

Increased prostaglandin e2 has a positive correlation with plasma calcium during goldfish reproduction

メタデータ	言語: eng 出版者: 公開日: 2017-10-05 キーワード (Ja): キーワード (En): 作成者: メールアドレス: 所属:
URL	http://hdl.handle.net/2297/37548

Increased Prostaglandin E₂ Has a Positive Correlation with Plasma Calcium during Goldfish Reproduction

Nobuo SUZUKI^{1*}, Yusuke MARUYAMA², Masaki NAKANO³, Atsuhiko HATTORI², Masato HONDA³, Yohei SHIMASAKI³, Toshio SEKIGUCHI¹, Hiroyuki MISHIMA⁴, Shigehito WADA⁵, Ajai K. SRIVASTAV⁶, Kazuichi HAYAKAWA⁷ and Yuji OSHIMA³

Department of Bioscience and Biotechnology, Graduate School of Bioresource and Bioenvironmental Sciences, Kyushu University, Hakozaki 6–10–1, Higashi-ku, Fukuoka, 812–8581, Japan

(Received October 24, 2013 and accepted November 11, 2013)

We recently demonstrated that prostaglandin E₂ (PGE₂) increases osteoclastic activity and induces bone resorption in both *in vitro* and *in vivo* experiments using goldfish. In the fish reproductive period, the plasma calcium (Ca) level in female teleosts increases remarkably to make vitellogenin, which is a major component of egg protein and a Ca-binding protein. In this period, however, there is no reported relationship between PGE₂ and Ca metabolism in fish. To clarify the Ca metabolism in fish reproduction, we examined plasma PGE₂ and Ca levels and measured tartrate-resistant acid phosphatase (TRAP) activities as an indicator of osteoclastic activity in goldfish. Plasma PGE₂ levels in the reproductive stage significantly increased as compared with those in non-reproductive stages. Also, both plasma Ca and TRAP increased in the reproductive stage. Significant positive correlations were recognized between plasma Ca and the gonad somatic index ($r=0.81$, $p<0.001$), plasma Ca and plasma PGE₂ levels ($r=0.635$, $p<0.05$), and plasma Ca and plasma TRAP activities ($r=0.584$, $p<0.05$) from the analysis using samples of both reproductive and non-reproductive stages. Taking these data into consideration, we suggested that PGE₂ acts on osteoclasts and increases plasma Ca as a result of osteoclastic bone resorption, and we concluded that PGE₂ is an important hormone in Ca metabolism during fish reproduction.

Key words: goldfish, plasma Ca, Prostaglandin E₂, reproduction, TRAP

INTRODUCTION

Prostaglandin E₂ (PGE₂) functions to bone metabolism and is an important hormone in bone and promoter of osteoclastogenesis (Kaji *et al.*, 1996; Gardner, 2007; Kaneko *et al.*, 2007). The bone-resorbing activity of mature osteoclasts in osteoblast-containing mouse bone cell cultures was increased by PGE₂, although it did not affect osteoclast-like cell formation in osteoblast-free mouse spleen cell cultures (Kaji *et al.*, 1996). Therefore, we focused on fish scales that coexist with both osteo-

clasts and osteoblasts (Bereiter-Hahn and Zylberberg, 1993; Suzuki *et al.*, 2000; Yoshikubo *et al.*, 2005; Suzuki *et al.*, 2007; Suzuki *et al.*, 2008; Suzuki *et al.*, 2011; Yano *et al.*, 2013). Using the goldfish scale *in vitro* assay system, we recently demonstrated that PGE₂ acts on osteoblasts and then increases the osteoclastic activity in the scales of goldfish as it does in the bone of mammals (Omori *et al.*, 2012). In addition, the intraperitoneal injection of PGE₂ into goldfish induced hypercalcemia (Omori *et al.*, 2012).

