

Effect of gemcitabine on the expression of apoptosis-related genes in human pancreatic cancer cells

メタデータ	言語: English 出版者: 公開日: 2017-10-05 キーワード (Ja): キーワード (En): 作成者: Jiang, Pei-Hong, Motoo, Yoshiharu, Sawabu, Norio, Minamoto, Toshinari メールアドレス: 所属:
URL	http://hdl.handle.net/2297/11866

Effect of gemcitabine on the expression of apoptosis-related genes in human pancreatic cancer cells

Pei-Hong Jiang, Yoshiharu Motoo, Norio Sawabu, Toshinari Minamoto

Pei-Hong Jiang, Norio Sawabu, Department of Internal Medicine and Medical Oncology, Cancer Research Institute, Kanazawa University, Kanazawa, Japan

Yoshiharu Motoo, Department of Medical Oncology, Kanazawa Medical University, Uchinada, Ishikawa, Japan

Toshinari Minamoto, Division of Diagnostic Molecular Oncology, Cancer Research Institute, Kanazawa University, Kanazawa, Japan

Correspondence to: Yoshiharu Motoo, MD, Department of Medical Oncology, Kanazawa Medical University, 1-1 Daigaku, Uchinada, Ishikawa 920-0293,

Japan. motoo@kanazawa-med.ac.jp

Telephone: +81-76-2188284 Fax: +81-76-2188283

Received: 2005-07-11 Accepted: 2005-11-18

and GSK-3 β , and the activation of *TP53INP1* and phospho-GSK-3 β ^{ser9}.

© 2006 The WJG Press. All rights reserved.

Key words: Gemcitabine; Pancreatitis-associated protein; TP53INP1; GSK-3 β ; PANC-1

Jiang PH, Motoo Y, Sawabu N, Minamoto T. Effect of gemcitabine on the expression of apoptosis-related genes in human pancreatic cancer cells. *World J Gastroenterol* 2006; 12(10): 1597-1602

<http://www.wjgnet.com/1007-9327/12/1597.asp>

Abstract

AIM: To investigate the expression of genes involved in the gemcitabine-induced cytotoxicity in human pancreatic cancer cells.

METHODS: A human pancreatic cancer cell line, PANC-1, was cultured. 1×10^4 PANC-1 cells were plated in 96-well microtiter plates. After being incubated for 24 h, gemcitabine was added to the medium at concentrations ranging 2.5 -1 000 mg/L. The AlamarBlue dye method was used for cell growth analysis. DNA fragmentation was quantitatively assayed using a DNA fragmentation enzyme-linked immunosorbent assay (ELISA) kit. *PAP* and *TP53INP1* mRNA expression was determined using the reverse transcription-polymerase chain reaction with semi-quantitative analysis. The expression of GSK-3 β and phospho-GSK-3 β proteins was examined with Western blot analysis.

RESULTS: The IC₅₀ for the drug after a 48-h exposure to gemcitabine was 16 mg/L. The growth of PANC-1 cells was inhibited by gemcitabine in a concentration-dependent manner ($P < 0.0001$) and the cell growth was also inhibited throughout the time course ($P < 0.0001$). The DNA fragmentation rate in the gemcitabine-treated group at 48 h was 44.7 %, whereas it was 25.3 % in the untreated group. The *PAP* mRNA expression was decreased after being treated with gemcitabine, whereas the *TP53INP1* mRNA was increased by the gemcitabine treatment. Western blot analysis showed that phospho-GSK-3 β ^{ser9} was induced by the gemcitabine treatment.

CONCLUSION: Gemcitabine suppresses PANC-1 cell proliferation and induces apoptosis. Apoptosis is considered to be associated with the inhibition of PAP

INTRODUCTION

Pancreatic cancer is the fifth most common cause of cancer death in Japan^[1]. It is estimated that there are 20000 cases every year, which is similar to the number of deaths from this disease. The reasons for its very high mortality rate include the lack of early diagnosis, a low resectability at the time of initial diagnosis, and the rapid recurrence after resection. Surgery is rarely a curative option in pancreatic cancers because of local extension and metastasis. Chemotherapy for advanced pancreatic cancer is palliative. The use of 5-FU in combination with radiation in the locally advanced setting has been shown to enhance survival. In a randomized trial between gemcitabine and 5-FU, gemcitabine showed significantly better results in terms of the clinical benefit effect and survival^[2].

