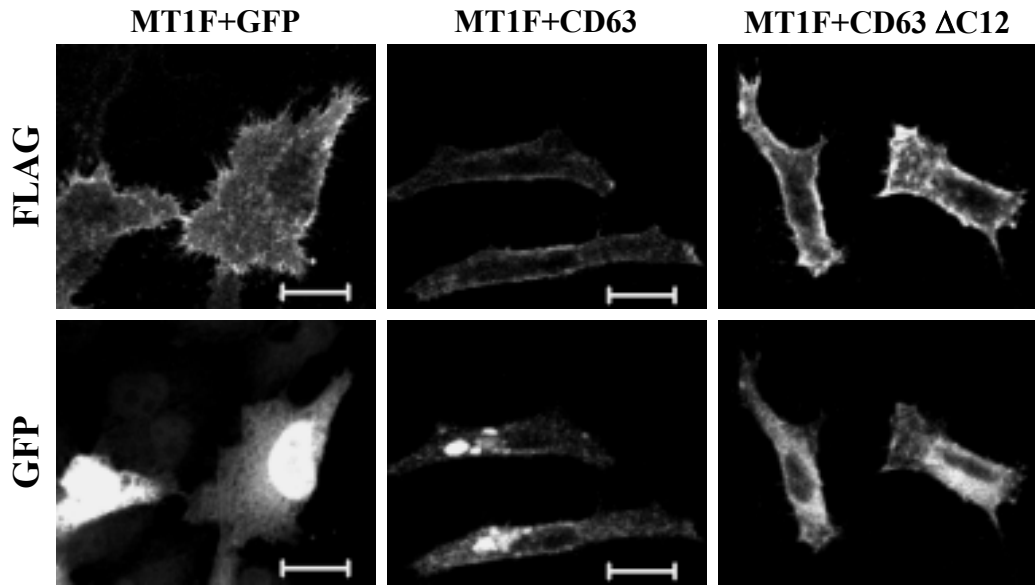
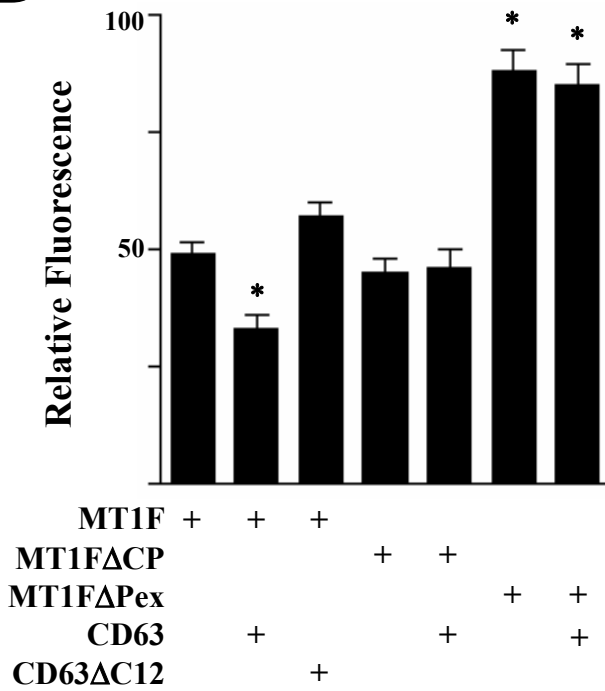


A**B****C**

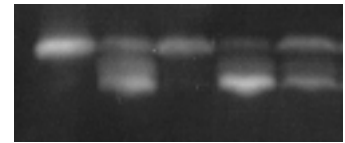
A**C****E**

MT1FΔPex		+		+
MT1F	+		+	
CD63			+	+

**B****D**

A**B****C**

MT1FΔCP	-	-	-	+	+
MT1F	-	+	+	-	-
CD63	-	-	+	-	+
MMP-2	+	+	+	+	+

**Gelatin Zymography**