In the reproductive period, the plasma calcium (Ca) level in female teleosts increases remarkably (Watts *et al.*, 1975; Yamauchi *et al.*, 1978; Norberg *et al.*, 1989; Suzuki *et al.*, 2004). This Ca is bound to vitellogenin, which is a major component of egg protein and the calcium-binding protein (Tinsley, 1985; Kwon *et al.*, 1993). In this period, PGE₂ synthesized in the ovaries functions to cause ovulation in fish (for a review, see Takahashi *et al.*, 2013). As van Anholt *et al.* (2003) reported that PGE₂ in the blood may serve some physiological roles in fish, PGE₂ secreted from the ovaries might influence plasma Ca in fish. However, there has been no reported relationship between PGE₂ and Ca metabolism during the fish reproductive period.

To clarify the Ca metabolism in fish reproduction, we examined plasma PGE₂ and Ca levels and measured tartrate-resistant acid phosphatase (TRAP) activities as an indicator of osteoclastic activity in goldfish.

We concluded that PGE₂ is an important hormone in Ca metabolism during fish reproduction.

¹ Noto Marine Laboratory, Institute of Nature and Environmental Technology, Kanazawa University, Housu-gun, Ishikawa 927-0553, Japan

² Department of Biology, College of Liberal Arts and Sciences, Tokyo Medical and Dental University, Ichikawa, Chiba 272-0827, Japan

³ Department of Bioscience and Biotechnology, Graduate School of Bioresource and Bioenvironmental Sciences, Kyushu University, Hakozaki 6–10–1, Higashi-ku, Fukuoka, 812–8581, Japan

⁴ Department of Medical Hygiene, Kochi Gakuen College, Kochi 780–0955, Japan

⁵ Department of Oral and Maxillofacial Surgery, Faculty of Medicine, University of Toyama, Sugitani, Toyama 930–0194, Japan

⁶ Department of Zoology, D.D.U. Gorakhpur University, Gorakhpur 273009, India

⁷ Faculty of Pharmaceutical Sciences, Institute of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa, Ishikawa 920–1192, Japan

* Corresponding author (E-mail: nobuos@staff.kanazawa-u.ac.jp)

MATERIALS AND METHODS

Animals

Female goldfish ($n=14$, 49.16 ± 3.77 g) were purchased from a commercial source (Higashikawa Fish Farm, Yamatokoriyama, Japan) and used in the present study. All experimental procedures were conducted in accordance with the Guide for the Care and Use of Laboratory Animals of Kanazawa University.

Measurement of plasma PGE₂, Ca, and TRAP levels of female goldfish in reproductive and non-reproductive stages

Goldfish in both the reproductive (March) ($n=8$) and non-reproductive (August) ($n=6$) stages were anesthetized with ethyl 3-aminobenzoate methanesulfonic acid salt (Sigma-Aldrich Inc., MO, USA). After weighing, the gonad somatic index (GSI) (%) was calculated. A blood sample was then collected from the dorsal aorta using a heparinized syringe. After centrifugation at 15,000 rpm for 3 min, the plasma was immediately frozen and kept at -80°C until use. The plasma total Ca (mg/100 ml) and PGE₂ (pg/ml) levels were determined using specific assay kits (Ca: Calcium E test; PGE₂: PGE₂-ELISA kit, Wako Pure Chemical Industries, Ltd., Osaka, Japan).

The plasma TRAP level was measured using $2\ \mu\text{l}$ of plasma from each goldfish. TRAP activities were measured using an acid tartrate buffer (a 20 mM tartrate in a 0.1 M sodium acetate buffer (pH 5.3)). An aliquot of $100\ \mu\text{l}$ of 20 mM para-nitrophenyl phosphate in an acid tartrate buffer was added to each well in a 96-well microplate. This plate was then incubated at 20°C for 30 min while being shaken. After incubation, the reaction was stopped by adding $50\ \mu\text{l}$ of a 3 N NaOH-20 mM EDTA solution, and the absorbance was then measured at 405 nm. The absorbance was converted into the amount of produced para-nitrophenol (pNP) using a standard curve for pNP.