Gemcitabine (2', 2'-difluorodeoxycytidine) is a nucleoside analogue well known for its antitumor activity in several solid tumors^[3]. It is one of the drugs effective for pancreatic cancers in the clinical treatment^[4]. Various general mechanisms of gemcitabine have been described^[5-7]. A primary mechanism of gemcitabine is the blocking of DNA synthesis by inhibiting DNA polymerase activity^[8]. Gemcitabine itself is not active until it enters the cells. Its intracellular transport is mediated by facilitated diffusion^[9]. Intracellularly, gemcitabine is phosphorylated to its active metabolites by deoxycytidine kinase to difluorodeoxycytidine monophosphate (dfdCMP), difluorodeoxycytidine diphosphate (dfdCDP) and difluorodeoxycytidine triphosphate (dfdCTP). All three of the metabolites interfere at different steps in the processing of DNA. The dfdCTP is incorporated into

DNA and as such can obstruct DNA replication and repair. The dfdCTP can also be incorporated into RNA and can inhibit CTP-synthetase. The dfdCMP can also inhibit dCMP-deaminase. The dfdCDP is an inhibitor of ribonucleotide reductase, and its action was shown to lead to the depletion of the DNA precursor pool, dNTP. Inhibition of DNA synthesis leads to growth inhibition or cell death. In previous studies, gemcitabine has been shown to play a major role in the apoptosis of certain tumor cell lines^[10-12]. There are several pathways that relate to gemcitabine-induced apoptosis. Nicole *et al*^[13] reported that gemcitabine-mediated apoptosis is caspase-dependent in pancreatic cancers; Jones *et al*^[14] showed that gemcitabine-induced apoptosis is achieved through the blocking of NF- κ B in non-small cell lung cancer cells (NSCLC). In addition, gemcitabine-induced apoptosis is also related to signal-regulated kinase (ERK), Akt, Bcl-2 and p38 mitogen-activated protein kinase (MAPK) pathways^[15-17]. Achiwa *et al*^[18] indicated that the increase in human equilibrate nucleoside transport (hENT) expression is a determinant of gemcitabine sensitivity in NSCLC cells. However, the molecular mechanism of gemcitabine-induced apoptosis has not been fully elucidated.

Pancreatitis-associated protein (PAP) is a secretory protein of pancreatic acinar cells. It is almost absent in the normal pancreas, but is induced in acute and chronic pancreatitis. We have reported the expression of *PAP* mRNA in cancer tissues and have measured *PAP* levels in the sera and pancreatic juice of patients with gastrointestinal cancers^[19-21]. We found that serum *PAP* levels were increased in 40% of patients with pancreatic cancer. We also reported that *PAP* levels in endoscopically aspirated pancreatic juice were positive in 55% of pancreatic cancers. *PAP* levels were significantly higher in both the serum and pancreatic juice in cases of pancreatic cancer, compared to chronic pancreatitis. Cytokines such as tumor necrosis factor- α , interferon- γ , and interleukin-6 induce *PAP* mRNA expression in the pancreatic acinar AR4-2J cell line. We found that the enhanced expression of *PAP* in pancreatic adenocarcinoma is caused by both ectopic expression in cancer cells and induction in acinar cells^[22].

TP53INP1^[23] was previously called stress-induced protein (SIP)^[24] or p53-dependent inducible nuclear protein (p53DINP1)^[25]. *TP53INP1* is strongly induced in acinar cells during acute pancreatitis in mice, and is also overexpressed in response to various stresses *in vitro*. *TP53INP1* gene expression is wild-type p53-dependent^[26]. There is a functional p53-response element within the promoter region of the *TP53INP1* gene, and *TP53INP1* mRNA expression is activated in cells expressing wild-type p53 in response to various stresses. One of the major functions of *TP53INP1* is promoting cellular apoptosis. Glycogen synthase kinase 3 β (GSK-3 β) is a multifunctional serine/threonine kinase mediating various cellular signaling pathways. The particular pathway depends on its substrates for phosphorylation^[27]. Since GSK-3 β is also an important mediator of an apoptotic signal, it is plausible that the GSK-3 β deregulation observed in cancer cells confers resistance to chemotherapy, which is a major cause of treatment failure in human cancers^[28].

In this study we investigated the effect of gemcitabine on the PANC-1 cells in terms of apoptosis-related factors.

MATERIALS AND METHODS

Cell culture and gemcitabine treatment

A human pancreatic cancer cell line, PANC-1, obtained from the American Type Culture Collection (ATCC, MD, USA), was maintained in Dulbecco's modified Eagle's medium supplemented with 100 mL/L fetal calf serum, penicillin, and kanamycin at 37°C in a 50 mL/L CO₂, 950 mL/L air atmosphere. Gemcitabine (Eli-Lilly Japan, Kobe, Japan) at a concentration of 50 mg/mL was dissolved in the serum free culture medium and stored at -20°C in the freezer. The concentration range of the treatment was from 2.5 mg/L to 1 000 mg/L.

