

Epileptic seizure in neuron-specific AP-3B knockout mice

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There are two isoforms of the AP-3 complex, the ubiquitous AP-3A, and the neuron-specific AP-3B. AP-3A consists of δ , $\beta 3A$, $\mu 3A$ and $\sigma 3$ subunits. AP-3B shares δ and $\sigma 3$ subunits with AP-3A, while the other two subunits of AP-3B, $\beta 3B$ and $\mu 3B$, are neuron-specific. In human, mutations in the $\beta 3A$ subunit of AP-3A were identified in patients suffering from the Hermansky-Pudlak syndrome (HPS), in which function and/or biogenesis of lysosomes and lysosome-related organelles such as melanosomes and platelet dense granules were impaired. *Pearl* mice, one of the HPS model mice, also bear a mutation in the $\beta 3A$ gene and share the same phenotypes with HPS. Another HPS model, *mocha* mice, has mutations in the δ subunit common to AP-3A and AP-3B. In addition to the phenotypes shared with *pearl* mice and HPS patients, *mocha* mice have been reported to suffer from neurological disorders, such as abnormal EEG and inner ear disorders (deafness and balance problems).

To elucidate that these neurological disorders of *mocha* mice result from the impairment in AP-3B, we established $\mu 3B^{-/-}$ mice. The inner ear disorder was not observed in our $\mu 3B^{-/-}$ mice, suggesting that the phenotype is only observed when both AP-3A and AP-3B are deficient in *mocha* mice. However, $\mu 3B^{-/-}$ mice exhibited epileptic seizures (figure). Kindling stimulation and a GABA_A receptor antagonist corroborated the seizure susceptibility of $\mu 3B^{-/-}$ mice. Morphologically, electron microscopy revealed that synaptic vesicles were not homogeneous in size in $\mu 3B^{-/-}$ mice when compared to those in wild-type mice. We also employed an *in vitro* synaptic vesicle formation assay. Brain lysates of wild-type mice, but not of $\mu 3B^{-/-}$ mice, promoted the formation of synaptic-like vesicle formation from isolated endosomal membranes of NGF-treated PC12 pheochromocytoma cells, suggested that AP-3B is required for the *de novo* formation of synaptic vesicles from endosomal membranes. These observations led us to measure the neurotransmitter release. Release of an excitatory glutamate and an inhibitory GABA from the hippocampus was measured in microdialysis experiments. The amount of basal release of the two neurotransmitters was similar between $\mu 3B^{-/-}$ mice and wild-type mice. Of note, however, was that the K⁺-evoked release of GABA was significantly impaired, while that of glutamate was slightly increased, in $\mu 3B^{-/-}$ mice. This difference could not be attributed to the changes in the content of the neurotransmitters themselves in the hippocampus. Taken together, these results suggest that epileptic seizures observed in $\mu 3B^{-/-}$ mice are likely due to impairment in the GABAergic inhibitory neurons. $\mu 3B^{-/-}$ mice may serve as a novel animal model for epilepsy, one of the most common neurological disorders.