Statistical analysis

All results are expressed as the means \pm SE. Statistical significance was assessed by Student's *t*-test. Simple correlation coefficients were calculated to assess the relationship among GSI values, plasma PGE₂ levels, plasma Ca levels, and plasma TRAP activities. The statistical significance of the correlation was evaluated using the method of Snedecor and Cochran (1980). In all cases, the selected significance level was $p < 0.05$.

RESULTS

Changes in GSI, plasma PGE₂ levels, Ca levels, and TRAP activities of female goldfish in reproductive and non-reproductive stages

There was a significant difference in the values of GSI between goldfish in March and August (Fig. 1). In addition, the plasma PGE₂ levels, Ca levels, and TRAP activities of female goldfish in March were significantly higher than those in August (Figs. 2, 3, and 4).

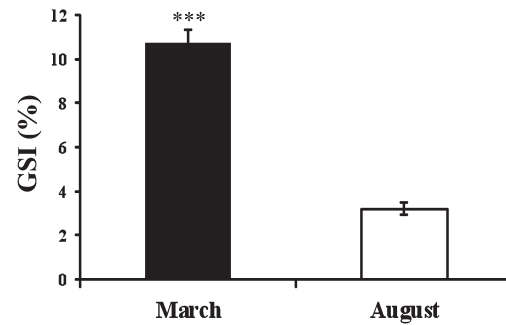


Fig. 1. GSI values of female goldfish in the reproductive (March) and non-reproductive (August) stages. *** indicates a statistically significant difference at $p < 0.001$ in the values of the reproductive and non-reproductive stages.

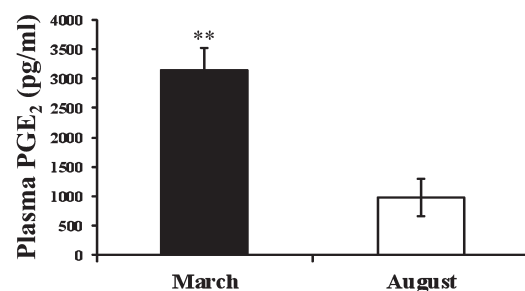


Fig. 2. Plasma PGE₂ values of female goldfish in the reproductive (March) and non-reproductive (August) stages. ** indicates a statistically significant difference at $p < 0.01$ in the values of the reproductive and non-reproductive stages.

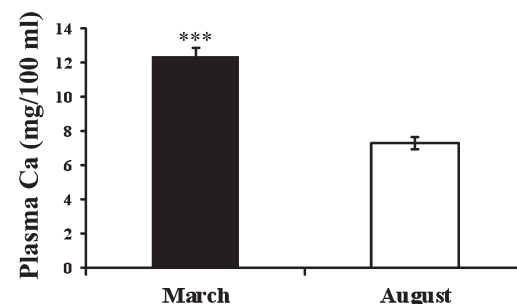


Fig. 3. Plasma Ca values of female goldfish in the reproductive (March) and non-reproductive (August) stages. *** indicates a statistically significant difference at $p < 0.001$ in the values of the reproductive and non-reproductive stages.

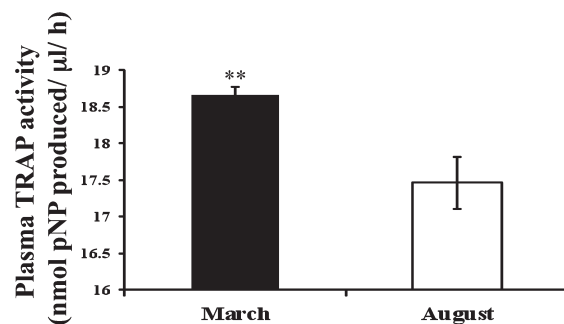


Fig. 4. Plasma TRAP activity (nmol pNP produced/ $\mu\text{l/h}$) values of female goldfish in the reproductive (March) and non-reproductive (August) stages. ** indicates a statistically significant difference at $p < 0.01$ in the values of the reproductive and non-reproductive stages.