Cell growth evaluation

The Alamarblue dye method was used for cell growth analysis. The 1×10^4 cells were plated in 96-well microtiter plates. After being incubated for 24 h, gemcitabine was added to the medium. Twenty μ L of AlamarBlue dye solution (Iwaki Glassware, Inc., Tokyo, Japan) was added to wells containing 200 μ L of medium at the time 12, 24, 48, and 72 h. After being incubated for 3 h, the cell growth was evaluated as the absorbance (A) using a spectrophotometer (Dai-Nippon Pharmaceutical Co., Osaka, Japan). An excitation wavelength of 540 nm was used, and the emission was read at 620 nm. The color of AlamarBlue stock is violet, and changes to red when oxidized. Each treatment was applied to 6 wells, and the experiments were repeated 3 times.

DNA fragmentation assay

DNA fragmentation was quantitatively assayed using a DNA fragmentation enzyme-linked immunosorbent assay (ELISA) kit (Boehringer Mannheim GmbH, Mannheim, Germany) according to the protocol. Cells were cultured in flat-bottom, 96-well microplates. After incubation in gemcitabine-supplemented media for 24 h, the cells were detached from the wells. The cells were lysed with lysis buffer, and the lysate was processed for streptavidin-coated microtiter plates. After incubation with biotin-labeled antihistone antibody and peroxidase-labeled anti-DNA antibody, the amount of fragmented DNA was determined using 2, 2'-azino-bis-3-ethylbenzthiazoline-sulfonate (ABTS) as a substrate. The plates were read on a Labsystems integrated EIA (Dai-Nippon Pharmaceutical Co., Osaka, Japan) at wavelengths of 540 nm and 620 nm. Each treatment was added to duplicate wells, and the experiment repeated 3 times.

Total RNA was extracted from pancreatic cancer cells using a SV Total RNA Isolation system kit (Promega, Madison, WI, USA). RNA concentrations were determined with spectrophotometry. RT was performed using a PowerScriptTM Reverse Transcriptase kit (Clontech Laboratories, Inc., Palo Alto, CA, USA). First strand cDNA was synthesized from 8 μ g of total RNA at 65°C for 5 min after RT; the reverse transcriptase was inactivated by incubating at 42°C for 60 min and the reaction was terminated by heating at 70°C for 25 min. Then, cycles

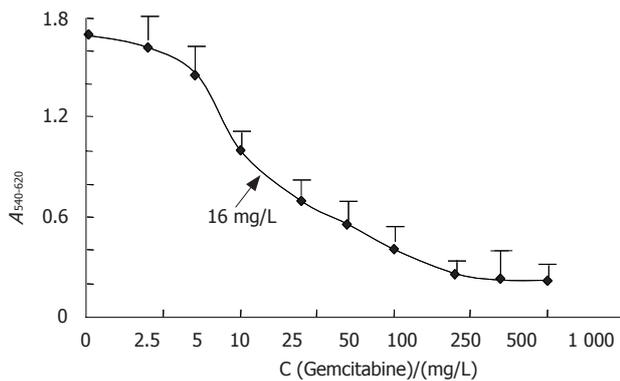


Figure 1 Effect of gemcitabine on the growth of PANC-1 cells.

of amplification were performed on a DNA thermal cycler (Perkin Elmer Cetus, Inc., Norwalk, CT, USA) as follows: *PAP* primers were denatured at 91°C for 60 sec, annealed at 54°C for 60 sec, and polymerized at 72°C for 45 sec. Then, extension was performed at 72°C for 10 min. *TP53INP1* primers were denatured at 94°C for 50 sec, annealed at 62°C for 40 sec, and polymerized at 72°C for 50 sec. The extension was performed at 72°C for 10 min. The *PAP* primer pairs were sense: 5'-CTCCTGATTGCCTCCTCAAG -3' and antisense: 5'-AAACGTACCCTCTCTTTAGG -3', producing a fragment of 441 bp. *TP53INP1* mRNA was specifically amplified with the following primers: sense 5'-CATCCAGCCAAACTCTCAGTC -3' and antisense 5'-GCGACGAAGGCTATTCTGT -3'. The size of the fragment was 703 base pairs (bp). The *GAPDH* of 452 bp was used as an internal control. Eight-microgram aliquots of the RT-PCR products were subjected to electrophoresis on a 20 g/L agarose gel and visualized with SYBR Gold (Molecular Probes, Inc., Eugene, OR, USA) at a 1:10000 dilution in dimethylsulfoxide, and exposed to ultraviolet 312-nm light.