Correlation among GSI, plasma PGE₂ levels, Ca levels, and TRAP activities

The results of correlation among GSI, plasma PGE₂ levels, Ca levels, and TRAP activities are indicated in Table 1. Significant positive correlations were recognized

Table 1. Correlation among GSI, plasma Ca levels, PGE₂ levels, and TRAP activities (n=14).

	<i>r</i> values	<i>p</i> values
GSI vs Plasma PGE ₂	<i>r</i> =0.790	<i>p</i> =0.0007
GSI vs Plasma Ca	<i>r</i> =0.813	<i>p</i> =0.0004
GSI vs Plasma TRAP	<i>r</i> =0.631	<i>p</i> =0.015
Plasma Ca vs Plasma PGE ₂	<i>r</i> =0.635	<i>p</i> =0.014
Plasma Ca vs Plasma TRAP	<i>r</i> =0.584	<i>p</i> =0.028
Plasma PGE ₂ vs Plasma TRAP	<i>r</i> =0.514	<i>p</i> =0.058

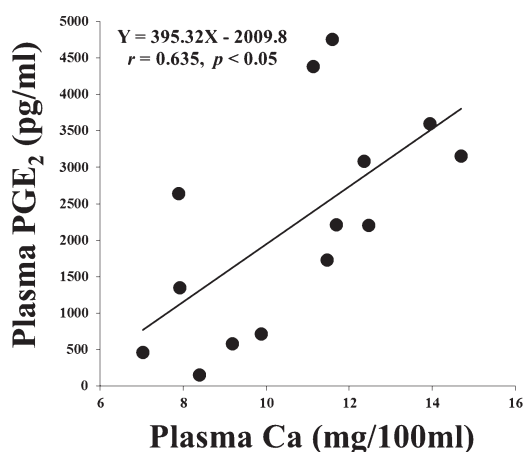


Fig. 5. Relationship between PGE₂ (pg/ml) and Ca (mg/100 ml) in the plasma of goldfish in the reproductive (March) and non-reproductive (August) stages.

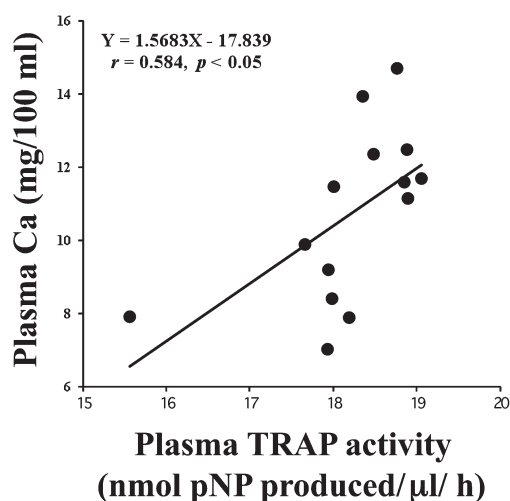


Fig. 6. Relationship between Ca (mg/100 ml) and TRAP activities (nmol pNP produced/μl/h) in the plasma of goldfish in the reproductive (March) and non-reproductive (August) stages.

between GSI and plasma PGE₂ (*r*=0.790, *p*<0.001), GSI and plasma Ca (*r*=0.813, *p*<0.001), and GSI and plasma TRAP (*r*=0.631, *p*<0.05) from the analysis using samples of both reproductive and non-reproductive stages.

As a result of having paid attention to the relations with plasma Ca, we discovered a significant positive relationship between plasma Ca and PGE₂ (*r*=0.635, *p*<0.05) (Fig. 5), and between plasma Ca and TRAP (*r*=0.584, *p*<0.05) (Fig. 6).

As PGE₂ levels increased, plasma TRAP activities tended to rise (*r*=0.514, *p*=0.058).

DISCUSSION

The present study is the first to demonstrate that PGE₂ is related to Ca metabolism in fish reproduction. Corresponding to increased GSI, plasma PGE₂ levels, Ca levels, and TRAP activities rose. In addition, significant correlations between plasma Ca and PGE₂ and between Ca and TRAP were observed. Because TRAP is known as an osteoclast-specific marker (for a review, see Vaes, 1988), the increased PGE₂ in the March fish activated osteoclasts and promoted osteoclastic bone resorption. As described in the introduction, we recently demonstrated that PGE₂ acts on osteoblasts and increases the osteoclastic activity in the scales of goldfish as it does in the bone of mammals (Omori *et al.*, 2012). In an *in vivo* experiment, furthermore, hypercalcemia was induced as a result of osteoclastic bone resorption after an intraperitoneal injection of PGE₂ into goldfish (Omori *et al.*, 2012). Taking these results into consideration together with the present study, we have concluded that PGE₂ acts as a calcemic hormone in fish reproduction.

In the present study, the highest correlation between GSI and plasma Ca was recognized. We think that several hormones, with the exception of PGE₂, are involved in Ca metabolism during fish reproduction. The candidate for this hypercalcemic hormone is estrogen. In female teleosts, estrogen enhances the synthesis of vitellogenin, which is a major component of egg protein and a Ca-binding protein (Tinsley, 1985; Kwon *et al.*, 1993). At the same time, estrogen promotes Ca resorption from the scales by activating osteoclasts (Persson *et al.*, 1995; Suzuki *et al.*, 2000; Suzuki and Hattori, 2003; Suzuki *et al.*, 2009). Consequently, plasma vitellogenin and Ca levels increase corresponding to the increase in estrogen level (Norberg *et al.*, 1989). PGE₂ is closely related to ovulation (late stage of fish reproduction) (for a review, see Takahashi *et al.*, 2013), suggesting that in the early stage of fish reproduction, estrogen acts as a hypercalcemic hormone, and then PGE₂ plays roles in both ovulation and Ca metabolism.

On the other hand, we previously demonstrated that a hypocalcemic hormone, calcitonin, acts on scales and inhibits osteoclastic activity using an *in vitro* scale assay system with goldfish (Suzuki *et al.*, 2000). As estrogen activates osteoclasts in some teleosts in both *in vivo* and *in vitro* experiments (Persson *et al.*, 1995; Suzuki *et al.*, 2000; Suzuki and Hattori, 2002; Suzuki and Hattori, 2003; Suzuki *et al.*, 2009), a counteraction may

exist between calcitonin and estrogen in osteoclasts of the scale. Using the *in vitro* scale assay system, the increased osteoclastic activity with estrogen was actually suppressed by calcitonin in goldfish (Suzuki *et al.*, 2000). Furthermore, our previous study demonstrated the interaction between calcitonin and estrogen. In the ultimobranchial gland, which is the secretion organ of calcitonin, estrogen receptors were detected by estrogen-specific binding assay and immunohistochemical analysis in goldfish (Suzuki *et al.*, 2004). Also, three types of estrogen receptors were detected in the ultimobranchial gland of goldfish (Suzuki *et al.*, 2004). Moreover, just after injecting estrogen into goldfish, plasma calcitonin level increased before the rise of plasma Ca (Suzuki *et al.*, 2004). Considering from our present data, we strongly suggested that PGE₂ affects other calcemic hormones in fish reproduction. Thus, in the future, we will examine the interaction among calcemic hormones, such as PGE₂, calcitonin, and estrogen, and elucidate the mechanism of teleost bone metabolism during the reproductive stages.

ACKNOWLEDGMENTS

This study was supported in part by grants to N.S. (Grant-in-Aid for Scientific Research [C] No. 24620004 by JSPS), to A.H. (Grant-in-Aid for Scientific Research [C] No. 24570068 by JSPS), to T. S. (Grant-in-Aid for Young Scientists [B] No. 40378568 by JSPS), to H.M. (Grant-in-Aid for Scientific Research [C] No. 23592727 by JSPS), and to K.H. (Grant-in-Aid for Scientific Research for Exploratory Research No.24651044 by JSPS).