Semiquantitative RT-PCR analysis

The gene expression of *PAP* and *TP53INP1* was semiquantitatively analyzed with an image analyzer (ATTO Densitograph ver. 3.02, ATTO, Inc., Tokyo, Japan). The relative expression intensity was calculated according to the following formula: *PAP* and *TP53INP1* mRNA in a sample/*GAPDH* mRNA in a sample.

Western blot analysis

For Western blot analysis, 1×10^6 PANC-1 cells were lysed in a CellLytic™ mammalia tissue lysis/ extraction reagent buffer (Sigma-Aldrich, St. Louis, MO, USA). Aliquots were boiled for 5 minutes with Laemli buffer, resolved on a 125 g/L SDS-PAGE gel and electrophoretically transferred to nitrocellulose membrane (Amersham, Buckingham, UK). Membranes were blocked for 1 h in TBS containing 50 g/L dry milk and 1 g/L Tween 20. Membranes were then incubated with either anti-rabbit phospho-GSK-3 β (ser9) polyclonal antibody (Cell Signaling Technology, Inc. Beverly, MA, USA) or anti-mouse GSK-3 β monoclonal antibody (BD Transduction Laboratories, Lexington, KY, USA) for one night at 4°C. After incubation with anti-

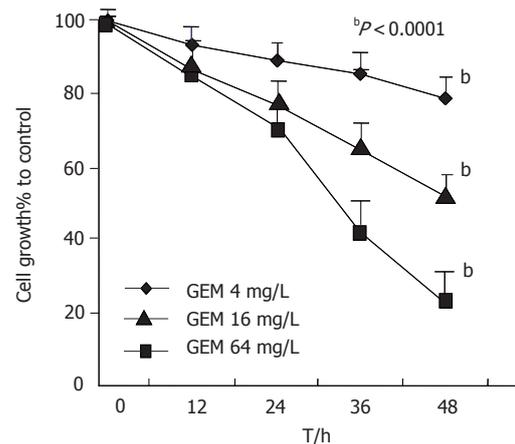


Figure 2 Inhibitory effect of gemcitabine on the growth of PANC-1 cells.

rabbit or anti-mouse IgG second antibody labeled with peroxidase, the membranes were visualized with an ATTO image analyzer.

Statistical analysis

Experimental results were expressed as the mean \pm SE. The difference between the means was evaluated with the Mann-Whitney *U* test. $P < 0.05$ was considered statistically significant. The statistical analysis was performed using StatView-5.0 (SAS Institute Inc. Tokyo, Japan).

RESULTS

Gemcitabine inhibited cell growth

The median 50% inhibitory concentration (IC_{50}) of gemcitabine after a 48-h exposure was 16 mg/L (Figure 1). We also measured the effect of 4, 16 and 64 mg/L gemcitabine at 12, 24, 36, and 48 h. The growth of PANC-1 cells was inhibited by gemcitabine in a concentration-dependent manner ($P < 0.0001$, Figure 2) and cell growth was also inhibited throughout the time course ($P < 0.0001$).

Gemcitabine induced apoptosis

1×10^4 PANC-1 cells were cultured in flat-bottom, 96-well microplates. After incubation in gemcitabine-supplemented media for 24 h, a DNA fragmentation assay was performed. The concentration of gemcitabine used in the DNA fragmentation assay was 16 mg/L. The DNA fragmentation rate in the gemcitabine-treated group was 44.7%, whereas the fragmentation rate in the untreated group was 25.3%. The fragmentation in the gemcitabine treated group was significantly higher than that in the untreated group ($P = 0.0157$).

Expression of *PAP*, *TP53INP1* mRNA

We exposed the PANC-1 cells to 4, 16 and 64 mg/L gemcitabine for 48 h. The expression of *PAP* and *TP53INP1* mRNA was assessed using RT-PCR. The *PAP* mRNA expression was decreased after being treated with gemcitabine, and disappeared after being treated with 16 mg/L gemcitabine. The semiquantitative analysis showed the intensity of 16 mg/L gemcitabine-treated