REFERENCES

- Bereiter-Hahn, J. and L. Zylberberg 1993 Regeneration of teleost fish scale. *Comp. Biochem. Physiol.*, **105A**: 625–641
- Gardner, C. R. 2007 Comparison of morphological effects of PGE₂ and TGFβ on osteoclastogenesis induced by RANKL in mouse bone marrow cell cultures. *Cell Tissue Res.*, **330**: 111–121
- Kaji, H., T. Sugimoto, M. Kanatani, M. Fukase, M. Kumegawa and K. Chihara 1996 Prostaglandin E₂ stimulates osteoclast-like cell formation and bone-resorbing activity via osteoblasts: Role of cAMP-dependent protein kinase. *J. Bone Mineral Res.*, **11**: 62–71
- Kaneko, H., M. Mehrotra, C. Alander, U. Lerner, C. Pilbeam and L. Raisz 2007 Effects of prostaglandin E₂ and lipopolysaccharide on osteoclastogenesis in RAW 264.7 cells. *Prostaglandins Leukot. Essent. Fatty Acids*, **77**: 181–186
- Kwon, H.C., S. Hayashi and Y. Mugiya 1993 Vitellogenin induction by estradiol-17β in primary hepatocyte culture in the rainbow trout, *Oncorhynchus mykiss*. *Comp. Biochem. Physiol.*, **104B**: 381–386
- Norberg B, B. Th. Björnsson, C. L. Brown, U.-P. Wichardt, L. J. Deftos and C. Haux 1989 Changes in plasma vitellogenin, sex steroids, calcitonin, and thyroid hormones related to sexual maturation in female brown trout (*Salmo trutta*). *Gen. Comp. Endocrinol.*, **75**: 316–326
- Omori, K., S. Wada, Y. Maruyama, A. Hattori, K. Kitamura, Y. Sato, M. Nara, H. Funahashi, K. Yachiguchi, K. Hayakawa, M. Endo, R. Kusakari, S. Yano, A.K. Srivastav, T. Kusui, S. Ejiri, W. Chen, Y. Tabuchi, Y. Furusawa, T. Kondo, Y. Sasayama, T. Nishiuchi, M. Nakano, T. Sakamoto and N. Suzuki 2012 Prostaglandin E₂ increases both osteoblastic and osteoclastic activity in the scales and participates in calcium metabolism in goldfish. *Zool. Sci.*, **29**: 499–504
- Persson, P., Y. Takagi and B. T. Björnsson 1995 Tartrate resistant acid phosphatase as a marker for scale resorption in rainbow trout, *Oncorhynchus mykiss*: Effects of estradiol-17β treatment and refeeding. *Fish Physiol. Biochem.*, **14**: 329–339
- Snedecor, G. W. and W. G. Cochran 1980 *Statistical Methods*, seventh ed. Iowa State Univ. Press, Ames (USA)
- Suzuki, N., J. A. Danks, Y. Maruyama, M. Ikegame, Y. Sasayama, A. Hattori, M. Nakamura, M.J. Tabata, T. Yamamoto, R. Furuya, K. Saijoh, H. Mishima, A. K. Srivastav, Y. Furusawa, T. Kondo, Y. Tabuchi, I. Takasaki, V.S. Chowdhury, K. Hayakawa and T.J. Martin 2011 Parathyroid hormone 1 (1–34) acts on the scales and involves calcium metabolism in goldfish. *Bone*, **48**: 1186–1193
- Suzuki, N. and A. Hattori 2002 Melatonin suppresses osteoclastic and osteoblastic activities in the scales of goldfish. *J. Pineal Res.*, **33**: 253–258
- Suzuki, N. and A. Hattori 2003 Bisphenol A suppresses osteoclastic and osteoblastic activities in the cultured scales of goldfish. *Life Sci.*, **73**: 2237–2247
- Suzuki, N., K. Hayakawa, T. Kameda, A. Toriba, N. Tang, M.J. Tabata, K. Takada, S. Wada, K. Omori, A.K. Srivastav, H. Mishima and A. Hattori 2009 Monohydroxylated polycyclic aromatic hydrocarbons inhibit both osteoclastic and osteoblastic activities in teleost scales. *Life Sci.*, **84**: 482–488
- Suzuki, N., K. Kitamura, T. Nemoto, N. Shimizu, S. Wada, T. Kondo, M. J. Tabata, F. Sodeyama, K. Ijiri and A. Hattori 2007 Effect of vibration on osteoblastic and osteoclastic activities: Analysis of bone metabolism using goldfish scale as a model for bone. *Adv. Space Res.*, **40**: 1711–1721
- Suzuki, N., M. Somei, A. Seki, R.J. Reiter and A. Hattori 2008 Novel bromomelatonin derivatives as potentially effective drugs to treat bone diseases. *J. Pineal Res.*, **45**: 229–234
- Suzuki, N., T. Suzuki and T. Kurokawa 2000 Suppression of osteoclastic activities by calcitonin in the scales of goldfish (freshwater teleost) and nibbler fish (seawater teleost). *Peptides*, **21**: 115–124
- Suzuki, N., K. Yamamoto, Y. Sasayama, T. Suzuki, T. Kurokawa, A. Kambegawa, A. K. Srivastav, S. Hayashi and S. Kikuyama 2004 Possible direct induction by estrogen of calcitonin secretion from ultimobranchial cells in the goldfish. *Gen. Comp. Endocrinol.*, **138**: 121–127
- Takahashi, T., C. Fujimori, A. Hagiwara, and K. Ogiwara 2013 Recent advances in the understanding of teleost medaka ovulation: The roles of proteases and prostaglandins. *Zool. Sci.*, **30**: 239–247
- Tinsley, D. 1985 A comparison of plasma levels of phosphoprotein, total protein and total calcium as indirect indices of exogenous vitellogenesis in the Crucian carp, *Carassius carassius* (L.). *Comp. Biochem. Physiol.*, **80B**: 913–916
- Vaes, G. 1988 Cellular biology and biochemical mechanism of bone resorption. *Clin. Orthop. Relat. Res.*, **231**: 239–271
- van Anholt, R. D., T. Spanings, W. Koven and S. E. Wendelaar Bonga 2003 Effects of acetylsalicylic acid treatment on thyroid hormones, prolactins, and the stress response of tilapia