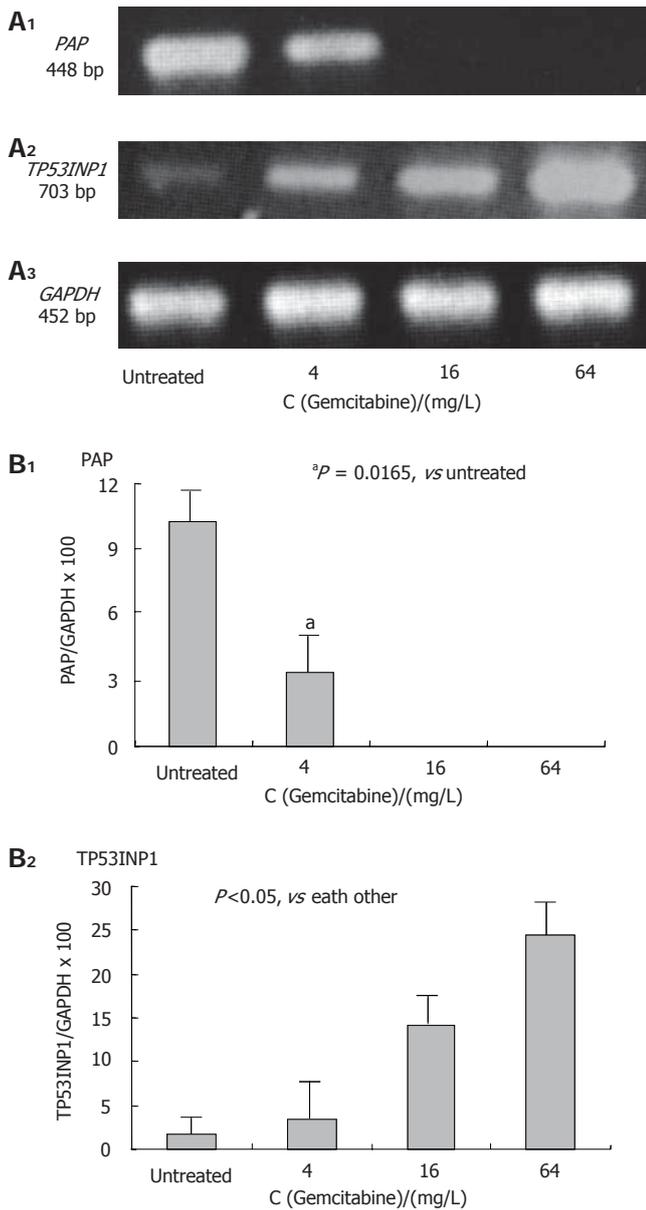


Figure 3 PAP mRNA expression after gemcitabine treatment.

cells to be significantly lower than that of the untreated cells ($P = 0.0165$, Figure 3). The *TP53INP1* mRNA was increased after exposure to gemcitabine. The intensity of the cells treated with 4, 16 and 64 mg/L gemcitabine was 2 times, 8.5 times and 14.6 times higher than in the untreated cells, respectively. The increasing density of *TP53INP1* mRNA was concentration-dependent. The P value of the interrelation of the untreated group to the 4 mg/L gemcitabine-treated group, 16 mg/L gemcitabine-treated group and 64mg/L gemcitabine-treated group were 0.0015, 0.0009 and 0.0352, respectively.

Expression of phospho-GSK-3β and GSK-3β proteins

The PANC-1 cell line showed no detectable phospho-GSK-3β^{ser9} in Western blotting analysis with a phospho-specific antibody, indicating GSK-3β activity in this cell line (Figure 4). On the other hand, phospho-GSK-3β^{ser9} was induced after 4 mg/L gemcitabine treatment, and gradually increased with the 16 and 64 mg/L gemcitabine

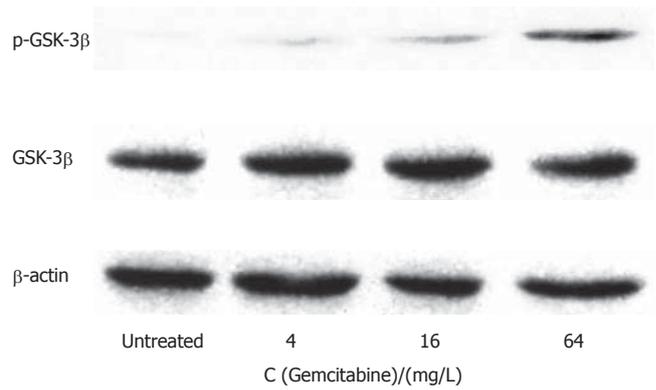


Figure 4 Effect of gemcitabine treatment on GSK-3β expression.

treatment. There was no significant difference in GSK-3β expression between the gemcitabine-treated and untreated groups.

DISCUSSION

Gemcitabine is a nucleotide analog that is converted to its triphosphate active form in cells and is subsequently incorporated into DNA to terminate strand elongation^[13-17]. It shows a favorable clinical outcome in the treatment of pancreatic cancer and non-small cell lung cancer^[14,18]. However, there have been few published *in vitro* analyses about gemcitabine. We therefore attempted to investigate the genes involved in the gemcitabine-induced cytotoxicity of pancreatic cancer cells. In the present study we showed that gemcitabine induced changes in expression of apoptosis-related genes in PANC-1 cells. The expression of anti-apoptotic pancreatitis-associated protein (PAP) was down-regulated, whereas the pro-apoptotic *TP53INP1* and p-GSK-3β were up-regulated after being treated with gemcitabine.

Apoptosis is believed to play an important role in tumors, and both pro- and anti-apoptotic factors are simultaneously activated in tumor development and progression. In this study, we showed that *PAP* expression was significantly decreased in the gemcitabine-induced anti-tumor process. The growth of human pancreatic cancer may require the up-regulation of *PAP* expression, which would suppress the apoptosis of cancer cells^[19]. *PAP* is one of the effectors of antiapoptosis induced by tumor necrosis factor-α through NF-κB and MAP kinases in pancreatic acinar cells^[29]. It was demonstrated that the activation status of the NF-κB, Akt and MAP kinase signaling pathways is often associated with anti-apoptotic signal transduction that is linked to chemotherapeutic agents^[16]. NF-κB plays an important role in oncogenesis and promotes cellular resistance to anticancer therapy. Bandala *et al*^[30] showed that gemcitabine treatment increased the activity of NF-κB, and Alexander *et al*^[31] showed that the induction of NF-κB by gemcitabine was dose-dependent in five pancreatic cancer cell lines. In addition, gemcitabine specifically activates p38 MAP kinases in the human pancreatic cancer cell lines, PK-1 and PCI43^[16]. Taken together, these findings confirmed that *PAP* is involved in the gemcitabine induced-apoptosis in a

dose-dependent manner.

Apoptosis is one of the major consequences of chemotherapy against malignancies. It is now established that the tumor suppressor p53 inhibits cell growth through the activation of cell cycle arrest and apoptosis. *TP53INP1* is a gene that is activated by wild type p53. It cooperates with HIPK2 to promote p53 phosphorylation at ser 46, and then induces apoptosis-inducing protein activation^[25, 26]. In our conditions, we showed *TP53INP1* was up-regulated by gemcitabine treatment from 4 mg/L to 64 mg/L, in a dose-dependent manner. *TP53INP1* may be involved in gemcitabine-mediated apoptosis. The precise mechanism of the *TP53INP1* increase after gemcitabine treatment is unclear. It has been shown that phosphorylation of the NH₂-terminal residues of p53 mediates its stabilization and nuclear accumulation after anticancer drug treatment^[28].

Beurel *et al.*^[28] has reported that GSK-3 β is hyperphosphorylated on serine 9 in human hepatoma cell lines as well as in human and murine tumoral livers. We examined the impact of gemcitabine on GSK-3 β activity in PANC-1 cells by measuring the phosphorylation level of GSK-3 β at serine 9 as an indicator of GSK-3 β inactivation. GSK-3 β can act as a positive or negative physiological regulator of the p53 protein. During endoplasmic reticulum stress, p53 is inhibited through a mechanism involving its phosphorylation at ser 315 and ser 376 by GSK-3 β . On the other hand, GSK-3 β plays a crucial role in cell survival mediated by nuclear factor-kappaB (NF- κ B) signaling. Phospho-GSK-3 β ^{ser9} is an inactive form in normal tissues that suggests the regulation of the balanced expression of the active and inactive forms of this kinase. In our data, phospho-GSK-3 β ^{ser9} was induced by gemcitabine treatment, indicating that GSK-3 β gradually became inactive in the gemcitabine-treated cells. Our results are similar to those of Beurel *et al.*^[28], who showed that GSK-3 β ^{ser9} phosphorylation by lithium treatment on tumor cells conferred resistance to anticancer therapy.

In conclusion, gemcitabine decreases cell proliferation of a human pancreatic cancer cell line, PANC-1, and promotes cellular apoptosis. The antiapoptotic gene, *PAP*, is down-regulated by gemcitabine treatment, whereas the pro-apoptotic *TP53INP1* gene and GSK-3 β ^{ser9} protein are up-regulated. Gemcitabine can induce apoptosis in cancer cells through GSK-3 β and *PAP* inhibition, and *TP53INP1* and GSK-3 β ^{ser9} activation.