- (*Oreochromis mossambicus*). *Am. J. Physiol. Regul. Integr. Comp. Physiol.*, **285**: R1098–R1106
- Watts, E. G., D. H. Copp and L. J. Deftos 1975 Changes in plasma calcitonin and calcium during the migration of salmon. *Endocrinology*, **96**: 214–218
- Yamauchi, H., H. Orimo, K. Yamauchi, K. Takano and H. Takahashi 1978 Increased calcitonin levels during ovarian development in the eel, *Anguilla japonica*. *Gen. Comp. Endocrinol.*, **36**: 526–529
- Yano, S., K. Kitamura, Y. Satoh, M. Nakano, A. Hattori, T. Sekiguchi, M. Ikegame, H. Nakashima, K. Omori, K. Hayakawa, A. Chiba, Y. Sasayama, S. Ejiri, Y. Mikuni-Takagaki, H. Mishima, H. Funahashi, T. Sakamoto and N. Suzuki 2013 Static and dynamic hypergravity responses of osteoblasts and osteoclasts in medaka scales. *Zool. Sci.*, **30**: 217–223
- Yoshikubo, H., N. Suzuki, K. Takemura, M. Hoso, S. Yashima, S. Iwamuro, Y. Takagi, M. J. Tabata and A. Hattori 2005 Osteoblastic activity and estrogenic response in the regenerating scale of goldfish, a good model of osteogenesis. *Life Sci.*, **76**: 2699–2709