REFERENCES

- 1 Okusaka T, Matsumura Y, Aoki K. New approaches for pancreatic cancer in Japan. *Cancer Chemother Pharmacol* 2004; **54** Suppl 1: S78-S82
- 2 Burris HA 3rd, Moore MJ, Andersen J, Green MR, Rothenberg ML, Modiano MR, Cripps MC, Portenoy RK, Storniolo AM, Tarassoff P, Nelson R, Dorr FA, Stephens CD, Von Hoff DD. Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. *J Clin Oncol* 1997; **15**: 2403-2413
- 3 Lawrence TS, Davis MA, Hough A, Rehemtulla A. The role of apoptosis in 2',2'-difluoro-2'-deoxycytidine (gemcitabine)-mediated radiosensitization. *Clin Cancer Res* 2001; **7**: 314-319
- 4 Diaz-Rubio E. New chemotherapeutic advances in pancreatic, colorectal, and gastric cancers. *Oncologist* 2004; **9**: 282-294
- 5 Moufarij MA, Phillips DR, Cullinane C. Gemcitabine potentiates cisplatin cytotoxicity and inhibits repair of cisplatin-DNA damage in ovarian cancer cell lines. *Mol Pharmacol* 2003; **63**: 862-869
- 6 Serrano MJ, Sanchez-Rovira P, Algarra I, Jaen A, Lozano A, Gaforio JJ. Evaluation of a gemcitabine-doxorubicin-paclitaxel combination schedule through flow cytometry assessment of apoptosis extent induced in human breast cancer cell lines. *Jpn J Cancer Res* 2002; **93**: 559-566
- 7 Galmarini CM, Clarke ML, Jordheim L, Santos CL, Cros E, Mackey JR, Dumontet C. Resistance to gemcitabine in a human follicular lymphoma cell line is due to partial deletion of the deoxycytidine kinase gene. *BMC Pharmacol* 2004; **4**: 8
- 8 Smith JA, Brown J, Martin MC, Ramondetta LM, Wolf JK. An in vitro study of the inhibitory activity of gemcitabine and platinum agents in human endometrial carcinoma cell lines. *Gynecol Oncol* 2004; **92**: 314-319
- 9 van Putten JWG, Groen HJM, Smid K, Peters GJ, Kampinga HH. End-joining deficiency and radiosensitization induced by gemcitabine. *Cancer Res* 2001; **61**: 1585-1591
- 10 Satoh K, Kaneko K, Hirota M, Toyota T, Shimosegawa T. The pattern of CPP32/caspase-3 expression reflects the biological behavior of the human pancreatic duct cell tumors. *Pancreas* 2000; **21**: 352-357
- 11 Xu ZW, Friess H, Buchler MW, Solioz M. Overexpression of Bax sensitizes human pancreatic cancer cells to apoptosis induced by chemotherapeutic agents. *Cancer Chemother Pharmacol* 2002; **49**: 504-510
- 12 Meggiato T, Calabrese F, De Cesare CM, Baliello E, Valente M, Del Favero G. C-JUN and CPP32 (CASPASE 3) in human pancreatic cancer: relation to cell proliferation and death. *Pancreas* 2003; **26**: 65-70
- 13 Chandler NM, Canete JJ, Callery MP. Caspase-3 drives apoptosis in pancreatic cancer cells after treatment with gemcitabine. *J Gastrointest Surg* 2004; **8**: 1072-1078
- 14 Jones DR, Broad RM, Comeau LD, Parsons SJ, Mayo MW. Inhibition of nuclear factor kappaB chemosensitizes non-small cell lung cancer through cytochrome c release and caspase activation. *J Thorac Cardiovasc Surg* 2002; **123**: 310-317
- 15 Chang GC, Hsu SL, Tsai JR, Wu WJ, Chen CY, Sheu GT. Extracellular signal-regulated kinase activation and Bcl-2 downregulation mediate apoptosis after gemcitabine treatment partly via a p53-independent pathway. *Eur J Pharmacol* 2004; **502**: 169-183
- 16 Habiro A, Tanno S, Koizumi K, Izawa T, Nakano Y, Osanai M, Mizukami Y, Okumura T, Kohgo Y. Involvement of p38 mitogen-activated protein kinase in gemcitabine-induced apoptosis in human pancreatic cancer cells. *Biochem Biophys Res Commun* 2004; **316**: 71-77
- 17 Schniewind B, Christgen M, Kurdow R, Haye S, Kremer B, Kalthoff H, Ungefroren H. Resistance of pancreatic cancer to gemcitabine treatment is dependent on mitochondria-mediated apoptosis. *Int J Cancer* 2004; **109**: 182-188
- 18 Achiwa H, Oguri T, Sato S, Maeda H, Niimi T, Ueda R. Determinants of sensitivity and resistance to gemcitabine: the roles of human equilibrative nucleoside transporter 1 and deoxycytidine kinase in non-small cell lung cancer. *Cancer Sci* 2004; **95**: 753-757
- 19 Xie MJ, Motoo Y, Iovanna JL, Su SB, Ohtsubo K, Matsubara F, Sawabu N. Overexpression of pancreatitis-associated protein (PAP) in human pancreatic ductal adenocarcinoma. *Dig Dis Sci* 2003; **48**: 459-464
- 20 Motoo Y, Satomura Y, Mouri I, Mouri H, Ohtsubo K, Sakai J, Fujii T, Taga H, Yamaguchi Y, Watanabe H, Okai T, Sawabu N. Serum levels of pancreatitis-associated protein in digestive diseases with special reference to gastrointestinal cancers. *Dig Dis Sci* 1999; **44**: 1142-1147
- 21 Motoo Y, Itoh T, Su SB, Nakatani MT, Watanabe H, Okai T, Sawabu N. Expression of pancreatitis-associated protein (PAP) mRNA in gastrointestinal cancers. *Int J Pancreatol* 1998; **23**: 11-16
- 22 Motoo Y, Taga K, Su SB, Xie MJ, Sawabu N. Arginine induces

- apoptosis and gene expression of pancreatitis-associated protein (PAP) in rat pancreatic acinar AR4-2J cells. *Pancreas* 2000; **20**: 61-66
- 23 **Tomasini R**, Samir AA, Vaccaro MI, Pebusque MJ, Dagorn JC, Iovanna JL, Dusetti NJ. Molecular and functional characterization of the stress-induced protein (SIP) gene and its two transcripts generated by alternative splicing. SIP induced by stress and promotes cell death. *J Biol Chem* 2001; **276**: 44185-44192
- 24 **Okamura S**, Arakawa H, Tanaka T, Nakanishi H, Ng CC, Taya Y, Monden M, Nakamura Y. p53DINP1, a p53-inducible gene, regulates p53-dependent apoptosis. *Mol Cell* 2001; **8**: 85-94
- 25 **Tomasini R**, Samir AA, Carrier A, Isnardon D, Cecchinelli B, Soddu S, Malissen B, Dagorn JC, Iovanna JL, Dusetti NJ. TP53INP1s and homeodomain-interacting protein kinase-2 (HIPK2) are partners in regulating p53 activity. *J Biol Chem* 2003; **278**: 37722-37729
- 26 **Tomasini R**, Samir AA, Pebusque MJ, Calvo EL, Totaro S, Dagorn JC, Dusetti NJ, Iovanna JL. P53-dependent expression of the stress-induced protein (SIP). *Eur J Cell Biol* 2002; **81**: 294-301
- 27 **Igney FH**, Krammer PH. Death and anti-death: tumour resistance to apoptosis. *Nat Rev Cancer* 2002; **2**: 277-288
- 28 **Beurel E**, Kornprobst M, Blivet-Van Eggelpoel MJ, Ruiz-Ruiz C, Cadoret A, Capeau J, Desbois-Mouthon C. GSK-3beta inhibition by lithium confers resistance to chemotherapy-induced apoptosis through the repression of CD95 (Fas/APO-1) expression. *Exp Cell Res* 2004; **300**: 354-364
- 29 **Malka D**, Vasseur S, Bodeker H, Ortiz EM, Dusetti NJ, Verrando P, Dagorn JC, Iovanna JL. Tumor necrosis factor alpha triggers antiapoptotic mechanisms in rat pancreatic cells through pancreatitis-associated protein I activation. *Gastroenterology* 2000; **119**: 816-828
- 30 **Bandala E**, Espinosa M, Maldonado V, Melendez-Zajgla J. Inhibitor of apoptosis-1 (IAP-1) expression and apoptosis in non-small-cell lung cancer cells exposed to gemcitabine. *Biochem Pharmacol* 2001; **62**: 13-19
- 31 **Arlt A**, Gehrz A, Muerkoster S, Vorndamm J, Kruse ML, Folsch UR, Schafer H. Role of NF-kappaB and Akt/PI3K in the resistance of pancreatic carcinoma cell lines against gemcitabine-induced cell death. *Oncogene* 2003; **22**: 3243-3251

S- Editor Pan BR L- Editor Zhang JZ E- Editor Bai SH